

CHAPTER III

RESULTS

3.1 Cloning and characterization of AcMRJPs cDNA

The full length of AcMRJP4 and AcMRJP5 cDNA family was prepared by PCR of the first stranded cDNA with specific primers which designed from AmMRJPs cDNA sequence in GenBank Database. A pair of primers for AcMRJP4 cDNA amplification were 5'-ATA TCC TAG AAA AAA AAT GAC AAA ATG GTT GC-3' (MRJP4) and 5'-GGG GTA CCC TTT TTT TTT TTT TTT TTT TTT TTT TA-3' (RMJ2). A pair of primers for AcMRJP5 cDNA amplification were 5'-CTG TCG TTT GCA AAA TAT TTG CAG C-3' (MRJP5_2) and 5'-GGG GTA CCC TTT TTT TTT TTT TTT TTT TTT TTT TA-3' (RMJ2) (Table 2.1 and Figure 2.1). The expected full length of AcMRJP4 and AcMRJP5 cDNA were about 1,600 bp and 1,900 bp as estimated from their corresponding size of AmMRJPs cDNA.

PCR product of 4 different sizes were detected when amplified by primers for AcMRJP4 cDNA (Figure 3.1). The sizes of PCR products were 400, 1,500, 1,600 and 2,000 bp in length. The PCR product size of 1,500, 1,600 and 2,000 bp were recovered from agarose gel by QIAquick gel extraction kit and used for cloning. PCR products were ligated with pGEM[®]-T easy vector and electro-transformed to *E. coli* JM 109 host.

Amplification of first stranded cDNAs with specific primers (MRJP5_2 and RMJ2) of AcMRJP5 cDNA, only one PCR product of 1,900 bp was obtained (Figure 3.2). The PCR product size of 1,900 bp was the expected size as calculated from AmMRJP5 cDNA. This PCR product was then clone as described before.

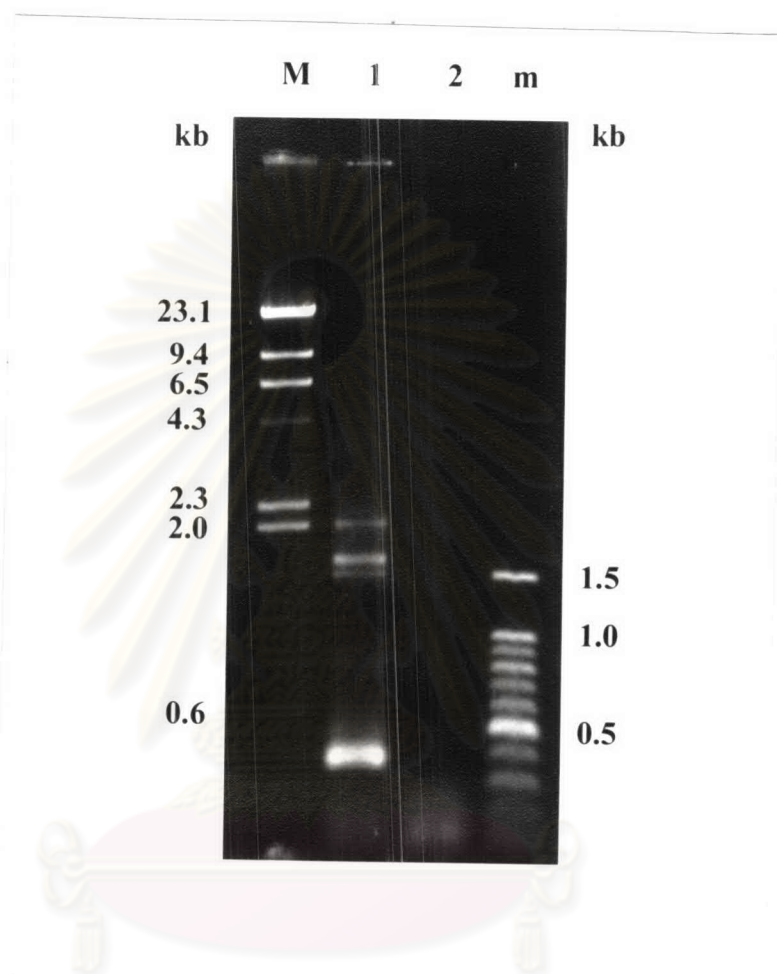


Figure 3.1 PCR amplification for full length cDNA of AcMRJP4

Lane M = λ / *Hind* III standard molecular weight marker

Lane 1 = The amplification products of first strand cDNA

Lane 2 = Negative control

Lane m = A 100 bp DNA ladder

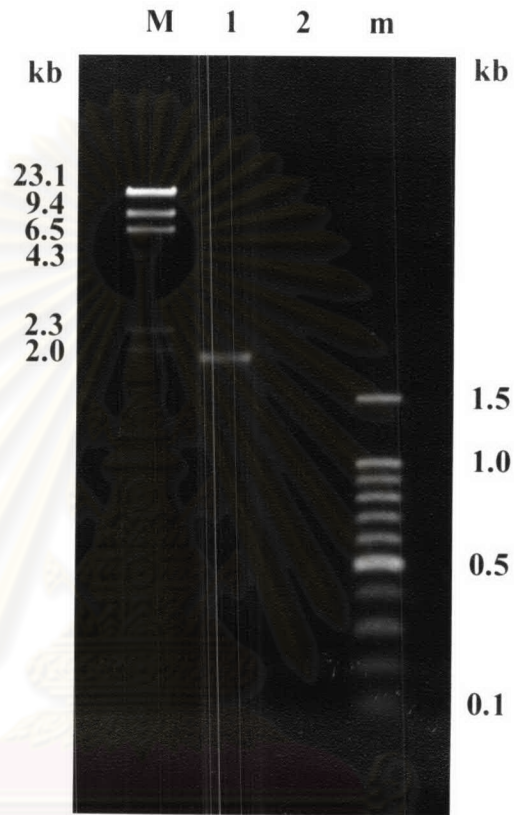


Figure 3.2 PCR amplification for full length cDNA of AcMRJP5

Lane M = λ / *Hind* III standard molecular weight marker

Lane 1 = The amplification products of first strand cDNA

Lane 2 = Negative control

Lane m = A 100 bp DNA ladder

The recombinant clones were screened by blue-white screening on selective plate. The white colonies were randomly picked for culture. The recombinant plasmids were extracted from the cell culture. After plasmid extraction, recombinant plasmids were mapped by digestion with restriction endonucleases. If they contained MRJP cDNA sequence, they should have the restriction map related to restriction map predicted from *A. mellifera* MRJPs cDNA. The restriction enzymes *EcoRI* and *SspI* were selected for characterized the MRJP cDNA sequence. The pGEM[®]-T easy vector multiple cloning region was flanked by recognition sites for the restriction enzyme *EcoRI*. For AmMRJPs sequence, the restriction enzyme *EcoRI* could not cut any AmMRJP cDNA sequence except in AmMRJP6. There is 1 recognition site of *EcoRI* in AmMRJP6 cDNA, after digestion 2 fragments of 324 and 1125 bp will be obtained. Thus, the recombinant plasmids containing other AmMRJP cDNA, restriction enzyme *EcoRI* can be used to determined size of the insert. The pGEM[®]-T easy vector has 2 *SspI* recognition sites and all AmMRJP cDNA sequences also contain *SspI* recognition sites. The position of recognition sites for the restriction enzyme *SspI* are differed in each AmMRJP cDNA sequence. So, it can be used to characterized the family of MRJP cDNA. Restriction fragments size of various recombinant plasmids of pGEM[®]-T AmMRJP3-6 cDNA after digestion with *SspI* are shown in Table 3.1.

Recombinant plasmids containing 1,500, 1,600, 1,900 and 2,000 bp insert DNA were characterized by restriction endonucleases digestion with *EcoRI* and *SspI* (Figure 3.3, 3.4 and 3.5). The recombinant plasmid containing 1,500 bp insert DNA, when digested with *EcoRI* two fragments of 380 and 950 bp was obtained. In addition, the digested product of restriction enzymes *SspI* was 790, 1200 and 2300 bp (Figure 3.3).

Comparison of size of digested product to those of AmMRJP cDNA (Table 3.1). It showed that 1500 bp cDNA insert was most likely be AcMRJP6 cDNA.

Digestion of the recombinant plasmids containing 1,600 bp insert DNA with restriction enzymes *EcoRI*, two DNA fragments size of 1,600 and 3,000 bp was obtained. It showed that *EcoRI* can not cut within the cDNA insert. In addition, the digested product of restriction enzymes *SspI* was 410, 800, 1150 and 2200 bp (Figure 3.4). These digested products sizes were similar to those obtained from AmMRJP4 cDNA (Table 3.1). Therefore, it was mostly that the 1600 bp cDNA insert was AcMRJP4.

The recombinant plasmids containing 1,900 bp insert DNA were digested with restriction enzymes *EcoRI*. The digested products size of 1,900 and 3,000 bp were detected. It showed that restriction enzymes *EcoRI* can not cut within the cDNA insert. In addition, the digested products size of 790, 1850 and 2150 bp were obtained after digested with *SspI* (Figure 3.5). The sizes of digested product were compared with those of AmMRJP cDNA (Table 3.1). The result showed that 1,900 bp cDNA insert might be AcMRJP5.

The recombinant plasmids containing 2,000 bp insert DNA were digested with restriction enzymes *EcoRI*. The digested products size of 2,000 and 3,000 bp were detected. It showed that restriction enzymes *EcoRI* can not cut within the cDNA insert. In addition, the digested products size of 1300, 1400 and 2150 bp were obtained after digested with *SspI*. The sizes of digested product were compared with those of AmMRJP cDNA (Table 3.1). The result showed that 2,000 bp cDNA insert might be AcMRJP3.

These four types of recombinant plasmids were further identified by DNA sequencing using M13 forward and M13 reverse primers. The nucleotide sequences were

compared with the DNA sequences deposited in the GenBank database. The results of nucleotide sequence and restriction pattern of recombinant plasmids showed that 1,500, 1,600, 1,900 and 2,000 bp cDNA insert were AcMRJP6, AcMRJP4, AcMRJP5 and AcMRJP3 respectively.

AcMRJP3

Recombinant plasmid containing 2,000 bp cDNA insert from one clone was sequenced using M13 forward and M13 reverse primer. Nucleotide sequences obtained (Figure 3.6) was almost 100% identical with AcMRJP3 cDNA previously reported by Srisuparbh (2002). The single nucleotide substitutions also called Single Nucleotide Polymorphism (SNP) were found in this nucleotide sequence. These changes led to both silent or non-silent substitution. From deduced amino acid sequence, amino acid residue at position 66, 529, 533 and 536 were change when compared with those of Srisuparbh's AcMRJP3 cDNA (Figure 3.7).

The consensus polyadenylation signal sequences were observed. The sequences AATAAATAAAATAAA contained two separated or three partially overlapping consensus polyadenylation signal sequences (AATAAA) is located 14 bp upstream from the poly(A) tail.

Table 3.1 Restriction map of AmMRJP3-AmMRJP6 cDNA in pGEM[®]-T easy vector digested with restriction enzyme *Ssp*I

Family	Direction of insert DNA	Predicted size of digested DNA fragment (bp)	Reference
MRJP3	+	182, 1159, 1250, 2256	Klaudiny <i>et al.</i> (1994)
MRJP3	-	182, 783, 1159, 2723	Klaudiny <i>et al.</i> (1994)
MRJP4	+	182, 391, 732, 1072, 2228	Klaudiny <i>et al.</i> (1994)
MRJP4	-	182, 391, 755, 1072, 2205	Klaudiny <i>et al.</i> (1994)
MRJP5	+	42, 182, 717, 1875, 2167	Albert <i>et al.</i> (1999a)
MRJP5	-	42, 182, 694, 1875, 2190	Albert <i>et al.</i> (1999a)
MRJP6	+	84, 182, 842, 1128, 2210	Albert <i>et al.</i> (2004)
MRJP6	-	84, 182, 737, 842, 2601	Albert <i>et al.</i> (2004)

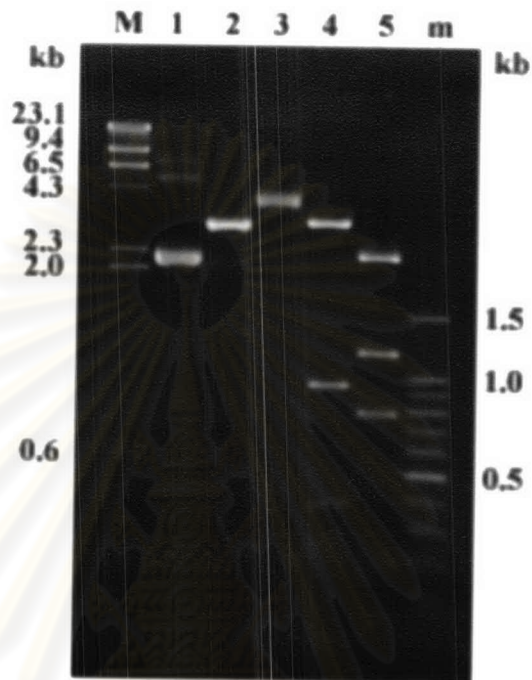


Figure 3.3 Restriction analysis of recombinant plasmid containing 1,500 bp cDNA insert.

Lane M = A λ /*Hind* III standard DNA marker

Lane 1 = Undigested pGEM

Lane 2 = pGEM digested with *EcoR* I

Lane 3 = Undigested recombinant plasmid

Lane 4 = Recombinant plasmid digested with *EcoR* I

Lane 5 = Recombinant plasmid digested with *Ssp* I

Lane m = A 100 bp DNA ladder

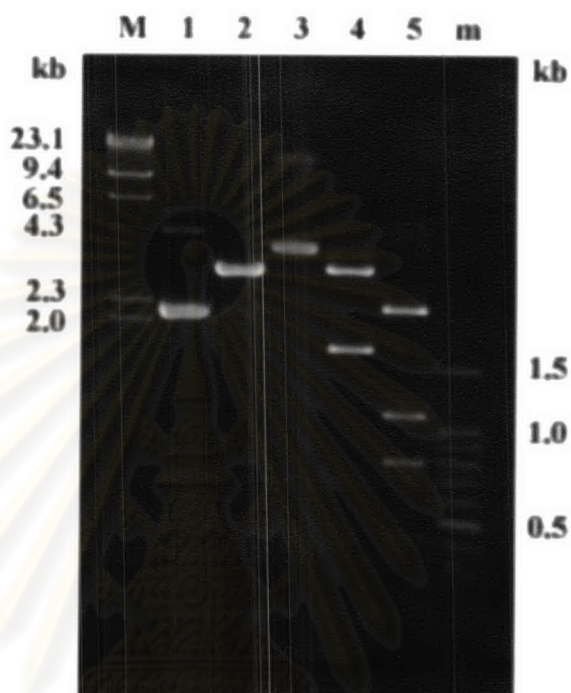


Figure 3.4 Restriction analysis of recombinant plasmid containing 1,600 bp cDNA insert.

Lane M = A λ /*Hind* III standard DNA marker

Lane 1 = Undigested pGEM

Lane 2 = pGEM digested with *EcoR* I

Lane 3 = Undigested recombinant plasmid

Lane 4 = Recombinant plasmid digested with *EcoR* I

Lane 5 = Recombinant plasmid digested with *Ssp* I

Lane m = A 100 bp DNA ladder

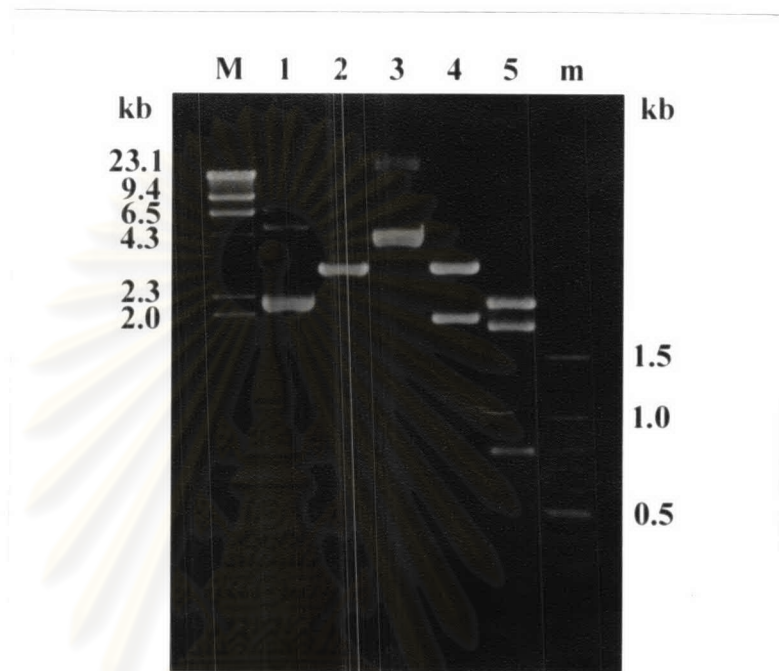


Figure 3.5 Restriction analysis of recombinant plasmid containing 1,900 bp cDNA insert.

Lane M = A λ /*Hind* III standard DNA marker

Lane 1 = Undigested pGEM

Lane 2 = pGEM digested with *EcoR* I

Lane 3 = Undigested recombinant plasmid

Lane 4 = Recombinant plasmid digested with *EcoR* I

Lane 5 = Recombinant plasmid digested with *Ssp* I

Lane m = A 100 bp DNA ladder

CLUSTAL X (1.81) multiple sequence alignment

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AcMRJP3      ATGACAAAGTGGTTGTTGCTGGTGGTGTGTCTTGGTATAGCTTGTCAAGATGTGACAAGC
2000bpRC     ATGACAAAATGGTTGCTGCTGGTGGTGTGTCTTGGTATAGCTTGTCAAGATGTGACAAGC
*****
AcMRJP3      GCAGCTGTGAACCATCAAAGAAAATCTTCAAAAAATTTGGCACATTTCGATGAAGGTGATC
2000bpRC     GCAGCTGTGAACCATCAAAGAAAATCTTCAAAAAATTTGGCACATTTCGATGAAGGTGATC
*****
AcMRJP3      TACGAATGGAACATATTGATTATGATTTTGGTAGCGTTGAAAGAAGAGATGCTGCGATT
2000bpRC     TACGAATGGAACATATTGATTATGATTTTGGTAGCGTTGAAAGAAGAGATGCTGCGATT
*****
AcMRJP3      AAATCTGGCGAATTTGATCACACAAAAAATTACCCTTTCGATGTGGATAGATGGCGTGAT
2000bpRC     AAATCTGGCGAATTTAATCACACAAAAAATTACCCTTTCGATGTGGATAGATGGCGTGAT
*****
AcMRJP3      AAGACATTTGTCACCGTAGAAAGGTTTCGATGGTGTACCTTCTTCTTTGAACGTGGTAACT
2000bpRC     AAGACATTTGTCACCGTAGAAAGGTTTCGATGGTGTACCTTCTTCTTTGAACGTGGTAACT
*****
AcMRJP3      AATAAAAAGGCAAAGGTGGACCTCTTCTACATCCATATCCTGATTGGTCGTGGGCGAAC
2000bpRC     AATAAAAAGGCAAAGGTGGACCTCTTCTACATNNNNNNNNNNNNNNNNNNNNNNNNNNNN
*****
AcMRJP3      TATAAAGATTGCTCTGGAATTGTGAGCGCTTTCAAAATTGCGGTTCGACAAATTCGACAGA
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      TTATGGGTTCTGGACTCAAGTCTTGTCAATAATAATCAACCCATGTGCTCTCCAAAATTG
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      GTAACCTTCGATTTGAATACCTCAAATTGCTTAAGCAAGTCGAGATACCACATAATATT
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      GCCGTAAATGCCACCACGAATGGGGAGAATTAGTATCACTAGCTGTTCAAGCTGTAGAT
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      CCTACGAATACTATGGTGTACATAGCAGACGAAAGAGGTGAAGCTTCAATCATCTATCAA
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      AATTCGACGATTCCTTCCATCGATTGACTTCCAATACTTTTCGATTACGATCCCAGATAT
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
AcMRJP3      ACCAACTGACAGTCGCTGGAGAAAGTTTACAGTGAAAAATGGAATTTGTGGAATTGCA
2000bpRC     NNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN

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(continued)

Figure 3.6 Alignment of the partial nucleotide sequence of recombinant plasmid containing 2,000 bp cDNA insert (2000bpRC) with AcMRJP3 cDNA (Srisuparbh, D., 2002). Conserve residues are indicated by asterisks.

AcMRJP3 2000bpRC CTTAGTCCCGTGACGAACAATCTTTATTACAGTCCTCTCGCTTCTCACAGTTTGTATTAT
NN

AcMRJP3 2000bpRC GTTAACACAGAACAATTCCAGGAATCCACAATATGAAGAAAATAACGTCCAATATGAAGGA
NN

AcMRJP3 2000bpRC TCCCAAGATATTTTGAACACTCAATCATTCGCTAAAGCAGTATCGAAAAATGGCGTCGTT
NN

AcMRJP3 2000bpRC TTCTTGGGACTCGTGAGTAATCAACTGTTGGCTGTGTGAATGAACATCAAGTACTTCAG
NN

AcMRJP3 2000bpRC AAAGAAAATTTTGATGTTGTCGCTCAGAATGAAGAGACACTTCAAATGATCGTTAGTATG
NN

AcMRJP3 2000bpRC AAAATCATGCAAGATCTTCCACAATCCGGCAGAATTAATGATCCAGGAAATGAATATATG
NN

AcMRJP3 2000bpRC TTGGCTTTAAGTAACAAAATGCAAAAAATAATAACAATGATTTTAATTTCAACGACGTA
NN

AcMRJP3 2000bpRC AATTTCCGAATTTTGGGTGCGAATGTAAATCACTTACAAGAAACACTCGTTGCGCAAAA
NN

AcMRJP3 2000bpRC TCTAATAATCAGAATGCTAACAATCAGAATGCTAACAATCAAAATGCTACCAATCAGAAT
NN

AcMRJP3 2000bpRC GATACCAACCAGAATGATAATGGTACCAACAGGAGGAATGGTAACAACCAAAATGGTAAC
NN

AcMRJP3 2000bpRC AGACAAAATGATAATAACAGAATGATAACAAGCAGAATGCTAACAAGCAGAATGCTAAC
NN

AcMRJP3 2000bpRC AAGCAGAATGCTAACAAGCAAAATGATAACAAGCAAAATGATAACAAGCAAAATGGTAAC
NN

AcMRJP3 2000bpRC AGACAAAATGATAATAGGCAGAATGATAACAAGCAAAATGATAATAGGCAGAATGATAAC
NNCAAGCAAAATGATAATAGGCAGAATGATAAC

AcMRJP3 2000bpRC AAGCAAAATGGTAACAGACAAAATGGTAATAGACAGAATGATAACAAGCGGAATGGTAAC
AAGCAAAATGGTAACAGACAAAATGATAATAGACAGAAAGATAACCAGCGGAATGGTAAC

AcMRJP3 2000bpRC AGGCAAAATGATAATAGACAGAATGATAACAAGCGGAATAGTAACAGGCAAAATGATAAT
AGGCAAAATGATAATAGACAGAATGATAACAAGCGGAATAGTAACAGGCAAAATGATAAT

AcMRJP3 2000bpRC AGACAGAATGATAACAAGCGGAATGGAAACAGGCAAAATGATAACAAGCAAAATGATAAC
AGACAGAATGATAACAAGCGGAATGGTAACAGGCAAAATGATAACAAGCAAAATGATAAC

Figure 3.6 (continued)

AcMRJP3
2000bpRC
AAGCAAAATGATAACAGGCAGAATGATAACAATCAGAATGATAATCAGAATGATAATAAT
AAGCAAAATGATAACAGGCAGAATGATAACAATCAGAATGATAATCAGAATGATAATAAT

AcMRJP3
2000bpRC
CGAAATAATCAAGCTCATCATCTTAA-----
CGAAATAATCAAGCTCATCATCTTAAAAATCACATTAATCAATTAATTATCAATTAAA

AcMRJP3
2000bpRC

ATCAATTAATTAGGATGTAAACCAAATTATTTTTTAAAATATTTTTTCGATGTAAACAAA

AcMRJP3
2000bpRC

ATTTTTTTAAATCTTTCATTATATTATAAATAAATAAATAAATATCGTTTTTCGCATAAA

AcMRJP3
2000bpRC

AAAAAAAAAAAAAAAAAAAAAAAAA

Figure 3.6 (continued)



CLUSTAL X (1.81) multiple sequence alignment

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AcMRJP3      MTKWLLLVVCLGIACQDVTSAAVNHQRKSSKNLAHSMKVIYEWKHIDYDFGSVERRDAAI
2000bpRC     MTKWLLLVVCLGIACQDVTSAAVNHQRKSSKNLAHSMKVIYEWKHIDYDFGSVERRDAAI
*****

AcMRJP3      KSGEFDHTKNYPFDVDRWRDKTFVTVERFDGVPSSLNVVTNKKGKGGPLLHPYDWSWAN
2000bpRC     KSGEFDHTKNYPFDVDRWRDKTFVTVERFDGVPSSLNVVTNKKGKGGPLLH-----
*****:*****

AcMRJP3      YKDCSGIVSAFKIAVDKFDRLWLVDSSLVNNNQPMCSPKLVTFDLNTSKLLKQVEIPHNI
2000bpRC     -----

AcMRJP3      AVNATTEWGELVSLAVQAVDPTNTMVYIADERGEASIIYQNSDDSFHRLTSNTFDYDPRY
2000bpRC     -----

AcMRJP3      TKLTVAGESFTVKNGICGIALSPVTNNLYYSPLASHSLYYVNTSEQFRNPQYEENNVQYEG
2000bpRC     -----

AcMRJP3      SQDILNTQSFAKAVSKNGVFLGLVSNSTVGCVNEHQVLQKENFDVVAQNEETLQMIVSM
2000bpRC     -----

AcMRJP3      KIMQDLPOSGRINDPGNEYMLALS NKMQKI INNDFNFNDVNFRI LGANVNHLTRNTRCAK
2000bpRC     -----

AcMRJP3      SNNQNANNQNANNQATNQNDTNQNDNGTNRNRNGNNQNGNRQNDNKQNDNKQANKQANAN
2000bpRC     -----

AcMRJP3      QKQNDNKQNDNKQNDNKQNGNRQNDNRQNDNKQNDNRQNDNKQNGNRQNGNRQNDNKRNNGN
2000bpRC     -----QNDNRQNDNKQNGNRQNDNRQKDNQRNGN
*****.***:*.***

AcMRJP3      RQNDNRQNDNKRNNSNRQNDNRQNDNKRNNGNRQNDNKQNDNKQNDNRQNDNNQNDNQNDNN
2000bpRC     RQNDNRQNDNKRNNSNRQNDNRQNDNKRNNGNRQNDNKQNDNKQNDNRQNDNNQNDNQNDNN
*****

AcMRJP3      RNNQAHHS
2000bpRC     RNNQAHHS
*****

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Figure 3.7 Alignment of the partial deduced amino acid sequence of recombinant plasmid containing 2,000 bp cDNA insert (2000bpRC) with AcMRJP3 (Srisuparbh, D., 2002). Conserve residues are indicated by asterisks.

AcMRJP4

Recombinant plasmids containing 1,600 bp cDNA insert which was expected to be AcMRJP4 cDNA as study by digestion with restriction enzyme *EcoRI* and *SspI*. Further identification of this cDNA insert, the recombinant plasmids containing 1,600 bp cDNA insert were sequenced by four primers (M13 forward and M13 reverse, MRJP4_2 and MRJP4_4) that shown in Table 2.1 and Figure 2.1. Two internal sequencing primers (namely MRJP4_2 and MRJP4_4) were designed from nucleotide sequences obtained using M13 forward and M13 reverse primers. Recombinant plasmids from two clones were sequenced along the entire length. The nucleotide sequences of the insert from these two recombinant plasmids, partial sequence of the insert DNA from another clone and AcMRJP4 cDNA sequence retrieved from EST library of *A. cerana* hypopharyngeal glands (GenBank Acc. CB350335) were assembled (Appendix C). The complete nucleotide sequence was compared with the DNA sequence deposited in GenBank database using nucleotide Blast (BlastN) and translated protein Blast (BlastX). The result showed that similar to AmMRJP4 cDNA. This sequence was most likely to be AcMRJP4 cDNA.

AcMRJP4 cDNA indicated a length of 1608 bp (including poly(A) tail) The sequence contained an open reading frame (nucleotides 1-1458) which encoded 485 amino acid residues (Figure 3.8). The sequence AATAAAATAAA containing two partially overlapping consensus polyadenylation signal sequences (AATAAA) was located 15 bp upstream from the poly(A) tail. The computer sequence analysis predicted that the signal peptidase cleavage site was located between Gly 20 and Ala 21. A comparison of AcMRJP4 and AmMRJP4 nucleotide sequences analysis by blast N

program revealed an identity of 89%. For blast X program protein sequence of AcMRJP4 was showed 79% sequence identity and 85% sequence similarity to AmMRJP4. But program was calculated by disregard partial amino acid sequence of the C-terminal. The deduced amino acid (without putative signal peptide) composition of AcMRJP4 comprised of 36.77% hydrophobic, 35.70% neutral and 27.53% hydrophilic amino acid residues. The essential amino acid content was 42.4%.

The pI value was estimated to be 5.84. The estimated molecular weight was 52.8 kDa. Seven putative *N*-glycosylation sites were found.

The nucleotide sequence and deduced amino acid sequence of AcMRJP4 cDNA were compared with the nucleotide sequence and deduced amino acid sequence of AmMRJP4 cDNA. The result of alignment was shown in Figure 3.9 and Figure 3.10.



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

ATGACAAAATGGTTGCTGTTGATGGCATGCCTTGGCATAGCTTGTCAAATATTAGAGGT 60
 M T K W L L L M A C L G I A C Q N I R G **↑**
GCCGTTGTTTCGAGAAAATTCTCTCGAGAAAAAATTAACAAATACGTTGAACGTGATTAC 120
 A V V R E **N S S R** K K L T N T L N V I H
GAATGGAAGTATGTCGATTATGATTTCCGGTAGCGACGAAAAAAGGCAAGCTGCGATTCAA 180
 E W K Y V D Y D F G S D E K R Q A A I Q
TCTGGCGAATATGATCGTACGAAAAATTATCCTCTTGACGTGATCAATGGCATGATAAG 240
 S G E Y D R T K N Y P L D V D Q W H D K
ACTTTTGTCACTATGTTAAGATACGATGGTGTGCCTTCTCTTTGAACGTGGTATCTGAC 300
 T F V T M L R Y D G V P S S L N V V S D
AAAACCTGGCAACGGTGGACCGCTTCTACAACCTTATCCCGATTGGTCATTTGCTAAGTAT 360
 K T G N G G P L L Q P Y P D W S F A K Y
GAAGATTGCTCTGGAATCGTGAGCGCCAACAAAATTGCTATCGACGAATATGAGAGATTG 420
 E D C S G I V S A N K I A I D E Y E R L
TGGGTTCTGGACTCGGGCCTTGTCAATAATATTCAACCTATGTGTTCTCCAAAATTGCTT 480
 W V L D S G L V N N I Q P M C S P K L L
GCCTTTGATTTGACTACTTCGAAATTGCTCAAGCAAGTCGAGATACCGCACGATGTTGCC 540
 A F D L T T S K L L K Q V E I P H D V A
GTAATGCCACCACAGGAAAGGGCGGATTAGCATCTTTAGCTGTTCAAGCTATGGATTCT 600
 V **N A T T** G K G G L A S L A V Q A M D S
GTAATACTATGGTGTACATGGCAGATAACAAAGATGATGCTTTAATTGTCTACCAAAT 660
 V N T M V Y M A D N K D D A L I V Y Q N
GCCGATGATTCTTTCCATCGATTGTCTTCCCACATTTCCAATCACAACCTTTAGATCTGAC 720
 A D D S F H R L S S H I S N H N F R S D
AAAATGTCGCAAGAAAATCTCACCTTGAAAGAAGTAGACAACAGAGTTTTTGGAAATGGCA 780
 K M S Q E **N L T L** K E V D N R V F G M A
CTTAGTCCGTCGACGATAATCTTTATTATAGTCTCTCTCTTCTCAGAATTTATATTAC 840
 L S S V T H N L Y Y S P L S S Q N L Y Y
GTTAACACAACATCGTTAATGAACTCGCAAAATCAAGGAAATGACGTGCAGTATGAAAGT 900
 V **N T T S** L M N S Q N Q G N D V Q Y E S
GTCCAAGACGTTTTTCAGCAGTCAATTATCCGCTAAAGCAGTATCGAAAAATGGCGTACTC 960
 V Q D V F S S Q L S A K A V S K N G V L
TTTTTCGGATTACGAATAATACTCTTGGTTGCTGGAATGAGCATCAGTCACTTGACAGA 1020
 F F G F T **N N T L** G C W N E H Q S L D R
CAAAATATCGATATTGTAGCTCGAAATGAGACGCTTCAAATGGTCGTTGGTATGAAGATT 1080
 Q N I D I V A R **N E T L** Q M V V G M K I
AAGCAAAACCTTCCACAATCTGGCAAAGTTAATAATACACAAAGAAATGAACATTTGTTG 1140
 K Q N L P Q S G K V **N N T Q** R N E H L L

(continued)

Figure 3.8 Nucleotide and deduced amino acid sequences of AcMRJP4. Initiation and termination of translational codons and putative polyadenylation signal are boldfaced. The signal peptidase cleavage site was indicated by arrow. *N*-linked glycosylation sites are underlined.

GCTTTAACCAACAAAAAGCAGGACGTGCTAAACAACGATCTTAATCTCGAACATGTGAAC 1200
 A L T N K K Q D V L N N D L N L E H V N

TTCCAAATTTTGGATGCTAATGTAAACGACTTGATACGGAATAGTCGTTGCGCAAATTCT 1260
 F Q I L D A N V N D L I R N S R C A N S

GACAATCAGGATAATAATCAACATAATTATAATCATAATCAAGTTCGTCATTCTTCAAAA 1320
 D N Q D N N Q H N Y N H N Q V R H S S K

TCTGACAATCAGAATAACAATCAACATAACAATCAAGCTTATCATTCTTCAAAGTCTGAC 1380
 S D N Q N N N Q H N N Q A Y H S S K S D

AATTGGGATAACAATAACAATCAAGCTCATCATTCTTCAAATTTGATAATCAGAATAAC 1440
 N W D N N N N Q A H H S S K F D N Q N N

AATCAATATAACAAT**TAG**GTTTCATCATTCTTTCATCAAATCATGTTAAATCTGATAATTA 1500
 N Q Y N N *

TCTTTTCTCGATGTAAGTCAAATATTTTAAAAAATTTTATTACATTATAAAA**CGAATAAA** 1560
ATAAATATCGTTTTTCGCATAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA 1608

Figure 3.8 (continued)



CLUSTAL X (1.81) multiple sequence alignment

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AcMRJP4 -----ATGACAAAATGGTTGCTGTTGAT
AmMRJP4 GTCACCTGTAAAATATTTGTAATATCCTAGAAAAAATGACAAAATGGTTGCTGTTGAT
*****

AcMRJP4 GGCATGCCTTGGCATAGCTTGTCAAAATATTAGAGGTGCCGTTGTTTCGAGAAAATTCCTC
AmMRJP4 GGTATGCCTTGGCATAGCTTGTCAAAATATTAGAGGTGCCGTTGTTTCGAGAAAATTCCTC
** *****

AcMRJP4 GAGAAAAAATTAACAAATACGTTGAACGTGATTACGAATGGAAGTATGTCGATTATGA
AmMRJP4 GGGAAAAAATGACAAATACGTTGAACGTGATTACAAATGGAAGTATGTCGATTATGA
* ***** *

AcMRJP4 TTTCCGTAGCGACGAAAAAAGGCAAGCTGCGATTCAATCTGGCGAATATGATCGTACGAA
AmMRJP4 TTTTCGATAACGACGAAAGGAGGCAAGCTGCGATTCAATCTGGCGAATATGATCGTACAAA
***** **

AcMRJP4 AAATTATCCTCTTGACGTGATCAATGGCATGATAAGACTTTTGTCACTATGTTAAGATA
AmMRJP4 AAATTATCCTCTTGACGTGATCAATGGCACAACAAGACTTTTCTCGCTGAATAAGATA
***** *

AcMRJP4 CGATGGTGTGCCTTCTCTTTGAACGTGGTATCTGACAAAATGGCAACGGTGGACCGCT
AmMRJP4 CAATGGTGTGCCTTCTCTTTGAACGTGGTATCTGACAAAATGGCAACGGTGGACGACT
* ***** *

AcMRJP4 TCTACAACCTTATCCCGATTGGTCATTTGCTAAGTATGAAGATTGCTCTGGAATCGTGAG
AmMRJP4 TCTACAACCGTATCCTGATTGGTCATTTGCCAAGTACGAAGATTGCTCTGGAATCGTGAG
*****

AcMRJP4 CGCCAACAAAATGCTATCGACGAATATGAGAGATTGTGGGTTCTGGACTCGGGCCTTGT
AmMRJP4 CGCTCATAAAATGCTATCGACGAATATGAGAGATTGTGGGTTCTGGATTCTGGGTTCTCGT
*** *

AcMRJP4 CAATAATATCAACCTATGTGTTCTCCAAAATGCTTGCCTTTGATTGACTACTTCGAA
AmMRJP4 CAATAATACGCAACCCATGTGTTCTCCAAAATGTTGCTTTGATCTTAATACCTCGCA
*****

AcMRJP4 ATTGCTCAAGCAAGTCGAGATACCGCACGATGTTGCCGTAAATGCCACCACAGGAAAGGG
AmMRJP4 ATTGCTCAAGCAAGTCGAGATACCGCACGATGTTGCC-----ACCACAGGAAAGGG
*****

AcMRJP4 CGGATTAGCATCTTTAGCTGTTCAAGCTATGGATTCTGTAAATACTATGGTGTACATGGC
AmMRJP4 CGAATTAGTATCTTTAACTGTTCAAGCTATGGATTCTGACAAATACTATGGTGTACATGGT
** *****

AcMRJP4 AGATAACAAAGATGATGCTTTAATTGTCTACAAAATGCCGATGATTCTTTCCATCGATT
AmMRJP4 AGACAACAAAATA--CTTTGATCATCTACAAAATGCCGATGATTCTTTTCATCGATT
*** *****

AcMRJP4 GTCTTCCACATTTCCAATCACAACCTTTAGATCTGACAAAATGTCG-CAAGAAAATCTCA
AmMRJP4 GTCTTCCCACTTTGAATCACAACCTCT-GACAAAATGTCAGATCAACAAGAAAATCTCA
***** **

```

(continued)

Figure 3.9 Alignment of the nucleotide sequence of AcMRJP4 cDNA with AmMRJP4 cDNA published sequence (GenBank Acc. Z26319). Conserve residues are indicated by asterisks.

AcMRJP4 CCTTGAAAGAAGTAGACAACAGAGTTTTTGGAAATGGCACTTAGTTCCGTGACGCATAATC
AmMRJP4 CCTTGAAAGAAGTAGACAACAAAGTTTATGGAATGGCACTTAGTCCCCTGACGCATAATC

AcMRJP4 TTTATTATAGTCTCTCTCTCTCAGAATTTATATTACGTTAACACAACATCGTTAATGA
AmMRJP4 TTTATTACAATTCTCCGTCTTCTGAGAATTTGTATTATGTTAACACAGAATCGTTAATGA
***** * * * * *

AcMRJP4 ACTCGCAAATCAAGGAAATGACGTGCAGTATGAAAGTGTCCAAGACGTTTTTCAGCAGTC
AmMRJP4 AATCGGAAATCAAGGAAATGACGTGCAATATGAAAGAGTCCAAGACGTTTTTCGACAGTC
* * * * *

AcMRJP4 AATTATCCGCTAAAGCAGTATCGAAAAATGGCGTACTCTTTTTCGGATTACGAATAATA
AmMRJP4 AATTAACCGTTAAAGCAGTATCGAAAAATGGCGTACTCTTTTTCGGACTCGGAATAATA
***** * * * * *

AcMRJP4 CTCTGGTTGCTGGAATGAGCATCAGTCACTTGACAGACAAAATATCGATATGTAGCTC
AmMRJP4 CTCTTAGTTGCTGGAACGAGCATCAGTCACTTGACAGACAAAATATCGATGTCGTAGCTC
***** * * * * *

AcMRJP4 GAAATGAG--ACGCTTCAAATGGTCGTTGGTATGAAGATTAAGCAAACCTTCCACAAT
AmMRJP4 GAAATGAGGACACGCTTCAAATGGTCGTTAGTATGAAGATTAAGCAAACGTTCCACAAT
***** * * * * *

AcMRJP4 CTGGCAAAGTTAATAATACACAAAGAAATGAACATTTGTTGGCTTTAACCACAAAAGC
AmMRJP4 CTGGCAGAGTTAATAATACGCAAAGAAATGAATATTTGTTGGCTTTAAGCGACAGAAACC
***** * * * * *

AcMRJP4 AGGACGTGCTAAACAACGATCTTAATCTCGAACATGTGAACTTCCAAATTTGGATGCTA
AmMRJP4 AGAACGTGCTAAACAACGATCTTAATCTCGAACACGTGAACTTCCAAATTTGGGCGCTA
** * * * * *

AcMRJP4 ATGTAACGACTTGATACGGAATAGTCGTTGCGCAAATTTGACAATCAGGATAATAATC
AmMRJP4 ACGTAAACGACTTGATACGGAATAGTCGTTGCGCAAATTTGACAATCAGGATAATAATC
* * * * *

AcMRJP4 AACATAATTATAATCATAATCAAGTTCGTCATTTCTTCAAATCTGACAATCAGAATAACA
AmMRJP4 ACTATAATCATAATCATAATCAAGCTCGTCATTTCTTCAAATCTGACAATCAGAATAACA
* * * * *

AcMRJP4 ATCAACATAACATCAAGCTTATCATTCTTCAAAGTCTGACAATTTGGGATAACAATAACA
AmMRJP4 ATCAACATAACGATCAAGCTCATCATTCTTCAAAGTCTAACAATCGGCATAACAATAACG
***** * * * * *

AcMRJP4 ATCAAGCTCATCATTCTTCAAATTTGATAATCAGAATAACAATCAATATAACAATTAGG
AmMRJP4 ATTAAGCTCATCATTCTTCAAATTTGATAATCAGAATAACAATCAGAATAACGATTAAT
** * * * * *

AcMRJP4 TTCATCATTCTTCAATCATGTTAAATCTGATAATTAATCTTTTTCTCGATGTAAGTC
AmMRJP4 ATAATAATCAATTTTATCATTCTTTAAATCTGTTAATTAATCTTTTTCTCGATGTAAGTC
* * * * *

AcMRJP4 AAATATTTTAAAAA-TTTCATTACATTATAAAACGAATAAAATAAATATCGTTTTTC-G
AmMRJP4 AAATATTTTAAAAAATTTTCATTACATTATAAAACGA-TAAATAAATATCGTTTTTTTG
***** * * * * *

AcMRJP4 CATAAAAAAAAAAAAAAAAAAAAAAAAAA
AmMRJP4 CATAAT-----

Figure 3.9 (continued)

CLUSTAL X (1.81) multiple sequence alignment

```

AcMRJP4      MTKWLLLMACLGIAQNIIRGAVVRENSSRKLTNTLNV IHEWKYVDYDFGSDEKRQAAIQ
AmMRJP4      MTKWLLLMVCLGIACQNIIRGGVVRENSSGKLTNTLNV IHKWKYLDYDFDNDERRQAAIQ
*****.*****.***** *:*****:***:***. .*:*****

AcMRJP4      SGEYDRTKNYPLDQVQWHDKTFVMTLRYDGVPSLNVVSDKTGNGGPLLQYPDWSFAKY
AmMRJP4      SGEYDRTKNYPLDQVQWHDKTFVMTLRYDGVPSLNVVSDKTGNGGRLQYPDWSFAKY
*****:***:***:*****:*****:*****:*****:*****:*****

AcMRJP4      EDCSGIVSANKIAIDEYERLWVLD SGLVNNIQPMCS PKLLAFDLTTSKLLKQVEI PHDVA
AmMRJP4      EDCSGIVSAHKIAIDEYERLWVLD SGLVNNIQPMCS PKLFAFDLNTS QLLKQVEI PHDVA
*****:*****:*****:*****:*****:*****:*****:*****

AcMRJP4      VNATGKGGGLASLAVQAMDSVNTMVY MADNKDDALIVYQNADDSFHRLSSHISNHNFRSD
AmMRJP4      T--TGKGELVSLTVQAMDSVNTMVY MVDNKN-TLIIYQNADDSFHRLSSHTLNHN--SD
.   **** *:***:*****.*****.***: :**:*:*****:***** ** **

AcMRJP4      KMS--QENLTLKEVDNRVFGMALSSVTHNLYYSPLSSQNL YVNTTSLMNSQNQGNDVQY
AmMRJP4      KMSDQQENLTLKEVDNKVYGMALSPVTHNLYYNSPSENLYYVNTESLMKSENQGNDVQY
***  *****:***:*****.*****. .*:***** ***:***:*****

AcMRJP4      ESVQDVFSSQLSAKAVSKNGVLF FFGFTNNTLGCWNEHQSLDRQNIDIVARN-ETLQMVVG
AmMRJP4      ERVQDVFDSQLTVKAVSKNGVLL FGLANNTLSCWNEHQSLDRQNIDIVARNEDTLQMVVS
*   *****.***: .*****:***: :***.*****:*****:*****:*****

AcMRJP4      MKIKQNL PQSGKVNNTQRNEHLLALTNKKQDVLNNDLNLEHVNFQILDANVNDLIRNSRC
AmMRJP4      MKIKQNV PQSGRVNNTQRNEYLLALSDRNQVNLNNDLNLEHVNFQILGANVNDLIRNSRC
*****:***:*****:*****:***: :*:*****:*****:*****

AcMRJP4      ANSDNQDNNQHYNHNQVRHSSKSDNQNNQHNNQAYHSSKSDNWDNNNNQAHSSKFDN
AmMRJP4      ANFDNQDNNQHYNHNQVQARHSSKSDNQNNQHNDQAHSSKSNRRHNNND-----
**  *****:***:*****.*****:***:*****:*.***:

AcMRJP4      QNNNQYNN
AmMRJP4      -----

```

Figure 3.10 Alignment of the deduced amino acid sequence of AcMRJP4 cDNA with AmMRJP4 cDNA published sequence (GenBank Acc. Z26319). Conserve residues are indicated in asterisks. : means amino acid which have the same group of side chains and similar size while . means amino acid which have the same group of side chains but different size.

AcMRJP5

Recombinant plasmids containing 1,900 bp cDNA which might be AcMRJP5 cDNA as analyzed by *EcoRI* and *SspI* digestion. Further identification was performed by sequencing the insert cDNA using five primers as showed in Table 2.1 and Figure 2.1.

The recombinant plasmids were sequenced using M13 forward and M13 reverse primer. Three internal sequencing primers (primer name MRJP5_A, MRJP5_B and MRJP5_C) were designed from nucleotide sequences obtained. The inserts of two recombinant clones were sequenced along the entire length and the insert of one recombinant clone was partially sequence. The nucleotide sequence of two recombinant clone and partial sequence of the another clone were assembled (Appendix D). The nucleotide sequence was compared with the DNA sequence deposited in GenBank database using nucleotide Blast (BlastN) and translated protein Blast (BlastX). The result shown that similar to AmMRJP5 cDNA. This sequence was most likely to be AcMRJP5 cDNA.

AcMRJP5 cDNA indicated a length of 1881 bp (including poly (A) tail) the sequence contained an open reading frame (nucleotides 1-1740) which encoded 579 amino acid residues (Figure 3.11). The sequence AATAAAATAAA containing two partially overlapping consensus polyadenylation signal sequences (AATAAA) was located 14 bp upstream from the poly (A) tail. The 3'-terminal sequence was observed nonanucleotide repeat sequence, GATAGAATG that encoded to tripeptide (DRM).

The computer sequence analysis predicted that the signal peptidase cleavage site was located between Gly 20 and Ala 21. A comparison of AcMRJP5 and AmMRJP5 nucleotide sequences analyzed by blast N program revealed an identity of 91%. For blast

X program, protein sequence of AcMRJP5 showed 90% sequence identity and 96% sequence similarity to AmMRJP5.

The deduced amino acid (without putative signal peptide) composition of AcMRJP5 comprised of 42.04% hydrophobic, 24.51% neutral and 33.45% hydrophilic amino acid residues. The essential amino acid content was 51.9%.

The pI value was estimated to be 8.75. The estimated molecular weight was 66.2 kDa. Five putative *N*-glycosylation sites were found.

The nucleotide sequence and deduced amino acid sequence of AcMRJP5 cDNA were compared with the nucleotide sequence and deduced amino acid sequence of AmMRJP5 cDNA. The result of alignment was shown in Figure 3.12 and Figure 3.13.

The deduced amino acid of AcMRJP5 inferred from AcMRJP5 cDNA show the extensive repeat region located between amino acid residue 367 and 520. This repeat region located at the C-terminal of this protein. The repetitive region consists of a 51-fold repeated tripeptide motif with dominance of DRM sequence motifs (Figure 3.13). The result from nucleotide sequencing of 3 recombinant clone contained AcMRJP5 cDNA insert show the repeat region of the AcMRJP5 was polymorphism that invariant in repeated tripeptide motif (Appendix D).

The repetitive region of *A. cerana* was located at the same position as found in *A. mellifera* but smaller in size, that occurred 51 times compared with 58 times in *A. mellifera* (Albert *et al.*, 1999a)

ATGACAAGTTGGTTGTTGCTGGTGGTGTGCCTTGGCATAGCTTGTCAAGGTATCACAGGC 60
 M T S W L L L V V C L G I A C Q G I T G **↑**
GCCACTGTTTCGAGAAAATTCCTTCGAGAAAATTTGGCAAATTCGATGAACGTGATTACAGAA 120
 A T V R E N S S R N L A N S M N V I H E
TGGAAGTATCTTGATTATGACTTCGGTAGCGACGAAAAAAGACAAGCTGCGATTCAATCT 180
 W K Y L D Y D F G S D E K R Q A A I Q S
GGCGAATATGACCATACGAAAAATTATCCCTTCGATGTCGATCGATGGCATGATATGACT 240
 G E Y D H T K N Y P F D V D R W H D M T
TTTGTCAACCGTACTAAGATACAAAGGTGTACCTTCTCTTTAAACGTGATATCTAAGAAA 300
 F V T V L R Y K G V P S S L N V I S K K
ATTGGCAACGGTGGACCTCTTCTGCAGCCATATCCTGATTGGTCGTGGGCGAACTATAAA 360
 I G N G G P L L Q P Y P D W S W A N Y K
GATTGCTCTGGAATCGTGAGCGCTTACAAAATTGCGATCGACAAGTTCGACAGATTGTGG 420
 D C S G I V S A Y K I A I D K F D R L W
GTTCTGGACTCAGGTATTATCAATAATACTCAACCCATGTGTTACCAAAAATTGCATGTC 480
 V L D S G I I N N T Q P M C S P K L H V
TTTGATCTCAATACCTCACAGCAGATTAAGCAAGTTATGATGCCGATGATATTGCCATA 540
 F D L N T S Q Q I K Q V M M P H D I A I
AATGCCACTACAGGAAAAAGGAGGACTAGAAAATCTAGTTGTTCAAGCTATGGATCCTATG 600
 N A T T G K G G L E N L V V Q A M D P M
AATACTCTGGTGTATATGGCAGATAACAAGGGTGTATGCTTTAATTGTTTATCAAAATTCC 660
 N T L V Y M A D N K G D A L I V Y Q N S
GATGATTCCTTCCATCGATTGACTTCCAACACTTTTCGATTACGATCCCAAATATATCAAA 720
 D D S F H R L T S N T F D Y D P K Y I K
ATGATGGCCGAGGAGAAAAGTTTCACATTGCAAGATGGAATTTTTGGAATGGCACTCAGT 780
 M M A A G E S F T L Q D G I F G M A L S
CCCATGACAAACAATCTTTATTACAGTCCTCTCGCTTCTCGCAGTTTGTATTATGTTAAT 840
 P M T N N L Y Y S P L A S R S L Y Y V N
ACGAAACCCTTCATGAAATCACAATATGGAACAAATAACGTACAACATGAAGGTGTCCAA 900
 T K P F M K S Q Y G T N N V Q H E G V Q
GATATTTTCAATACTCAATCAATTGCTAAAATAATGTCGAAAAATGGCGTTCTCTTTTTC 960
 D I F N T Q S I A K I M S K N G V L F F
GGTCTCATGAATAATTCAGCTATTGGTTGTTGGAATGAGCACCAACCACTTCAGAGACAA 1020
 G L M N N S A I G C W N E H Q P L Q R Q
AATATGGATATGGTCGCTCAGAATGAAGAGACACTTCAAACGGTCGTTGCTATGAAAATG 1080
 N M D M V A Q N E E T L Q T V V A M K M
ATGCATCTCCACAATCCAACAGGATGAATAGGATGCATAGGATGAATAGTATGAATAGA 1140
 M H L P Q S N R M N R M H R M N S M N R

(continued)

Figure 3.11 Nucleotide and deduced amino acid sequences of AcMRJP5. Initiation and termination of translational codons and putative polyadenylation signal are boldfaced. The signal peptidase cleavage site was indicated by arrow. *N*-linked glycosylation sites are underlined.

ATGGATAGGATGGATAGAAATGGATAGGATGGATAGGATGGATAGGATGGATAGGATGGAT 1200
 M D R M D R M D R M D R M D R M D R M D R M D
 AGGATGGATAGGATGGATAGGATGGATAGGATGGATAGAAATGGATAGAAATGGATAGGATG 1260
 R M D R M D R M D R M D R M D R M D R M D R M
 GATAGGATGGATATAATGGATAGGACGAATAAAATGGATAGGATGGATAGGATGGATATA 1320
 D R M D I M D R T N K M D R M D R M D I
 ATGGATAAGATGAATAAAATGGATAGGATGGATAGTATGATTAGAATAGATAAAATGGAT 1380
 M D K M N K M D R M D S M I R I D K M D
 AGAATGGATAGAAATGCATAGAATAGATATAATGAATAGAATGGATAGAAATGGATAGAAATG 1440
 R M D R M H R I D I M N R M D R M D R M
 GACACAAGAATAGATACAAGAATGGACAGAATGGATAGAAATGGATAAAATGGATAAGATA 1500
 D T R I D T R M D R M D R M D K M D K I
 AATAAGATGCATAGGATGGGTAGGATGGATAGGATGGATAGAAATGAATAGAATGAATAGA 1560
 N K M H R M G R M D R M D R M N R M N R
 CAAATGAATGAATATATGATGGCTTTAAGTATGAAATTACAGAAATTTATAACAATGAT 1620
 Q M N E Y M M A L S M K L Q K F I N N D
 TATAATTTCAACGAAGTAAATTTCCGAATTTTGGCTGCAAATGTAAACGATTTAATAATG 1680
 Y N F N E V N F R I L A A N V N D L I M
 AACACTCGTTGTGCAAATTTCTAACAATCAGAATGATAATCAAATAAGCATAATAATTA 1740
 N T R C A N S N N Q N D N Q N K H N N *
 GGTAGTCGTTCTTTATATTAATCTGTTAATTAGTCTTTTCTCGACTATAAACCAAATA 1800
 TTGTTTCAAATTTCTTTATATTATAAATGAATAAAATAAATATCGTTTTTGCTTAAAAAA 1860
 AAAAAAAAAAAAAAAAAAAAAA 1881

Figure 3.11 (continued)

ศูนย์วิทยทรัพยากร
 จุฬาลงกรณ์มหาวิทยาลัย

CLUSTAL X (1.81) multiple sequence alignment

```

AcMRJP5 -----ATGA
AmMRJP5 TACTACTGCGTTCTCTTGAAACTGTCGTTTGCAAATATTTGCAGCATCCAAGAACAATGA
          ****

AcMRJP5 CAAGTTGGTTGTTGCTGGTGGTGCCTTGGCATAGCTTGTCAAGGTATCACAGGCGCCA
AmMRJP5 CAACTTGGTTGTTGCTGGTGGTGCCTTGGCATAGCTTGTCAAGGTATCACAGGCGTCA
          *** ***** **

AcMRJP5 CTGTTTCGAGAAAATTCTTCGAGAAAATTTGGCAAATTCGATGAACGTGATTACGAATGGA
AmMRJP5 CTGTTTCGAGAAAATTCTCCGAGAAAGTTGGCAAATTCGATGAACGTGATTACGAATGGA
          ***** **

AcMRJP5 AGTATCTTGATTATGACTTCGGTAGCGACGAAAAAGACAAGCTGCGATTCAATCTGGCG
AmMRJP5 AGTATCTCGATTATGATTTGGTAGCGACGAAAGGAGGCAAGCTGCGATGCAATCTGGCG
          ***** **

AcMRJP5 AATATGACCATACGAAAAATTATCCCTTCGATGTCGATCGATGGCATGATATGACTTTTG
AmMRJP5 AGTATGACCATACGAAAAATTATCCCTTCGATGTCGATCAATGGCGTGGTATGACTTTTG
          * ***** **

AcMRJP5 TCACCGTACTAAGATACAAAGGTGTACCTTCCTCTTTAAACGTGATATCTAAGAAAATTG
AmMRJP5 TAACCGTACCAAGATACAAAGGTGTACCTTCCTCTTTGAACGTGATATCTGAGAAAATTG
          * ***** **

AcMRJP5 GCAACGGTGGACCTCTTTCGAGCCATATCCTGATTGGTTCGTGGGCGAACTATAAAGATT
AmMRJP5 GCAACGGTGGACGACTTCTACAACCGTATCCTGATTGGTTCGTGGGCGAACTATAAAGATT
          ***** **

AcMRJP5 GCTCTGGAATCGTGAGCGCTTACAAAATGCGATCGACAAGTTCGACAGATTGTGGGTTG
AmMRJP5 GCTCTGGAATAGTGAGCGCTTACAAAATGCGATCGACAAGTTCGACAGATTGTGGATTG
          ***** **

AcMRJP5 TGGACTCAGGTATTATCAATAACTCAACCCATGTGTTACCAAAAATTGCATGTCTTTG
AmMRJP5 TGGACTCAGGTATTATCAATAACTCAACCCATGTGTTACCAAAAATTGCATGTCTTTG
          ***** **

AcMRJP5 ATCTCAATACCTCACAGCAGATTAAGCAAGTTATGATGCCGCATGATATTGCCATAAATG
AmMRJP5 ATCTCAATACCTCACATCAGCTTAAGCAAGTTGTGATGCCGCACGATATTGCCGTAATG
          ***** **

AcMRJP5 CCACTACAGGAAAAGGAGGACTAGAAAATCTAGTTGTTCAAGCTATGGATCCTATGAATA
AmMRJP5 CCAGCACAGGGAATGGGGACTCGTATCACTAGTTGTTCAAGCTATGGATCCTGTGAATA
          *** ***** **

AcMRJP5 CTCTGGTGTATATGGCAGATAACAAGGGTGATGCTTTAATGTTTATCAAAAATTCGATG
AmMRJP5 CTATCGTGTATATGGCAGATGACAAAGGTGATGCTTTAATCGTCTACCAAAAATTCGACG
          ** * ***** **

AcMRJP5 AATCTTCCATCGATTGACTTCCAACACTTTCGATTACGATCCCAAATATATCAAAAATGA
AmMRJP5 AATCTTCCATCGATTGACTTCCAACACTTTCGATTACGATCCCAAATATATCAAAAATGA
          * ** ***** **

```

(continued)

Figure 3.12 Alignment of the nucleotide sequence of AcMRJP5 cDNA with AmMRJP5 cDNA published sequence (GenBank Acc. AF004842). Conserve residues are indicated by asterisks.

AcMRJP5 TGGCCGCAGGAGAAAGTTTCACATTGCAAGATGGAATTTTTGGAATGGCACTCAGTCCCA
AmMRJP5 TGGACGCGGGAGAAAGTTTCACAGCGCAAGATGGAATTTTTGGAATGGCACTCAGTCCCA
*** **

AcMRJP5 TGACAAACAATCTTTATTACAGTCTCTCGCTTCTCGCAGTTTGTATTATGTTAATACGA
AmMRJP5 TGACAAACAATCTTTATTACAGCCTCTTTCTTCTCGCAGTTTGTATTATGTTAATACAA
***** **

AcMRJP5 AACCTTCATGAAATCACAATATGGAACAAATAACGTACAACATGAAGGTGTCCAAGATA
AmMRJP5 AACCTTCATGAAATCAGAATATGGAGCAAATAACGTACAATATCAAGGTGTCCAAGATA
**** **

AcMRJP5 TTTTCAATACTCAATCAATTGCTAAAATAATGTCGAAAAATGGCGTTCCTTTTTCCGGTC
AmMRJP5 TTTTCAACACTGAATCGATTGCTAAAATAATGTCGAAAAATGGCGTTCCTTTTTCCGGCC
***** **

AcMRJP5 TCATGAATAATTCAGCTATTGGTTGTTGGAATGAGCACCAACCACTTCAGAGACAAAATA
AmMRJP5 TCATGAATAATTCAGCTATTGGTTGTTGGAACGAGCATCAACCACTTCAGAGAGAAAATA
***** **

AcMRJP5 TGGATATGGTCGCTCAGAATGAAGAGACTTCAAACGGTCGTTGCTATGAAAATGATGC
AmMRJP5 TGGATATGGTCGCTCAGAATGAAGAGACTTCAAACGGTCGTTGCTATGAAAATGATGC
***** **

AcMRJP5 ATCTCCACAATCCAACAGGATGAATAGGATGCATAGGATGAATAGTATGAATAGATGG
AmMRJP5 ATCTCCACAATCCAACAAGATGAATAGGATGCATAGGATGAATAGAGTGAATAGAGTGA
***** **

AcMRJP5 ATAGGATGGATAGAATGGATAGGATGGATAGGATGGATAGGATGGATAGGATGGATAGGA
AmMRJP5 ATAGAATGGATAGAATGGATAGAATAGATAGGATGGATAGGATGGATAGGATGGATACAA
**** **

AcMRJP5 TGGATAGGATGGATAGGATGGATAGGATGGATAGAATGGATAGAATGGATAGGATGGATA
AmMRJP5 TGGATACAATGGATAGAATAGATAGGATGGATAGGATGGATAGAATAGATAGGATAGATA
***** **

AcMRJP5 GGATGGATATAATGGATA-----
AmMRJP5 GGATGCATACAATGGATACAATGGATACAATGGATAGAACAGATAAGATGAGTAGCATGG
***** **

AcMRJP5 ---GGACGAATAAAATGGATAGGATGGATAGGATGGATATAATGGATAAGATGAATAAAA
AmMRJP5 ATAGGATGGATAGAATGGATAGGGTGGATAGGATGGATACAATGGATAGAACAGATAAGA
*** **

AcMRJP5 TGGATAGGATGGATAGTATGATTAGAATAGATAAAAATGGATAGAATGGATAGAATGCATA
AmMRJP5 TGAGTAGCATGGATAGGATGGATAGAATGGATAGGGTGGATACAATGGATACAATGGATA
** **

AcMRJP5 GAATAGATATAATGAATAGAATGGATAGAATGGATAGAATGGACACAA-----GAATAG
AmMRJP5 CAATGGATAGAATGGATAGGATGGATAGGATGGATAGAATGGATAGAATGGATAGGATGG
*** **

AcMRJP5 ATACAA-----GAATGGACAGAATGGATAGAATGGATAAAAATGGATAAGATAAATAAGA
AmMRJP5 ATACAATGGATAGAACAGATAAGATGAGTAGGATAGATAGAATGGATAAAAATAGATAGAA
***** **

AcMRJP5 TGCATAGGATGGGTAGGATGGATAGGATGGATAGAATGAATAGAATGAATAGACAAATGA
AmMRJP5 TGGATAGGATGGATAGGACAAAATAGAATGGATAGAATGAATAGGATGAATAGACAAATGA
** *****

Figure 3.12 (continued)

```

AcMRJP5      ATGAATATATGATGGCTTTAAGTATGAAATTACAGAAATTTATAAACAATGATTATAATT
AmMRJP5      ATGAATATATGATGGCTTTAAGTATGAAATTACAGAAATTTATAAATAATGATTATAATT
*****

AcMRJP5      TCAACGAAGTAAATTTCCGAATTTTGGCTGCAAATGTAAACGATTTAATAATGAACACTC
AmMRJP5      TCAACGAAGTAACTTCCGAATTTTGGGTGCAAATGTAAACGATTTAATAATGAATACTC
*****

AcMRJP5      GTTGTGCAAATTCTAACAATCAGAATGATAATCAAATAAGCATAATAATTAAGGTAGTC
AmMRJP5      GTTGTGCAAATTCTGACAATCAGAATAACAATCAAATAAGCATAATAATTAAGATGATC
*****

AcMRJP5      GTTCTTTATATTAAAATCTGTTAATTAGTCTTTTCTCGACTATAAACCAAATATTGTTTC
AmMRJP5      GTTCTTTATATTAAAATCTGTTAATCAGTCTTTTCTCGA-TATAAACCAAATATTCTTTA
*****

AcMRJP5      AAATTTCTTTATATTATAAATGAATAAAATAAATATCGTTTTTGCTTAAAAAAAAAAAA
AmMRJP5      AAATTTCTTTATATTATAAATGAATAAAATAAATAT---TTTGCATGAT-----
*****

AcMRJP5      AAAAAAAAAAAAAA
AmMRJP5      -----

```

Figure 3.12 (continued)



AcMRJP6

Further identification of 1500 bp cDNA insert of recombinant plasmid was performed by sequencing of the insert DNA using four primers that showed in Table 2.1 and Figure 2.1. The insert DNA was sequenced using M13 forward and M13 reverse primers. One internal sequencing primer (MRJP6_A) was designed from nucleotide sequences obtained. Another primer (MRJP4_2) that had been used for sequence analysis of AcMRJP4 was used for sequence analysis of this DNA insert, even though one base mismatch at 5' termini was found in this primer.

The sequence of AcMRJP6 was obtained from sequencing of the insert DNA from three recombinant clones (Appendix E). One of these sequences shows 100% identity with assembled sequence. The nucleotide sequence obtained was compared with the DNA sequence deposited in GenBank database using nucleotide Blast (BlastN) and translated protein Blast (BlastX). The result showed that the sequence is nearly the same to AmMRJP6 cDNA. This sequence was designated AcMRJP6.

AcMRJP6 cDNA had a length of 1450 bp (including poly(A) tail). The sequence contains an open reading frame (nucleotides 1-1308) which encoded 435 amino acid residues (Figure 3.14). The sequence AATAAAATAAA containing two partially overlapping consensus polyadenylation signal sequences (AATAAA) was located 14 bp upstream from the poly(A) tail. The computer sequence analysis predicted that the signal peptidase cleavage site was located between Ser 20 and Ala 21. A comparison of AcMRJP6 and AmMRJP6 nucleotide sequences analysis by blast N program revealed an identity of 92%. For blast X program protein sequence of AcMRJP6 was shown 88% sequence identity and 93% sequence similarity to AmMRJP6. The deduced amino acid

(without putative signal peptide) composition of AcMRJP6 comprised of 42.89% hydrophobic, 29.40% neutral and 27.71% hydrophilic amino acid residues. The essential amino acid content was 46.8%.

The pI value was estimated to be 6.44. The estimated molecular weight was 47.4 kDa. Two putative *N*-glycosylation sites were found.

The nucleotide sequence and deduced amino acid sequence of AcMRJP6 cDNA were compared with the nucleotide sequence and deduced amino acid sequence of AmMRJP6 cDNA. The result of alignment was shown in Figure 3.15 and Figure 3.16.

AcMRJPs characterization

A summary for molecular characterization of cDNA and deduced amino acid sequences of AcMRJPs are illustrated in Table 3.2

The amino acid composition of AcMRJPs are illustrated in Table 3.3. Four AcMRJPs contained high amounts of the 10 essential amino acid: MRJP5 (51.9%), MRJP1 (47.4%), MRJP6 (46.8%) and MRJP2 (45%). MRJP5 is rich in Arg and Met (8.9% and 12.5%). MRJP6 is rich in Ile and Phe (7.0% and 4.1%). MRJP3 and MRJP4 have a lower overall content of essential amino acids, but they possess relatively higher amount of some of them; MRJP3 had Arg (5.6%), Lys (6.6%) and MRJP4 had Leu (8.6%), Val (7.7%).

Alignment of deduced amino acid sequence of AcMRJP1-6 was show in Figure 3.17. Four cysteine residues are conserved in these proteins. The regions of high sequence similarity was found in N-terminal region of the protein. Protein sequence of PYPDWS, DCSGIVS, RLWVLDS, NLYYSP and LYYVNT are conserved among AcMRJP.

ATGACAAAATGGTTGCTGCTGATAGTGTGTCTTAGCATAGCTTGTCAAGATGTCACAAGC 60
 M T K W L L L I V C L S I A C Q D V T S **↑**
 GCGATTCATCGAAGAAAATCTTCAAAAAATTTGGAACATTTCGATGAACGTGATTACAGAA 120
 A I H R R K S S K N L E H S M N V I H E
 TGGAAATATCTTGATTATGATTTTCGATACCAATGAAAAAAAACAAGCTGCGATTCAATTT 180
 W K Y L D Y D F D T N E K K Q A A I Q F
 GGTGAATACGACTATACGAAAAATTATCCCTTTGACGTCGATCAATGGCATGATAAGACT 240
 G E Y D Y T K N Y P F D V D Q W H D K T
 TTTGTCGCTGTAATAAGATACGATGGTGTACCTTCTCTTTGAACGTGATATCTGACAAA 300
 F V A V I R Y D G V P S S L N V I S D K
 ACTGGCAACGGTGGACGCCTTCTCCAACCGTATCTGATTGGTCGTGGACGAACTATAAA 360
 T G N G G R L L Q P Y P D W S W T N Y K
 GATTGTTCTGGAATCGTGAGCGTTTACAAAATTGCGATTGACAAAATTCGACAGATTGTGG 420
 D C S G I V S V Y K I A I D K F D R L W
 GTTCTGGACTCAGGTCTTATTAATAATATTCAACTTATGTGTTCTCCAAAATTGCTTGCC 480
 V L D S G L I N N I Q L M C S P K L L A
 TTTGATCTGACAACCTTCGAAATTGCTCAAGCAAGTCGAGATACCGTACGATATTGCTGTA 540
 F D L T T S K L L K Q V E I P Y D I A V
 AATGCCAGCACAGGAATGGGAGGACTCGTCTCATTAGTTGTTCAAGCTATGGATCCTATG 600
N A S T G M G G L V S L V V Q A M D P M
 AATACTATGGTATATATAGCAGATGACAGAGGTGACGCTTTAATCATCTATCAAAATTCC 660
 N T M V Y I A D D R G D A L I I Y Q N S
 GATGATTCTTTCCATCGATTGAGTTCCAATACTTTTGATAACGATCCCAGATATTCTGAA 720
 D D S F H R L S S N T F D N D P R Y S E
 TTGACGGTCGCGGGAGAAAGTTTTCACAGTGCATGATGGAATTTTTGGAATGGCACTTAGT 780
 L T V A G E S F T V H D G I F G M A L S
 CCTGTGACGAACAATCTTTATTATAGCCCTCTCACTTCTCACAGTTTGTATTACGTTAAC 840
 P V T N N L Y Y S P L T S H S L Y Y V N
 ACGGAACCATTTATGAAATCACAATATGGAGAAAATAATATAACAATATGAAGGAATTCAA 900
 T E P F M K S Q Y G E N N I Q Y E G I Q
 GATATTTTCAACACTCAATCATCCGCTAAAGTAATGTCGAAAAATGGCGTCCTTTTCTTC 960
 D I F N T Q S S A K V M S K N G V L F F
 GGACTTGTGAATAATTCAGCTATTGGTTGTTGGAACGAGCATCAACCCTTCAGAAACAA 1020
G L V N N S A I G C W N E H Q P L Q K Q
 AATATGGATATGGTCGCTCAGAATGAAGAGACACTTCAAATAATCACTAGTGTGAAAATT 1080
 N M D M V A Q N E E T L Q I I T S V K I
 ATACAAAATCTTCCATATTCGGGAAAGATGAATAGAATTCACAAGAATGAATATATGTTG 1140
 I Q N L P Y S G K M N R I H K N E Y M L

(continued)

Figure 3.14 Nucleotide and deduced amino acid sequences of AcMRJP6. Initiation and termination of translational codons and putative polyadenylation signal are boldfaced. The signal peptidase cleavage site was indicated by arrow. *N*-linked glycosylation sites are underlined.

GCTTTAAGTAACAGAATGCAGAAAATAGTAAACAATGATTTTAATTTCAACGACATAAAAT 1200
 A L S N R M Q K I V N N D F N F N D I N
 TTCCGAATATTGGGTGCGAATGTAAAGAACTTAATAAAAAACACTCGTTGTGCAAATTCT 1260
 F R I L G A N V K N L I K N T R C A N S
 AAAAATCAGAATAACAATCAAAAAGAAACATAAGAATCAAGCTCAT**TAGATCTTTTCCAAG** 1320
 K N Q N N N Q K K H K N Q A H *
 ATCATATTAAATTCTATAGATTAATTTTTTCTCGTGGTAAATCAAATATTTTTTAAAAATT 1380
 TATTTGCATTATAAATTA**AATAAAATAAA**TATCATTTCGCATAAAAAAAAAAAAAAAAAAAAA 1440
 AAAAAAAAAA 1450

Figure 3.14 (continued)



CLUSTAL X (1.81) multiple sequence alignment

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AcMRJP6 -----
AmMRJP6 ATTAAATATTTGCAGCTTCCCTCCATAAGTGTTCCATATATCTTAATTGTATTATTG

AcMRJP6 -----ATGACAAAATGGTTGCTGCT
AmMRJP6 CAATCTTTCATTTATCTAACACGAAATATTTTGTAGAAAAATGACAAATGGTTACTGCT
          *****  *****  *****

AcMRJP6 GATAGTGTGTCTTAGCATAGCTTGTCAAGATGTCAAGCGCGATTTCATCGAAGAAAATC
AmMRJP6 GATAGTGTGTCTTAGCATAGCTTGTCAAGATGTCAAGCGCGATTTCATCAAAGAAAATC
          *****  *****  *****

AcMRJP6 TTCAAAAAATTTGGAACATTTCGATGAACGTGATTACGAATGGAATATCTTGATTATGA
AmMRJP6 TTCAAAAAATTTGGAACATTTCGATGAACGTGATTACGAATGGAATATATCGATTATGA
          *****  *  *****

AcMRJP6 TTTTCGATACCAATGAAAAAACAAGCTGCGATTCAATTTGGTGAATACGACTATACGAA
AmMRJP6 TTTTGGTAGTGATGAAAAAAGACAAGCTGCGATTCAATCTGGCGAATACGATTATACGAA
          *** * *  *****  *****  *****  *****

AcMRJP6 AAATTATCCCTTTGACGTCGATCAATGGCATGATAAGACTTTTGTCTGCTGTAATAAGATA
AmMRJP6 AAATTATCCTTTTCGACGTCGATCAATGGCATAATAAGACTTTTCTGCTGTAATAAGATA
          *****  **  *****  *****  *****  *****

AcMRJP6 CGATGGTGTACCTTCCTCTTTGAACGTGATATCTGACAAAATGGCAACGGTGGACGCCT
AmMRJP6 CGATGGTGTACCTTCCTCTTTGAACGTGATATCTGAGAAAATGGCAACGGTGGATGCCT
          *****  *****  *****  *****  *****

AcMRJP6 TCTCCAACCGTATCCTGATTGGTCTGGACGAACATAAAGATTGTTCTGGAATCGTGAG
AmMRJP6 TCTACAACCGTATCCTGATTGGTCTGGGCGAACATAAAGATTGTTCTGGAATAGTGAG
          ***  *****  *****  *****  *****

AcMRJP6 CGTTTACAAAATTCGATTGACAAATTCGACAGATTGTGGGTTCTGGACTCAGGTCTTAT
AmMRJP6 CGTTTACAAAATTCGATTGACAAATTCGACAGATTGTGGGTTCTGGACTCAGGTCTTAT
          **  *****  *****  *****  *****

AcMRJP6 TAATAATATTCAACTTATGTGTTCTCCAAAATGCTTGCCTTTGATCTGACAACTTCGAA
AmMRJP6 TAATAATATTCAACTTATGTGTTCTCCAAAATTAATGCTTGCCTTTGATCTCAATACCTCAA
          *****  *****  *****  *****  *  **  *  *

AcMRJP6 ATTGCTCAAGCAAGTCGAGATACCGTACGATATTGCTGTAATGCCAGCACAGGAATGGG
AmMRJP6 GTTGCTTAAACAAATCGAGATACCACATAATATTGCCGTAATGCCAGCACAGGAATGGG
          *****  **  *  *  *****  *  *****  *****  *****

AcMRJP6 AGGACTCGTCTCATTAGTTGTTCAAGCTATGGATCCTATGAATACTATGGTATATATAGC
AmMRJP6 AGGACCCGTATCGTAGTTGTTCAAGCTATGGATCCTATGAATACTACGGTGTATATAGC
          *****  ***  *  *  *****  *****  *****  *****

AcMRJP6 AGATGACAGAGGTGACGCTTTAATCATCTATCAAATTCGATGATTCTTTCCATCGATT
AmMRJP6 AGACGACAGAGGTGACGCTTTAATCATCTATCAAATTCGATGATTCTTTCCATCGATT
          ***  *****  *****  *****  *****  *****

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(continued)

Figure 3.15 Alignment of the nucleotide sequence of AcMRJP6 cDNA with AmMRJP6 cDNA published sequence (GenBank Acc. AY313893). Conserve residues are indicated by asterisks.

AcMRJP6 GAGTTCCAATACTTTTGATAACGATCCCAGATATTCTGAATTGACGGTCGCGGGAGAAAG
AmMRJP6 GACTTCCAAAACCTTTTGATAACGATCTCAGATATTCTGAACTGGCCGTCGCGGGAGAAAG
** ***** ***** ***** ***** ** * *****

AcMRJP6 TTTACAGTGCATGATGGAATTTTGGAAATGGCACTTAGTCCTGTGACGAACAATCTTTA
AmMRJP6 TTTACAGTGCATGATGGAATTTTGGAAATGGCACTTAGTCCTGTGACGAACAATCTTTA

AcMRJP6 TTATAGCCCTCTCACTTCTCACAGTTTGTATTACGTTAACCGGAACCATTATGAAATC
AmMRJP6 TTACAGCCCTCTCACTTCTCACAGTTTGTATTATGTTAACATGGAACCATTATGAAATC
*** ***** ***** ***** *****

AcMRJP6 ACAATATGGAGAAAATAATATACAATATGAAGGAATTCAAGATATTTTCAACTCAATC
AmMRJP6 ACAATATGAAGAAAATAATATAGAATATGAAGGAATCCAAGATATTTTCAACTCAATC
***** ***** ***** ***** *****

AcMRJP6 ATCCGCTAAAGTAATGTCGAAAAATGGCGTCCTTTTCTTCGGACTTGTGAATAATTCAGC
AmMRJP6 GTCTGCTAAAGTAATGTCGAAAAATGGCGTCCTTTTCTTCGGACTTGTGAATAATTCAGC
** *****

AcMRJP6 TATTGGTTGTTGGAACGAGCATCAACCACTTCAGAAACAAAATATGGATATGGTCGCTCA
AmMRJP6 TATTGGTTGTTGGAACGAGCATCAACCACTTCAGAGACAAAATATGGATATGGTCGCTCA

AcMRJP6 GAATGAAAGACACTTCAAATAATCACTAGTGTGAAAATTATACAAAATCTCCATATTC
AmMRJP6 GAATGAAAAGACACTTCAAATGATCATTAGCGTGAAAATTATACAAAATCTGCATATTC
***** ***** ** * *****

AcMRJP6 CGGAAAGATGAATAGAATTCACAAGAATGAATATATGTTGGCTTTAAGTAACAGAATGCA
AmMRJP6 CGGAAGGATGAATAGAATTCACAAGAATGAATATATGTTGGCTTTAAGTAACAGAATGCA

AcMRJP6 GAAAATAGTAAACAATGATTTTAATTTCAACGACATAAATTTCCGAATATGGGTGCGAA
AmMRJP6 GAAAATAGTAAACAATGATTTTAATTTGACGAAAGTAACTTTTCAATTTGGGTGCGAA
***** ***** ** * *****

AcMRJP6 TGTAAAGAACTTAATAAAAAACACTCGTTGTGCAAATTTTAAAAATCAGAATAACAATCA
AmMRJP6 TGTAAATAACTTAATAAAAAACACTCGTTGTGCAAAGTCTAACAATCAGAATAACAATCA
***** ***** ***** ***** *****

AcMRJP6 AAAGAAACATAAGAATCAAGCTCATT-----AGATCTTTTCCAAGATCATATTAATTC
AmMRJP6 AAATAAATATAAGAATCAAGCTCATTTAGATTAGATCTTTTCCAAGATCATATTAATTC
*** ** ***** ***** ***** ***** **

AcMRJP6 TATAGATTAATTTTCTCGTGGTAAATCAAATATTTTAAAAATTTATTTGCATTATAA
AmMRJP6 TATAGATCAACTTTTCTCGTGGTAAATCAAATATTTTAAAAATTTATTTGCATTATAA
***** ** ***** ***** ***** ***** *****

AcMRJP6 ATTAATAAAATAAATATCATTTCGCATAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
AmMRJP6 ATGAATAAAATAAATATCGTTTCGCATG-----
** *****

AcMRJP6 AAAAAAAAAAAAA
AmMRJP6 -----

Figure 3.15 (continued)

Table 3.2 Molecular characterization of cDNAs and deduced amino acid sequences of AcMRJP.

Family	DNA insert size* (bp)	Deduced amino acid (residues)	No. of N- glycosylation site	Amino acid residues without signal peptide	Molecular weight (kDa)	pI	Reference
MRJP1	1421	433	3	413	46.7	5.40	AF525776
MRJP2	1565	463	2	446	50.6	7.78	AF525777
MRJP3	2005	608	6	588	67.3	8.79	Srisuparbh (2002)
MRJP4	1608	485	7	465	52.8	5.84	AY532368
MRJP5	1881	579	5	559	66.2	8.75	AY532369
MRJP6	1450	435	2	415	47.4	6.44	-

The characteristics were predicted by computer analysis of AcMRJP cDNA sequence and deduced amino acid sequence without their signal peptides.

* including polyA tail

Table 3.3 Amino acid composition of AcMRJPs

	MRJP1	MRJP2	MRJP3	MRJP4	MRJP5	MRJP6
Ala	4.1	5.2	5.1	5.2	4.3	4.8
Arg	4.1	2.5	5.6	3.2	8.9	2.9
Asn	8.0	14.8	20.2	14.0	10.0	10.6
Asp	8.0	6.1	8.0	8.0	11.3	7.0
Cys	1.5	1.1	0.9	0.9	0.7	1.0
Gln	3.6	5.6	8.3	6.9	4.3	5.3
Glu	4.1	4.3	3.2	3.2	2.1	3.4
Gly	5.6	5.6	4.6	3.9	3.8	5.1
His	2.4	2.7	2.0	3.7	2.3	2.7
Ile	5.8	5.4	3.2	3.0	5.4	7.0
Leu	9.2	8.1	4.9	8.6	5.2	7.7
Lys	5.3	8.1	6.6	5.6	5.4	6.7
Met	2.7	2.7	1.4	2.2	12.5	3.4
Phe	4.1	3.8	3.1	2.8	2.9	4.1
Pro	3.9	3.4	2.6	2.2	2.7	3.1
Ser	9.2	5.6	6.0	9.7	5.5	8.0
Thr	5.8	4.5	4.3	4.3	3.9	4.6
Trp	1.2	1.1	1.0	1.3	1.1	1.4
Tyr	4.6	3.6	2.7	3.9	3.4	5.1
Val	6.8	6.1	6.3	7.7	4.3	6.3
Ess. aa.	47.4 %	45 %	38.4 %	42.4 %	51.9 %	46.8 %

Percent content of amino acid in native protein was obtained by computer analysis of its sequence employing the program ProtParam. Essential amino acids are marked in boldface.

CLUSTAL X (1.81) multiple sequence alignment

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AcMRJP2      MTRWLFMVAACLGIAACQG---AIIRQ-NSAKNLENSLNVHEWKYIDYDFGSEERRQAAI
AcMRJP3      MTKWLLLIVVCLGIACQDVTSAAVNHQRKSSKNLAHSMKVIYEWKHIDYDFGSVERRDAAI
AcMRJP1      MTRWLFMVVCLGIVCQG-----TTSSILRGESLNKSLVSLVHEWKFFDYDFSDERRQDAI
AcMRJP5      MTSWLLLIVVCLGIACQGITG--ATVRENSSRNLANSNMNVIHEWKYLDYDFGSDEKRQAAI
AcMRJP6      MTKWLLLIVCLSIACQDVTS--AIHRRKSSKNLEHSMNVIHEWKYLDYDFDTNEKKQAAI
AcMRJP4      MTKWLLLMAACLGIAACQNIRG-AVVRENSSRKKLNTNLNVIHEWKYVDYDFGSDEKRQAAI
                **  **:::.*.*.*..          ..*  :::*:***.*****: *::: **

AcMRJP2      QSGEYDHTKNYPFDVDQWHDKTFVTILKYDGVPSLTLNMI SNKIGKGGRLLPYPDWSWAE
AcMRJP3      KSGEFDHTKNYPFDVDRWRDKTFVTVERFDGVPSSLNVVTKKGGKGPLLHPYPDWSWAN
AcMRJP1      LSGEYDYRKNYPSDVDQWHGKIFVTMLRYNGVPSSLNVI SKKIGDGGPPLQPYPDWFAK
AcMRJP5      QSGEYDHTKNYPFDVDRWHDMTFVTVLRYKGVPSLNVISKKI GNGGPLLQPYPDWSWAN
AcMRJP6      QFGEYDYTKNYPFDVDQWHDKTFVAVIRYDGVPSLNVISDKTGNNGGPLLQPYPDWSWTN
AcMRJP4      QSGEYDRTKNYPLDVDQWHDKTFVTMLRYDGVPSLNVVSDKTGNNGGPLLQPYPDWFAK
                **:*  ****  ***:*..  **::  ::.*****:*:..*  *.*  **:******:::

AcMRJP2      NKDCSGIVSAFKIAIDKFDRLWVLD SGLINRTEPICAPKLHVFDLKNTKHLKQIEIPHDI
AcMRJP3      YKDCSGIVSAFKIAVDKFDRLWVLD SSSLVNNQPMCS PKLVTFDLNTSKLLKQVEIPHNI
AcMRJP1      YDCSGIVSATKLAIDKCDRLWVLD SGLVNNTQPMCS PKLTFDLTTSQLLKQVEIPHDV
AcMRJP5      YKDCSGIVSAYKIAIDKFDRLWVLD SGI INNTQPMCS PKLHVFDLNTSQQIQVMMPHDI
AcMRJP6      YKDCSGIVSVYKIAIDKFDRLWVLD SGLINNIQLMCS PKLLAFDLTTSKLLKQVEIPYDI
AcMRJP4      YEDCSGIVSANKIAIDEYERLWVLD SGLVNNIQPMCS PKLLAFDLTTSKLLKQVEIPHDV
                .*****. *:***: *****:.*.  : :*:***  *****: :**: :***:

AcMRJP2      AVNATTGKGGLVSLVVQAMD P---MNTLVYIADHKGDALIVYQNSDDSFHRM TSNTFDYD
AcMRJP3      AVNATTEW GELVSLAVQAVDP---TNTMVYIADERGEASIIYQNSDDSFHRLTSNTFDYD
AcMRJP1      AVNATTGKGR LSSLAVQPLDCN INGDTMVYIAD EKGEGLIVYHDS DNSFHRLTSKTFDYD
AcMRJP5      AINATTGKG GLENL VVQAMD P---MNTLVY MADNKGDALIVYQNSDDSFHRLTSNTFDYD
AcMRJP6      AVNASTGM GGLVSLVVQAMD P---MNTMVYI ADDRGDALIIYQNSDDSFHRLSSNTFDND
AcMRJP4      AVNATTGKG GLASLAVQAMD S---VNTMVY MADNKDDALIVYQNA DDFS HRLSSHSI SNHN
                *:*:*  * * .*****: *  :*:***:***:..*  *:***:***:***:***: : :

AcMRJP2      PRYAKMTINGESFTLKNG-ICG MALSPVTNNLYYSPLASHGLYV NTEPFMKSQFGDN NN
AcMRJP3      PRYTKLTVAGESFTVKNG-ICG IALSPVTNNLYYSPLASHSLYV NTEQFRNPQYEENN-
AcMRJP1      PKFTKMTINGESFTTQSG-ISG MALSPMTNNLYYSPVASTSLYV NTEQFRTSNYEQNA-
AcMRJP5      PKYIKMMAAGESFTLQDG-IFG MALSPMTNNLYYSPLASRSLYV NTKPFMKSQYGTNN-
AcMRJP6      PRYSELTVAGESFTVHDG-IFG MALSPVTNNLYYSPLTSHSLYV NTEPFMKSQYGENN-
AcMRJP4      FRSDKMSQENLTLKEVDNRVFGMALSSVTHNLYYSPLSSQNLYV NTS LMNSQNQGN D-
                :  :  .  :  :  :  :  *:*:*:*:*:*:*:*:*  *  *****  :  :  *

AcMRJP2      VQYEGSQDTLNTQSLAKAVSKDGVLFVGLVGN S ALGCLNEHQPLQRENLELVAQNEKTLQ
AcMRJP3      VQYEGSQDILNTQSF AKAVSKNGV VFLGLVSNSTVGCVNEHQVLQKENFDVVAQNEETLQ
AcMRJP1      VHYEGVQNILDTQSSAKV VSKSGV LFFGLVGD S ALGCWNEHRS LERHNI RTVAQSD ETLQ
AcMRJP5      VQHEGVQDIFNTQSI AKIMSKNGV LFFGLMNNSAIGCWNEHQPLQRQNMDMVAQNEETLQ
AcMRJP6      IQYEGIQDIFNTQSSAKVMSKNGV LFFGLVNNSAIGCWNEHQPLQKQNM DMVAQNEETLQ
AcMRJP4      VQYESVQDVFSSQLSAKAVSKNGV LFFGFTNN-TLGCWNEHQSLDRQN IDIVARN-ETLQ
                :*: *  :*: *  **  **:*:*:*:*:*  :  :  :*: *  **  **:*:*:*
    
```

(continued)

Figure 3.17 Alignment of deduced amino acid sequence of AcMRJP cDNA. Accession number of AcMRJP as follows: AcMRJP1, AF525776; AcMRJP2, AF525777; AcMRJP4, AY532368; AcMRJP5, AY532369. AcMRJP3 amino acid sequence obtained from D. Srisuparbh (2002). Conserve residues are indicated by asterisks.

Phylogenetic relationships between AcMRJPs and AmMRJPs families

Nucleotide and deduced amino acid sequences of AcMRJP4 (AY532368), AcMRJP5 (AY532369) and AcMRJP6 cDNA obtained from this study and sequence of AcMRJP1 (AF525776), AcMRJP2 (AF525777) and AcMRJP3 (Srisuparbh, 2002) cDNA previously reported were aligned with the sequence of AmMRJP1-AmMRJP8 cDNA and *A. mellifera* yellow-f protein cDNA retrieved from the GenBank database [AmMRJP1 (AF000633), AmMRJP2 (AF000632), AmMRJP3 (Z26318), AmMRJP4 (Z26319), AmMRJP5 (AF004842), AmMRJP6 (AY313893), AmMRJP7 (BK001420), AmMRJP8 (AY398690) and AmYellowP (Albert and Klaudiny, 2004)].

Genetic distances of each MRJP were calculated at both nucleotide and protein levels. The lowest and highest divergence at the nucleotide level was 0.0660 (AcMRJP1-AmMRJP1) and 0.5066 (AcMRJP3-AmMRJP5), respectively. At the protein level, the lowest divergence was 0.0990 (AcMRJP1-AmMRJP1) whereas the highest divergence was 0.8556 (AcMRJP3-AcMRJP5) (Table 3.4).

The original data was then bootstrapped 1000, and 500 times for nucleotide and protein data, respectively. Bootstrapped neighbor-joining trees were then constructed (Figure 3.18 and 3.19). Relationships at both nucleotide and protein levels of MRJPs indicated phylogenetically closer relationships between the same MRJPs families from different species rather than different families of MRJPs within the same species.

The identical trees were obtained from two types data either nucleotide and amino acid sequences. The same families from different species were grouped together. The result showed *A. mellifera* Yellow-f protein was a monophyletic group distant from MRJPs. The MRJP8 exhibited the earliest divergence within MRJPs gene families.

Table 3.4 Estimated genetic distance among MRJPs families of *A. cerana* (Ac), *A. mellifera* (Am) and *A. mellifera* yellow protein (YP) obtained from nucleotide (above diagonal) and deduced amino acid (below diagonal) sequences.

	Ac1	Ac2	Ac3	Ac4	Ac5	Ac6	Am1	Am2	Am3	Am4	Am5	Am6	Am7	Am8	YP
Ac1	-	0.2630	0.2872	0.3276	0.3530	0.2882	0.0660	0.2573	0.2846	0.3613	0.3731	0.2899	0.2736	0.3848	1.0910
Ac2	0.4504	-	0.2131	0.3068	0.3332	0.2010	0.2910	0.0875	0.2237	0.3365	0.3573	0.2086	0.2018	0.3624	1.1438
Ac3	0.4900	0.4032	-	0.3460	0.4817	0.1981	0.3103	0.2137	0.1075	0.3489	0.5066	0.1883	0.2422	0.3846	1.1233
Ac4	0.5599	0.5790	0.6351	-	0.3995	0.2555	0.3419	0.3009	0.3590	0.0978	0.4393	0.2673	0.2825	0.4083	1.1892
Ac5	0.6246	0.5709	0.8556	0.6969	-	0.2747	0.3706	0.3359	0.4326	0.4258	0.0966	0.2716	0.2609	0.4316	1.2190
Ac6	0.4988	0.3627	0.3236	0.4668	0.4612	-	0.3163	0.2228	0.2201	0.2822	0.2785	0.0687	0.2106	0.3442	1.1964
Am1	0.0990	0.4687	0.5183	0.5665	0.6319	0.5093	-	0.2817	0.3086	0.3797	0.3929	0.3136	0.2894	0.4028	1.0972
Am2	0.4462	0.1683	0.4341	0.5214	0.5398	0.3930	0.4518	-	0.2256	0.3350	0.3462	0.2307	0.2020	0.3714	1.1227
Am3	0.4670	0.3913	0.1760	0.6046	0.7936	0.3733	0.5032	0.4103	-	0.3472	0.4654	0.2185	0.2612	0.3888	1.1101
Am4	0.6253	0.6560	0.6549	0.1969	0.7371	0.5060	0.6432	0.6135	0.5939	-	0.4383	0.2863	0.3149	0.4168	1.1986
Am5	0.6348	0.6166	0.7548	0.7498	0.1898	0.4823	0.6580	0.5710	0.7121	0.7645	-	0.2687	0.2759	0.4461	1.2227
Am6	0.5032	0.3680	0.3217	0.4936	0.4667	0.1036	0.5228	0.4061	0.3748	0.5250	0.4835	-	0.2253	0.3568	1.2031
Am7	0.4586	0.4001	0.4447	0.5309	0.5100	0.4165	0.4988	0.3963	0.4625	0.5971	0.5275	0.4420	-	0.3426	1.1580
Am8	0.6059	0.6192	0.6578	0.7297	0.7360	0.6390	0.6111	0.6299	0.6554	0.7887	0.7667	0.6390	0.6313	-	1.2479
YP	2.0515	2.0080	1.9782	2.2652	2.2173	2.0515	2.0773	2.1309	1.8883	2.4147	2.1877	2.0515	2.3122	1.8326	-

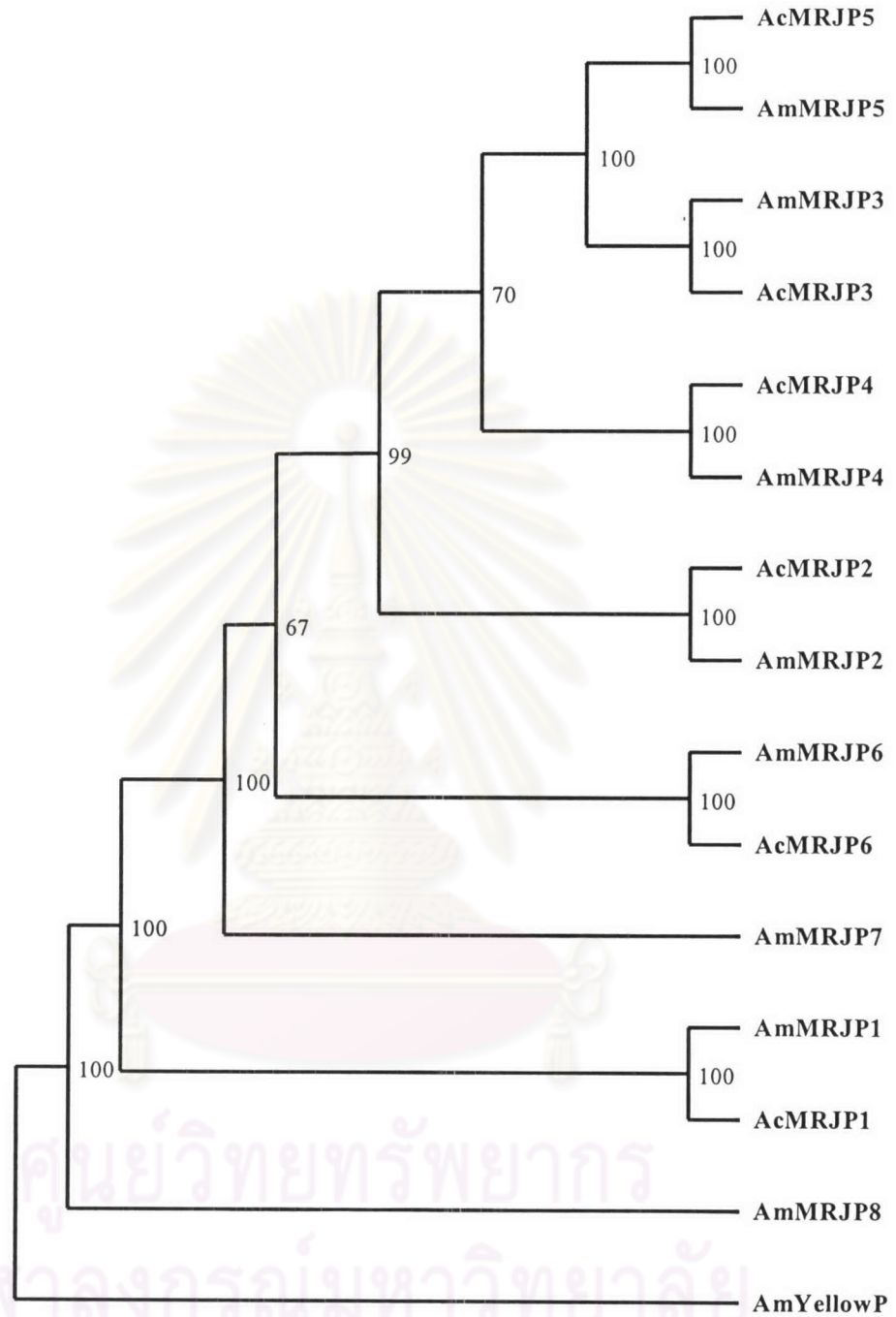


Figure 3.18 A bootstrapped tree illustrating relationship of MRJPs of *A. cerana*, *A. mellifera* and *A. mellifera* yellow protein. The original nucleotide sequence data was bootstrapped 1000 times. Values at the node indicate the percentage of times occurred out of 1000 trees.

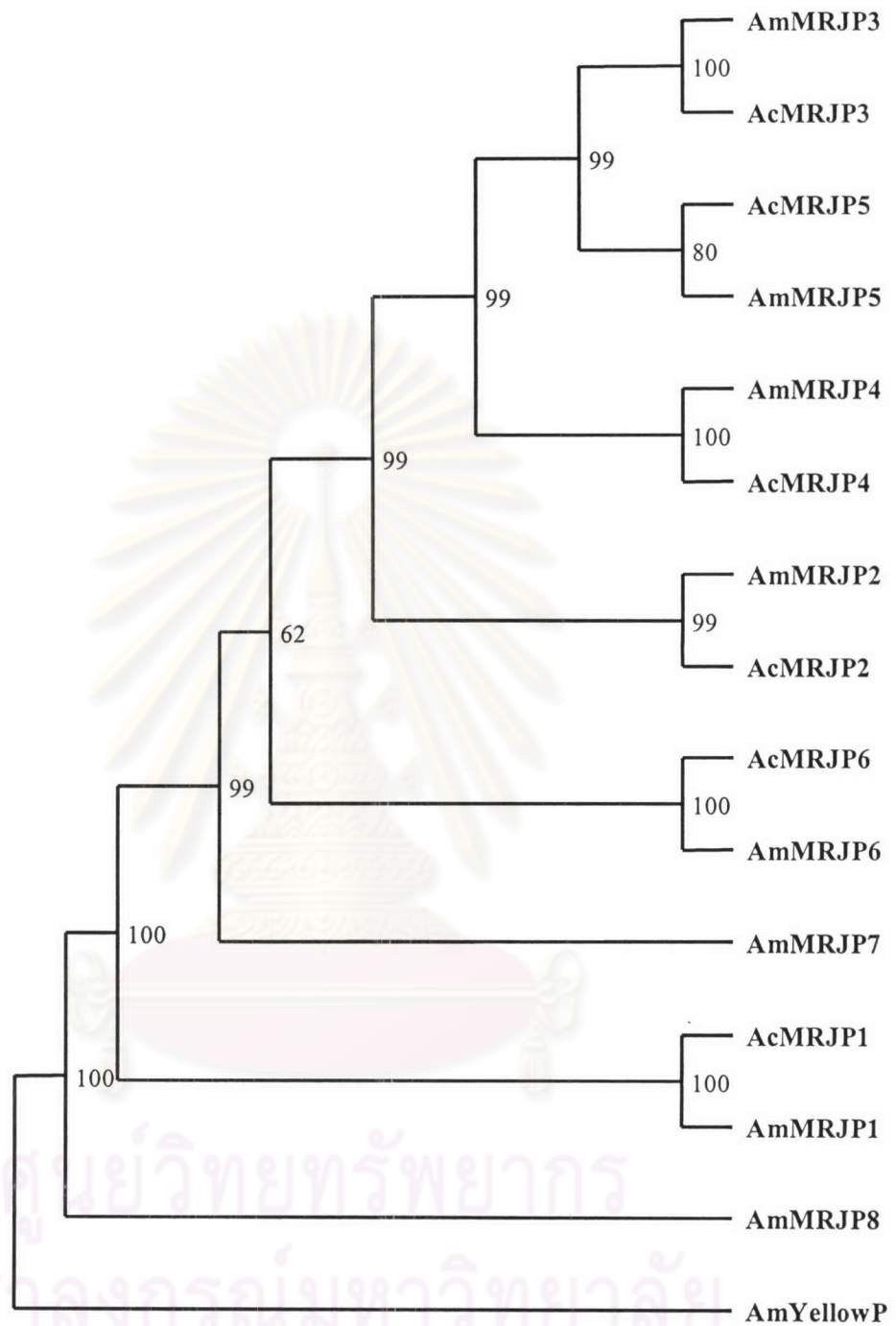


Figure 3.19 A bootstrapped tree illustrating relationship of MRJPs of *A. cerana*, *A. mellifera* and *A. mellifera* yellow protein. The original deduced amino acid sequence data was bootstrapped 500 times. Values at the node indicate the percentage of times occurred out of 500 trees.

3.2 Overexpression of AcMRJP4 protein in *E. coli*

In this study, The pET system was used for overexpression of AcMRJP4 protein in *E. coli* system. *E. coli* Rosetta (DE3) pLysS was selected for expression of the insert DNA, AcMRJP4 cDNA of pET 19b vector. The pET 19b contains T7 *lac* promoter, ampicillin resistance gene, *lacI* gene, sequence encoded a Histidine peptide (His-tags) at N-terminal of produced protein and sequence encoded protein sequences for enterokinase cleavage site. The His-tags was added for advantage in purification and identification of expressed protein while enterokinase cleavage site added for removed His-tags from the expressed protein.

E. coli Rosetta (DE3) pLysS contains T7 RNA polymerase gene and plasmid that harboured the rare tRNA genes, T7 lysozyme gene and chloramphenicol resistance gene.

For amplification of AcMRJP4 cDNA, primers were newly designed from AcMRJP4 cDNA sequence. The forward primers was designed over predicted N-terminal amino acid sequence without signal peptide and added *NdeI* restriction site to the 5'-end of the primer. The reverse primer was designed over stop codon and *BamHI* restriction site was added to the 5'-end of the primer. Recombinant plasmid contained AcMRJP4 cDNA from transformant number 5 (MRJP405) was used as template DNA for PCR amplification process. The *Pfu* DNA polymerase that have 3' to 5' exonuclease activity was used in PCR reaction. After amplification reaction was completed, amplified product was electrophoretically analyzed through 1 % agarose gel.

Only single PCR product of 1,400 bp was obtained (Figure 3.20). The PCR product was digested with proteinase K and purified by NucleoSpin® column. The pET 19b vector and purified PCR product were digested with *NdeI* and *BamHI* restriction

endonuclease. The digested products were electrophoretically analyzed and eluted from agarose gel by using QIAquick gel extraction kit. The *NdeI*-*Bam*HI digested PCR product was ligated with *NdeI*-*Bam*HI digested pET 19b vector, and then electro-transformed into *E. coli* JM109. Twelve white colonies containing the recombinant plasmid were randomly picked for plasmid extraction and double digested with *NdeI* and *Bam*HI. The result of electrophoretically analyzed show that all of twelve clones contained recombinant plasmid with AcMRJP4 cDNA fragment (Figure 3.21).

The transformed clone, transformant number 1 (clone name Exp401) was sent to Bioservice unit for plasmid extraction and sequencing. The T7 forward primer was used for sequencing. The amino acid sequence deduced from nucleotide sequence obtained showed that gene fragment had correct reading frame and 100% sequence identity with those of plasmid MRJP405 (Figure 3.22). The recombinant plasmid Exp401 and pET 19b vector were electro-transformed to a competent *E. coli* Rosetta (DE3) pLysS. The transformants that grown on selective plate containing ampicillin and chloramphenicol were expected to be *E. coli* Rosetta containing the recombinant plasmid. The recombinant clones were identified by colony PCR method. The recombinant clone number one that contained the inserted gene and one clone that carried pET 19b vector (Figure 3.23) were induced by IPTG for AcMRJP4 protein production. For culture cell harbouring the vector DNA (contained T7 *lac* promoter), IPTG final concentration of 1 mM was used to induce the protein production. The cell pellet was collected before induction with IPTG as a reference, and then collected after induced at 1 hour interval for 5 hours. The cell pellets were analyzed by SDS-PAGE. The results showed that *E. coli* Rosetta carried pET 19b vector and *E. coli* Rosetta carried the recombinant plasmid at 0 hours of induction, the

expected protein band that overexpressed was not observed. The protein band of 53 kDa was observed at 1-5 hours after induction. The highest expression level was 3 hours after induction with IPTG. The molecular weight of induced protein band was 53 kDa, which corresponded well to 55.7 kDa, the calculated molecular weight of recombinant AcMRJP4 protein deduced from DNA sequence (Figure 3.24).



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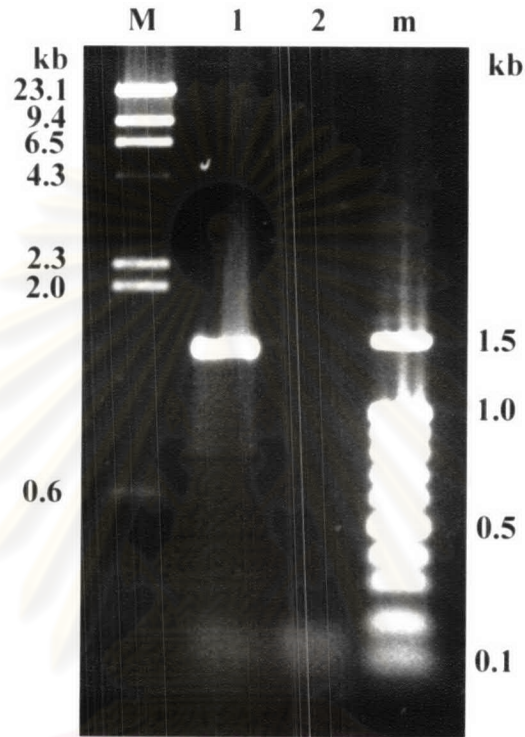


Figure 3.20 PCR amplification of AcMRJP4 cDNA without signal sq. for expression

Lane M = λ / *Hind* III standard molecular weight marker

Lane 1 = The amplification products of AcMRJP4 cDNA

Lane 2 = Negative control

Lane m = A 100 bp DNA ladder

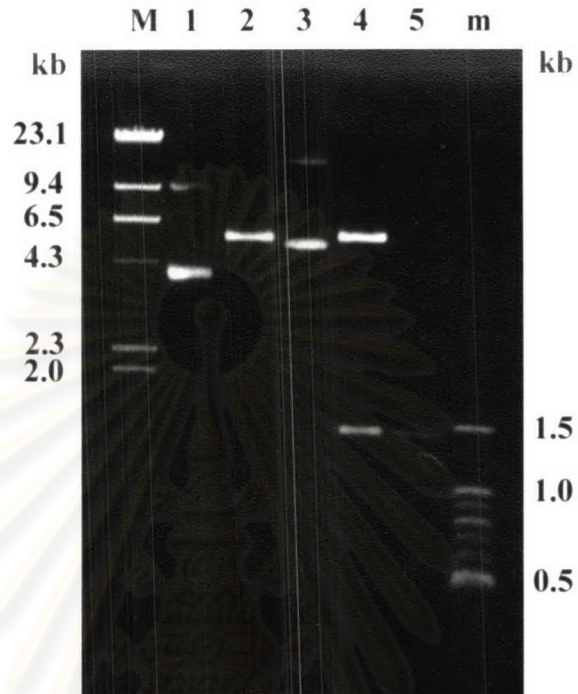


Figure 3.21 Cloning of pET19b expression vector containing AcMRJP4 cDNA in *E. coli*

JM 109

Lane M = λ / *Hind* III standard molecular weight marker

Lane 1 = undigested pET 19b

Lane 2 = pET 19b digested with *Nde* I and *Bam*HI

Lane 3 = undigested recombinant plasmid

Lane 4 = recombinant plasmid digested with *Nde* I and *Bam*HI

Lane 5 = AcMRJP4 insert digest with *Nde* I and *Bam*HI

Lane m = A 100 bp DNA ladder

CLUSTAL X (1.81) multiple sequence alignment

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MRJP405      ---MTKWLLMACPGIACQNIRGAVVRENSSRKKLTNTLNVIHEWKYVDYDFGSDEKRQ
Exp401      MGHHHHHHHHHSSGHIDDDDKHMAVVRENSSRKKLTNTLNVIHEWKYVDYDFGSDEKRQ
              :      :.  *  :: : *****

MRJP405      AAIQSGEYDRTKNYPLDVDQWHDKTFVTMLRYDGVPSLNVVSDKTGNGGPLLQPYPDWS
Exp401      AAIQSGEYDRTKNYPLDVDQWHDKTFVTMLRYDGVPSLNVVSDKTGNGGPLLQPYPDWS
              *****

MRJP405      FAKYEDCSGIVSANKIAIDEYERLWVLDSGLVNNIQPMCSPKLLAFDLTTSKLLKQVEIP
Exp401      FAKYEDCSGIVSANKIAIDEYERLWVLDSGL-----
              *****

MRJP405      HDVAVNATTGKGLASLAVQAMDSVNTMVYADNKKDALIVYQNADDSFHRLSSHISNHQ
Exp401      -----

MRJP405      FRSDKMSQENLTLKEVDNRVFGMALSSVTHNLYYSPLSSQNLYYVNTKSLMNSQNQGNDV
Exp401      -----

MRJP405      QYESVQDVFSSQLSAKAVSKNGVLFVFGFTNNTLGCWNEHQSLDRQNIDI VARNETLQMVV
Exp401      -----

MRJP405      GMKIKQNLPSGKVNNTQRNEHLLALTNKKQDVLNNDLNLEHVNFQILDANVNDLIRNSR
Exp401      -----

MRJP405      CANSDNQDNNQHNYNHNQVRHSSKSDNQNNQHNNQAYHSSKSDNWDNNNQAHSSKFD
Exp401      -----

MRJP405      NQNNNQYNN
Exp401      -----

```

Figure 3.22 Alignment of the deduced amino acid sequence of AcMRJP4 cDNA transformant number 5 (MRJP405) with deduced amino acid sequence of AcMRJP4 cDNA in pET 19b vector transformant number 1 (Exp401). Conserved residues are indicated in asterisks. : means amino acid which have the same group of side chains and similar size while . means amino acid which have the same group of side chains but different size.

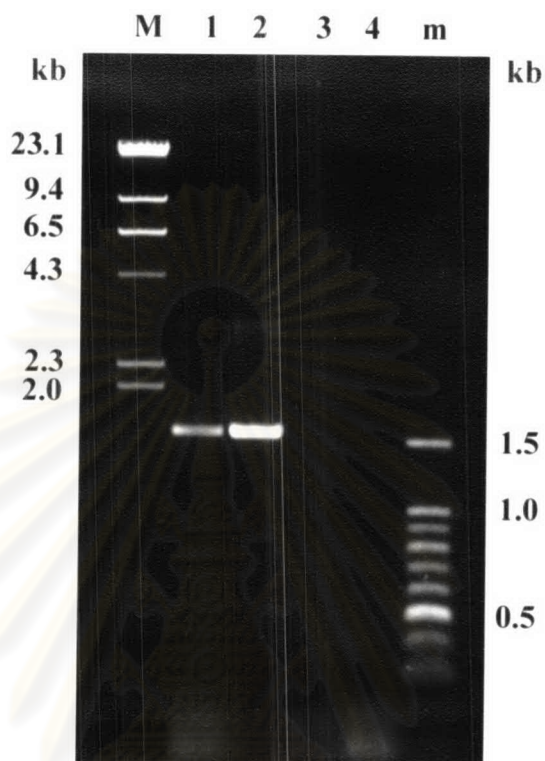


Figure 3.23 Colony PCR for identified cloning to expression host

Lane M = λ / *Hind* III standard molecular weight marker

Lane 1 = Positive control 5 ng of vector containing AcMRJP4 cDNA

Lane 2 = The amplification products of vector containing
AcMRJP4 cDNA in Rosetta host

Lane 3 = The amplification products of vector in Rosetta host

Lane 4 = Negative control

Lane m = A 100 bp DNA ladder

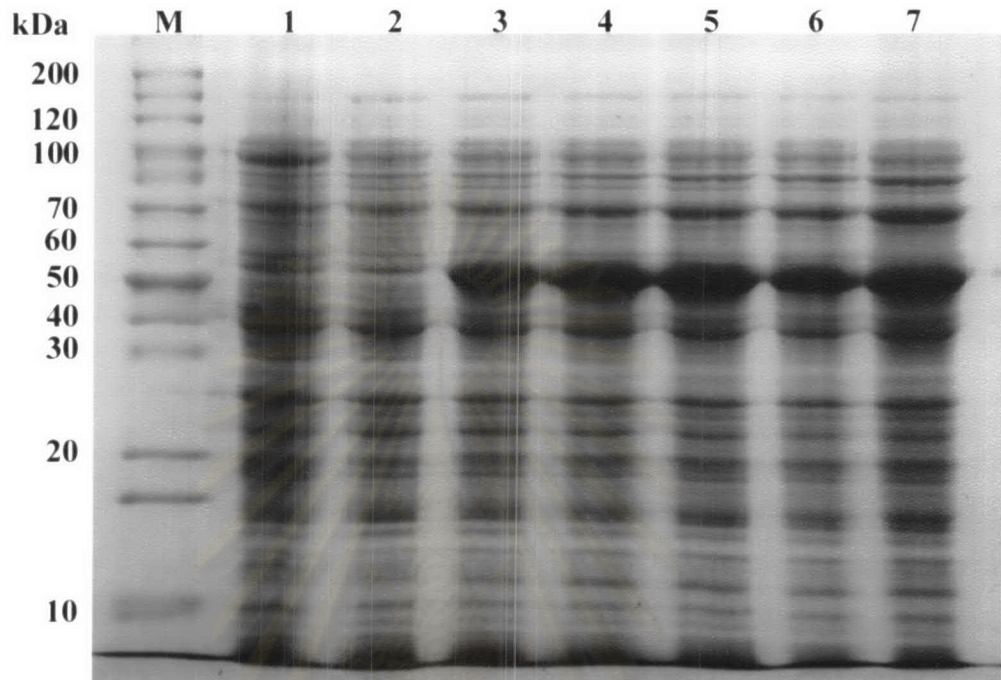


Figure 3.24 Protein pattern of crude extract of AcMRJP4 protein producing transformant vary in induction time.

Lane M Protein molecular weight marker

Lane 1 *E. coli* Rosetta cells carried pET 19b vector at 0 hours induction.

Lane 2 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 0 hours induction.

Lane 3 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 1 hour induction.

Lane 4 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 2 hours induction.

Lane 5 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 3 hours induction.

Lane 6 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 4 hours induction.

Lane 7 *E. coli* Rosetta cells carried recombinant AcMRJP4 plasmid at 5 hours induction.