

## References

- Avise, J.C., Arnold, J., Ball, R.M., Bermingham, E., Lamb, T., Neigel, J.E., Reeb, C.A., and Saunders, N.C. 1987. Intraspecific phylogeography: The mitochondrial DNA bridge between population genetics and systematics. Ann. Rev. Ecol. 18: 489-522.
- Bai, Y., Michaels, T.E., and Pauls, K.P. 1997. Identification of RAPD markers linked to common bacterial blight resistance genes in *Phaseolus vulgaris* L. Genome 40: 544-551.
- Bharadwaj, R.K. 1968. A new record of the mite *Tropilaelaps clareae* from *Apis dorsata* colonies. Bee wld. 49(4): 115-118.
- Biasiolo, A. 1992. Lack of allozyme variability among *Varroa* mite populations. Exp. Appl. Acarol. 16(4):287-294.
- Boyc, W., Elliott, L., Claek, R., and Jessup, D. 1990. Morphometric analysis of *Psoroptes* spp. mites from bighorn sheep mule deer cattle and rabbits. J. Parasitol. 76(6): 823-828.
- Bruce, W.A., Delfinado-Baker, M. and Vincent, D.L. 1997. Comparative morphology of the peritremes of *Varroa* and *Euvarroa* (Varroidae), parasites of honey bees (Apidae). Internat. J. Acarol. 23(1): 13-20.
- Burgett, M., Akwatanakul, P. and Morse, R.A. 1983. *Tropilaelaps clareae* : A parasite of honey bees in South-East Asia. Bee Wld. 64(1): 25-28.
- Carlson, J.E., Tulsieram, L.K., Glaubitz, V.W.K., Kauffeldt, C., and Rutledge, R. 1991. Segregation of random amplified DNA markers in F1 progeny of conifers. Theor. Appl. Genet. 83: 194-200.
- Crozier, R.H. and Crozier, Y.G. 1993. The mitochondrial genome of the honeybee *Apis mellifera*: Complete sequence and genome organization. Genet. Soc. Am. 133: 97-117.
- Davis, L.G., Battey, J.F. and Kuehl, W.M. 1994. Basic methods in molecular biology. 2<sup>nd</sup> ed. Connecticut: Appleton & Lange. pp. 113-178.
- De Jong, D. 1990. Mites: varroa and other parasite of brood. In Morse, R.A., Nowogrodzki, R. (eds.), Honey bee pests, predators and disease. Cornell University Press; Ithaca, NY, USA; pp. 201-218.

- Delfinado-Baker, M. and Baker, E.W. 1982. A new species of *Tropilaelaps* parasitic on honey bees. Am. Bee J. 122(6): 416-417.
- Delfinado-Baker, M. and Peng, C.Y.S. 1995. *Varroa jacobsoni* and *Tropilaelaps clareae*: A perspective of life history and why Asian bee-mites preferred European honey bees. Am. Bee J. 135(7): 415-420.
- Delfinado-Baker, M., Rath, W., and Boecking, O. 1992. Phoretic bee mites and honeybee grooming behavior. Internat. J. Acarol. 18(4): 315-322.
- Delfinado-Baker, M., Underwood, B.A., and Baker, E.W. 1985. The occurrence of *Tropilaelaps* mites in brood nests of *Apis dorsata* and *A. laboriosa* in Nepal, with descriptions of the nymphal stages. Am. Bee J. 135(10): 703-706.
- de Guzman, L.I., Rinderer, T.E., and Stelzer, J.A. 1997. DNA Evidence of the origin of *Varroa jacobsoni* oudemans in the Americas. Biochem. Genet. 35(9/10): 327-335.
- Donnell, K.O. 1992. Ribosomal DNA internal transcribed spacers are highly divergent in the phytopathogenic ascomycete *Fusarium sambucinum* (*Gibberella pulicaris*). Curr. Genet. 22: 213-220.
- Edwards, O.R., Melo, E.L., Smith, L., and Hoy, M.A. 1997. Discrimination of three *Typhlodromalus* species (Acari: Phytoseiidae) using random amplified polymorphic DNA markers. Exp. Appl. Acarol. 22(2): 101-109.
- Eickworth, G.C. 1988. The origins of mites associated with honey bees. In Needham, G.R., Page, R.E., Jr, Delfinado-Baker, M., and Bowman, C.E. (eds.), Africanized honey bees and bee mites. Ellis Hoewood Limited pp. 327-338.
- Eickworth, G.C. 1990. Mites: An Overview. In Morse, R.A. and Nowogrodzki, R. (eds.), Honey Bee Pests, Predators, and Diseases. London pp.188-199
- Eisses, K.T., Davies, S.L. and Chambers, G.K. 1994. Substrate and inhibitor specificities of the thermostable alcohol dehydrogenase allozymes ADH-71k and ADH-FCh.D. of *Drosophila melanogaster* Biochem. Genet. 32:91-103.
- Ellsworth, D.L., Rittenhouse, K.D. and Honeycutt, P.L. 1993. Artifactual variation in randomly amplified polymorphic DNA banding patterns. BioTechniques 14: 214-217.

- Fenton, B., Malloch, G. and Moxey, E. 1997. Analysis of eriophyid mite rDNA internal transcribed spacer sequences reveals variable simple sequence repeats. Insect Mol. Biol. 6(1):23-32.
- Garcia, G.M., Stalker, H.T., Shroeder, E. and Kochert, G. 1996. Identification of RAPD, SCAR, and RFLP markers tightly linked to nematode resistance genes introgressed from *Arachis cardenasii* into *Arachis hypogaea*. Genome 39: 836-845.
- Goka, K., Takafuji, A., Hamamura, T., Osakabe, M. and Komazaki, S. 1996. Genetic distinctness between two forms of *Tetranychus urticae* Koch (Acari: Tetranychidae) detected by electrophoresis. Exp. Appl. Acarol. 20(12): 683-693.
- Hadrys, H., Balick, M. and Schierwater, B. 1992. Applications of random amplified polymorphic DNA (RAPD) in molecular ecology. Mol. Ecol. 1: 55-63.
- Hall, H.G. and Smith, D.R. 1991. Distinguishing African and European honeybee matrilineages using amplified mitochondrial DNA. Proc. Natl. Acad. Sci. USA. 88: 4548-4552.
- Hoelzled, A.R. and Green, A. 1992. Analysis of population-level variation by sequencing PCR-amplified DNA. In Hoelzled, A.R. (ed.), Molecular Genetic Analysis of populations: A practical Approach. IRL Press. Oxford, pp. 159-188.
- Hoy, M.A. 1994. Insect molecular genetics an introduction to principles and applications. London: Academic Press. pp. 546.
- Hsiao, C., Chatterton, N.J., Asay, K.H. and Jensen, K.B. 1995. Phylogenetic relationships of the monogenomic species of the wheat tribe, *Triticeae* (poaceae), inferred from nuclear rDNA (internal transcribed spacer) sequences. Genome 38: 211-223.
- Hsiao, C., Chatterton, N.J., Asay, K.H. and Jensen, K.B. 1994. Phylogenetic relationships of 10 grass species: an assessment of phylogenetic utility of the internal transcribed spacer region in nuclear ribosomal DNA in monocots. Genome 37(1):112-120.

ต้นฉบับ หน้าขาดหาย

- Marillia, E.F. and Scoles, G.J. 1996. The use of RAPD markers in *Hordeum* phylogeny. Genome 39:646-654.
- Matheson, A. 1993. World bee health report. Bee wld. 74(4): 177-213.
- May, B. 1992. Starch gel electrophoresis of allozymes. In Hoelzel, A.R. (ed.), Molecular Genetic Analysis of Population: A Practical Approach. IRL Press, Oxford, pp. 1-27.
- McLain, D.K., Wesson, D.M., Olover, J.H., JR. and Collins, F.H. 1995. Variation in Ribosomal DNA Internal Transcribed Spacers I Among Eastern Populations of *Ixodes scapularis* (Acari: Ixodidae). Ent. Soc. Am. 32(3): 353-360.
- Meunier, J.R. and Grimont, P.A.D. 1993. Factors affecting reproducibility of random amplified polymorphic DNA fingerprinting. Res. Microbio. 144: 373-379.
- Moritz, R.F.A., Cornuet, J.M., Kryger, P., Garnery, L. and Hepburn, H.R. 1994. Mitochondrial DNA variability in South African honeybees (*Apis mellifera*). Apidologie 25: 169-178.
- Narang, S.K., Leopold, R.A., Kraeger, C.M. and DeVault, J.D. 1994. Amplifications of genetic to arthropods. London: CRC Press Inc.
- Navajas, M., Cotton, D., Kreiter, S. and Gutierrez, J. 1992. Molecular approach in spider mites (Acari: Tetranychidae): preliminary data on ribosomal DNA sequences. Exp. Appl. Acarol. 15: 211-218.
- Navajas, M., Fournier, D., Lagnel, J., Gutierrez, J. and Boursot, P. 1996. Mitochondrial COI sequences in mites: evidence for variations in base composition. Insect Mol. Biol. 5(4): 281-285.
- Navajas, M., Gutierrez, J., Bonato, O., Bolland, H.R. and Mmapangou-Divassa, S. 1994. Intraspecific diversity of the Cassava Green Mite *Mononychellus progresivus* (Acari: Tetranychidae) using comparisons of mitochondrial and nuclear ribosomal DNA sequences and cross-breeding. Exp. Appl. Acarol. 18:351-360.
- Norris, D.E., Klompen, J.S.H., Keirans, J.E. and Blackiv, W. 1996. Population genetic of *Ixodes scapularis* (Acari: Ixodidae) based on mitochondrial 16S and 12S genes. Ent. Soc. Am. 33(1): 78-89.

- Osakabe, Mh. and Sakagami, Y. 1994. RFLP analysis of ribosomal DNA in sibling species of spider mite, genus *Panonychus* (Acari: Tetranychidae). Insect Mol. Biol. 3(1): 63-66.
- Paskewitz, S.M., NG, K., Coetzee, M. and Hunt, R.H 1993. Evaluation of the Polymerase Chain Reaction Method for Identifying Members of the *Anopheles gambiae* (Diptera: Culicidae) Complex in Southern Africa. Ent. Soc. Am. 3(5): 953-957.
- Paskewitz, S.M., Wessont, D.M. and Collins, F.H. 1993. The internal transcribed spacers of ribosomal DNA in five members of the *Anopheles gambiae* species complex. Insect Mol. Biol. 2(4): 247-257.
- Porter, C.H. and Collins F.H. 1991. Species-diagnostic differences in a ribosomal DNA internal transcribed spacer from the sibling species *Anopheles freeborni* and *Anopheles hermsi* (Diptera: Culicidae). Am. J. Trop. Med. Hyg. 45(2): 271-279.
- Promega. 1996. Technical manual for OmniBase DNA cycle sequencing system. Promega: Medison.
- Rao, K.B., Bhat, K.V., and Totey, S.M. 1996. Detection of species-specific genetic markers in farm animals through random amplified polymorphic DNA (RAPD). Genet. Anal. 13(5): 135-138.
- Rath, W., Boecking, O. and Drescher, W. 1994. The phenomena of simultaneous infestation of *Apis mellifera* in Asia with the parasitic mites *Varroa jacobsoni* OUD. And *Tropilaelaps clareae* Delfinado & Baker. J. Apic. Res. 34(4):125-127.
- Richner, S.M., Meiring, J., and Kirby, R. 1997. A study of the genetic diversity of *Mycobacterium tuberculosis* from patients in the eastern province of South Africa random amplified polymorphic DNA profiling. Electrophoresis 18(9): 1570-1576.
- Rijpkema, S., Golubic, D., Molkenboer, M., Verbeek-De Kurif, N., and Schellekens, J. 1996. Identification of four genomic groups of *Borrelia burgdorferi sensu lato* in *Ixodes ricinus* ticks collected in a Lyme borreliosis endemic region of northern Croatia. Exp. Appl. Acarol. 20:23-30.

- Rinderer, T.E. 1986. Selection. In Rinderer, T.E. (ed.), Bee genetic and Breeding. Orlando: Academic Press. pp. 23-30.
- Ruano, G., Kidd, K.K., and Stephens, J.C. 1990. Haplotype of multiple polymorphisms resolved by enzymatic amplification of single DNA molecules. Proc. Natl. Acad. Sci. USA 87(16): 6296-6300.
- Saiki, R.K., Gelfand, D.H., Stoffel, S., Scharf, S.J., Higuchi, R., Horn, G.T., Mullis, K.B. and Erlich, H.A. 1988. Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. Nature (London) 239:487-497.
- Sanger, F., Nickle, S and Coulson, A.R. 1977. Sequencing with chain-terminating inhibitors. Proc. Natl. Sci. USA. 74:5463-5467.
- Sappal, N.P., Jeng, R.J., Hubbes, M. and Liu, F. 1995. Restriction fragment length polymorphisms in polymerase chain reaction amplified ribosomal DNAs of three *Trichogramma* (Hymenoptera: Trichogrammatidae) species. Genome 38:419-425.
- Scott, J.A., Brogdon, W.G. and Collins, F.H. 1993. Identification of single specimens of the *Anopheles gambiae* complex by the polymerase chain reaction. Am. J. Trop. Med. Hyg. 49(4): 520-529.
- Scot, M.P. and Williams, S.W. 1993. Comparative reproductive success of communally breeding burying beetles as assessed by PCR with randomly amplified polymorphic DNA. Proc. Natl. Acad. Sci. 90: 2242-2245.
- Scot, M.P. Haymes, K.M. and Williams, S.C. 1992. Parentage analysis using RAPD PCR. Nucleic Acids Res. 20: 54-93.
- Shankaranarayanan, P., Banerjee, M., Kacker, R.K., Aggarwal, R.K. and Singh, L. 1997. Genetic variation in Asiatic lions and Indian tigers. Electrophoresis 18(9): 1693-1700.
- Smith, D.R. and Brown, W.M. 1988. Polymorphisms in mitochondrial DNA of European and Africanized honeybee (*Apis mellifera*). Experientia 44: 257-260.
- Swoboda, I. and Bhalla, P. L., 1997. RAPD analysis of genetic variation in Australian fan flower, *Scaevola*. Genome. 40: 600-606.

- Tang, J., Toe, L., Back, C. and Unnasch, T.R. 1996. Intra-specific Heterogeneity of the rDNA Internal Transcribed Spacer in the *Simulium damnosum* (Diptera: Simuliidae) Complex. Mol. Bio. Evol. 13(1): 244-252.
- Tassanakajon, A., Pongsomboon, S., Rimphanitchayakit, V., Jarayabhand, P. and Boonsaeng, V. 1997. Randomly amplified polymorphic DNA (RAPD) markers for determination of genetic variation in wild populations of the black tiger prawn (*Penaeus monodon*) in Thailand. Mol. Marine Biol. and Biotech. 6:110-115.
- Todd, C.D., Walker, A. M., Walff, K., Northcott, S. J., Walker, A. F., Ritchie, M. G., Hoskins, R., Abbott, R. J. and Hazon, N. 1997. Genetic differentiation of population of the copepod sea louse *Lepeophtheirus salmonis* (Kroyer) ectoparasitic of wild and farmed salmonids around the coasts of Scotland: Evidence from RAPD markers. J. Exp. Mar. Biol. and Ecol. 210:(2) 251-274.
- Vogler, A. P. and DeSalle, R. 1994. Evolution and Phylogenetic Information Content of the ITS-1 Region in the Tiger Beetle *Cicindela dorsalis*. Mol. Biol. Evol. 11(3): 393-405.
- Walsh, P.S., Metzger, D.A. and Higuchi, R. 1994. Chelex<sup>®</sup> 100 as a Medium for Simple Extraction of DNA for PCR-Based Typing from Forensic Material. BioTechniques 10(4): 506-513.
- Weising, K., Nybom, H., Wolff, K. and Meyer, W. 1995. DNA fingerprinting in plants and fungi. CRC Press, pp. 322.
- Welsh, J. and McClelland, M. 1990. Fingerprint genomes using PCR with arbitrary primers. Nucleic Acids Res. 18(24):7213-7218.
- Wesson, D.M., McLain, D.K., Oliver, J.H., Piesman, J. and Collins, F.H. 1993. Investigation of the validity of species status of *Ixodes dammini* (Acari: Ixodidae) using rDNA. Proc. Natl. Acad. Sci. USA 90: 10221-10225.
- White, T.J., Bruns, T., Lee, S. and Taylor, J. 1990. Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In Innis, M.A., Gelfand, D.H., Sninsky, J.J., and White. T.J. (eds.), PCR protocols: a guide to methods and applications. Academic Press, San Diego. pp. 315-322.

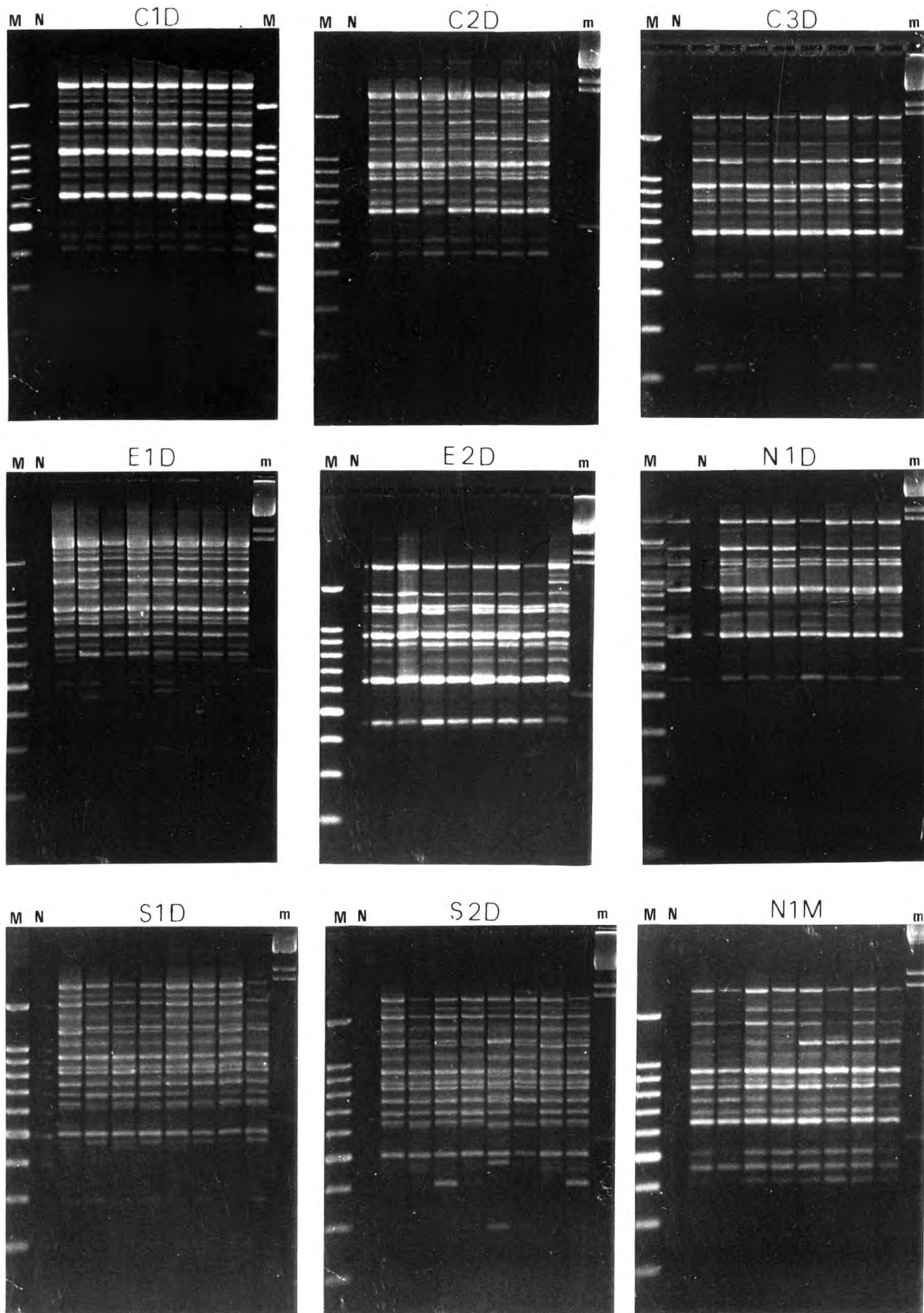


- Wilbur, W.J. and Lipman, D.J. 1983. Rapid similarity searches of nucleic acid and protein data banks. Proc. Natl. Acad. Sci. USA. 80: 726-730.
- Williams, J.G.K., Kanafey, M.K., Livak, Rafalski J.A. and Tingey, S.V. 1993. Genetic analysis using random amplified polymorphic DNA markers. Methods Enzymol 218: 704-740.
- Williams, J.G.K., Kubelik, A.R., Livak, K.J., Rafalski J.A. and Tingey, S.V. 1990. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. Nucleic Acids Res. 18(22): 6531-6535.
- Wolfe, S.L. 1993. Molecular and Cellular Biology. Wadsworth publishing Company, Belmont, California.
- Wongsiri, S. 1989. Biology of honey bees. 2<sup>nd</sup> ed. Bangkok: Ton-Or Co., LTD pp. 184.
- Wongsiri, S., Tangkanasing, P., Lekprayoon, C., Rinderer, T.E., and Sylvester, H.A. 1995. Biodiversity of parasitic mites of honey bees in Southeast Asia. pp. 6-13 In Wongsiri, S., Tangkanasing, P., Lekprayoon, C., Rinderer, T.E., Sylvester, H.A. and Delfinado Baker, M. (eds.) Biodiversity of bee mites and honey bees in Thailand. Publication of Bee Biology Research Unit, Chulalongkorn University (1991-1994).
- Zahler, M., Gothe, R. and Rinder, H. 1995. Diagnostic DNA amplification from individual tick eggs, laevae and nymphs. Exp. Appl. Acarol. 19: 731-736.
- Zavaleta, A.I., Martinez-Murcia, A.J. and Rodriguez-Valera, F. 1997. Intraspecific Genetic Diversity of *Oenococcus oeni* as Derived from DNA Fingerprinting and Sequence Analyses. Appl. and Environ. Microbiol. 63(4): 1261-1267.
- Zhuo, L., Sajdak S.L. and Phillips, R.B. 1994. Minimal intraspecific variation in the sequence of the transcribed spacer regions of the ribosomal DNA of lake trout (*Salvelinus namaycuch*). Genome 37(4): 664-671.

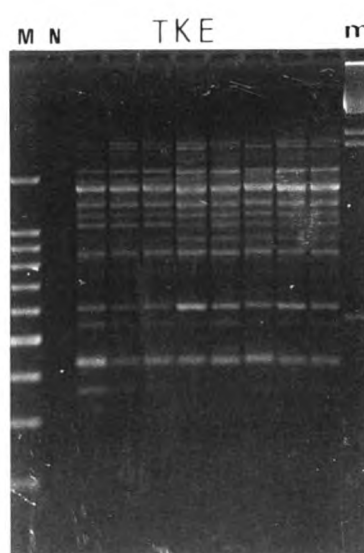
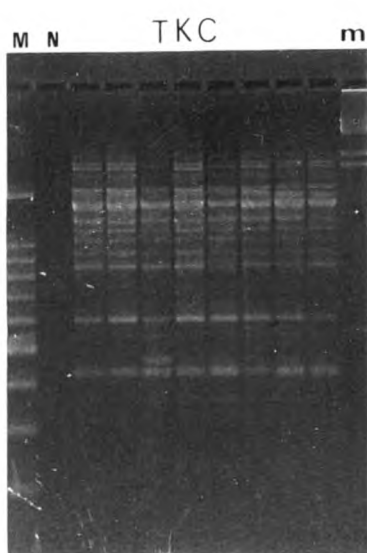
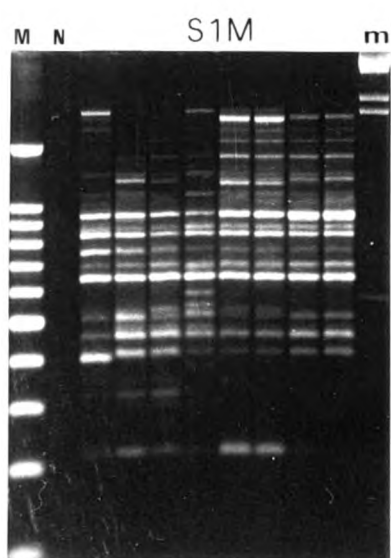
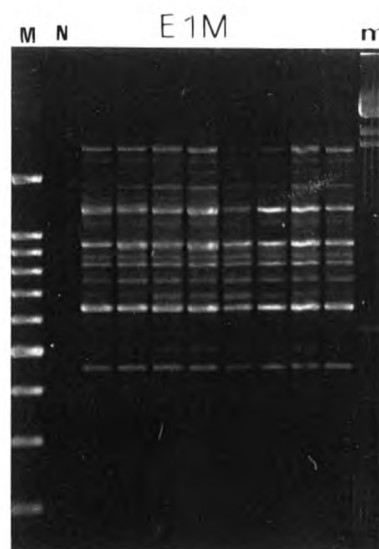
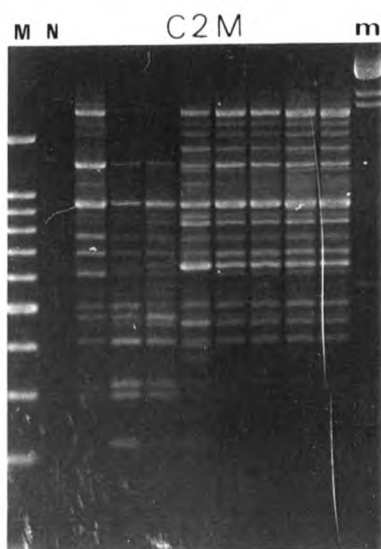
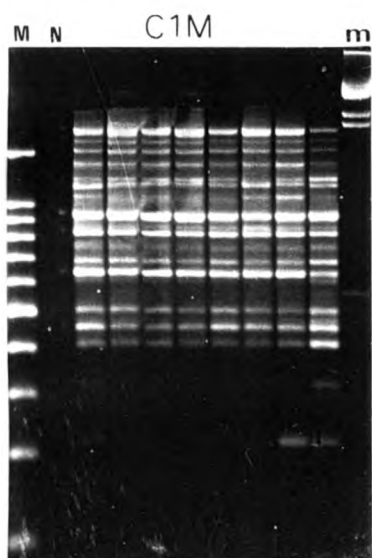
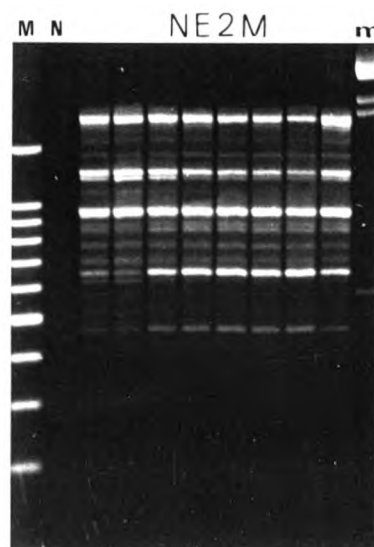
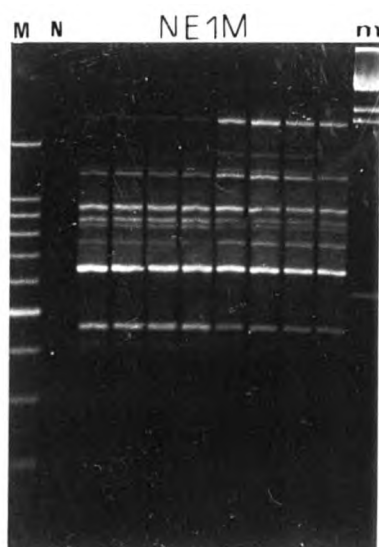
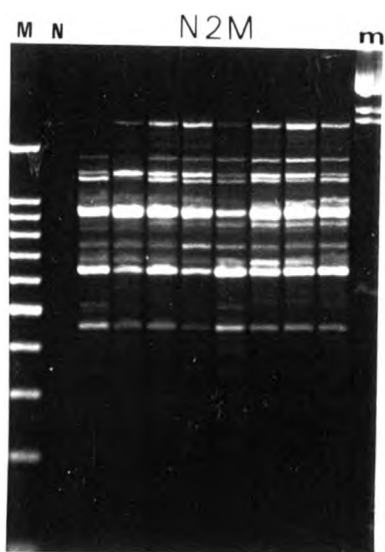
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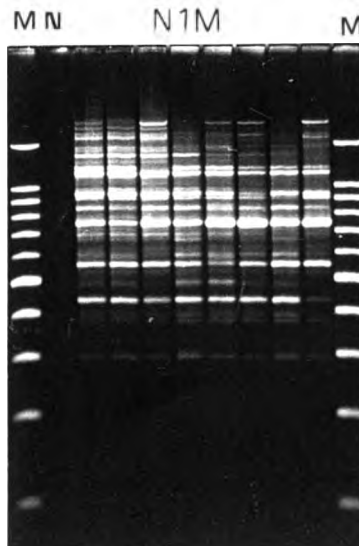
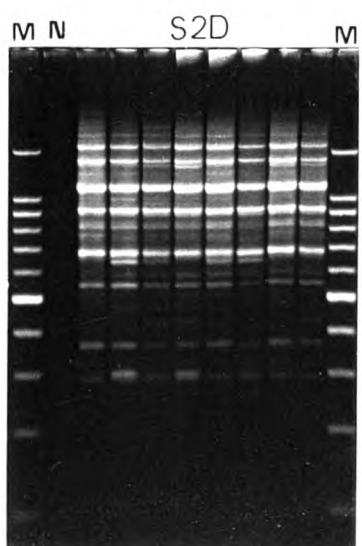
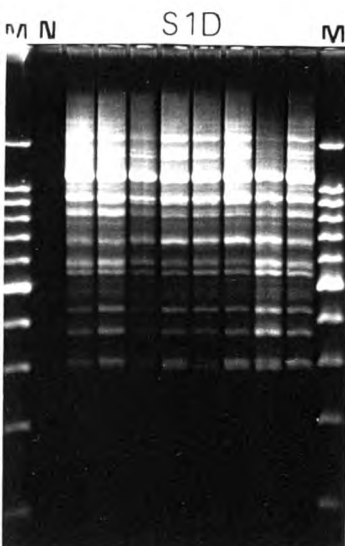
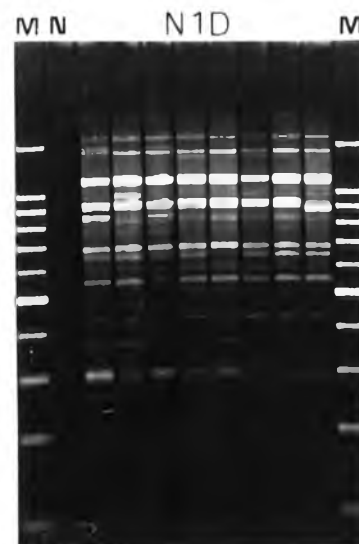
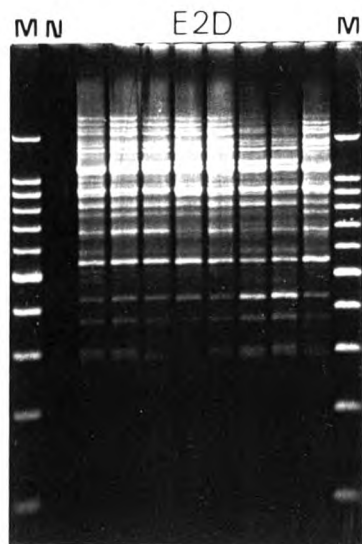
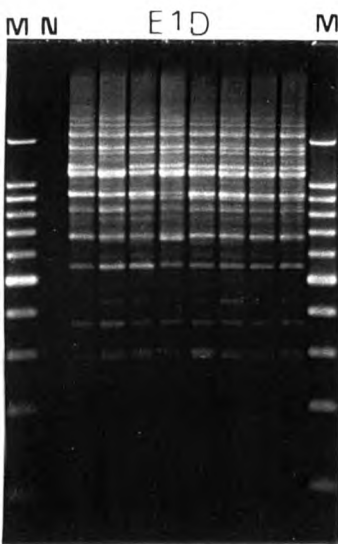
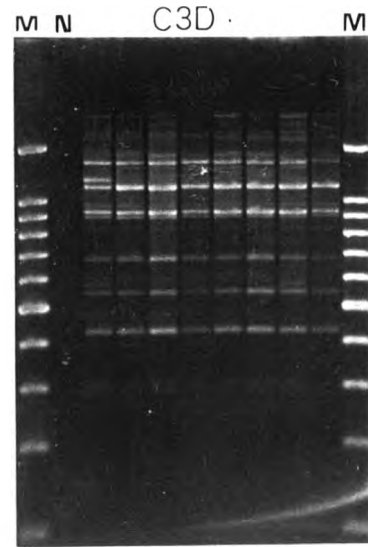
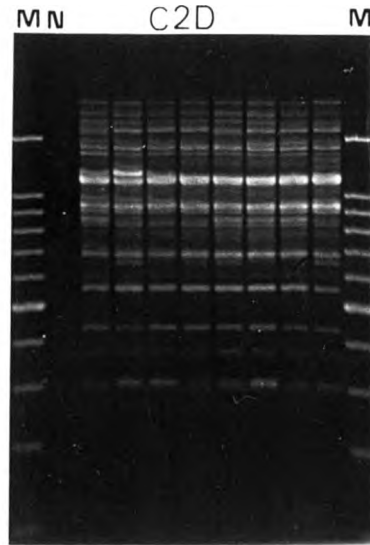
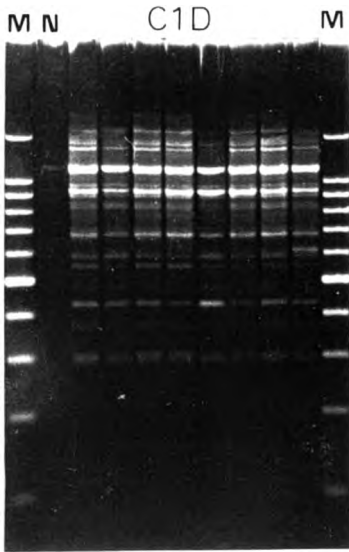
RAPD-PCR patterns using primer OPA07, OPA11 and OPA12 for each group of 16 groups of *T. clareae* and 2 groups of *T. koenigerum*.

Primer OPA07

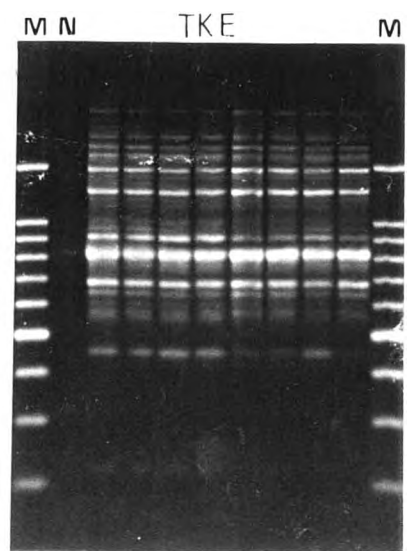
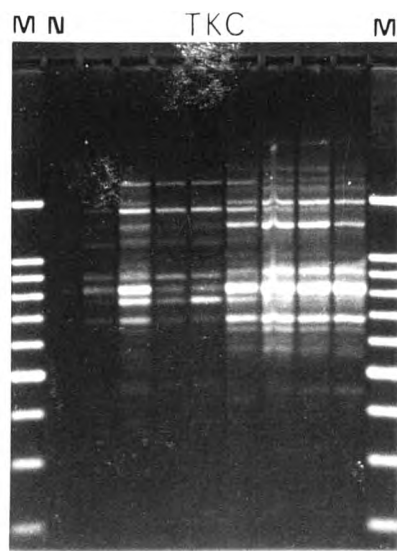
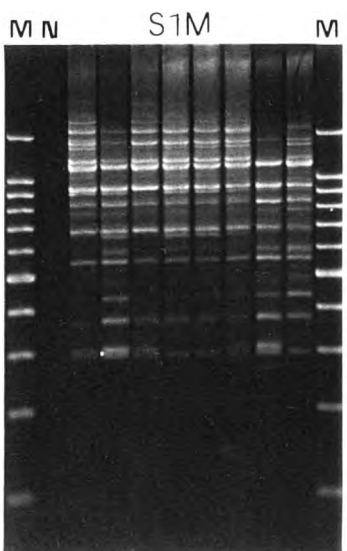
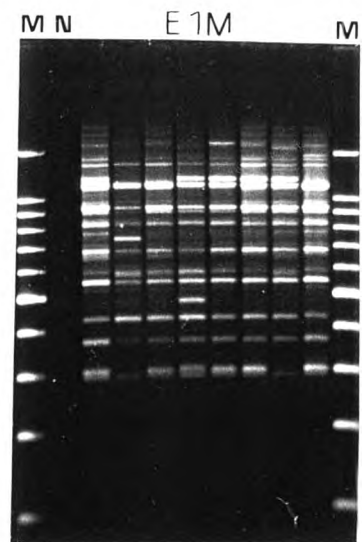
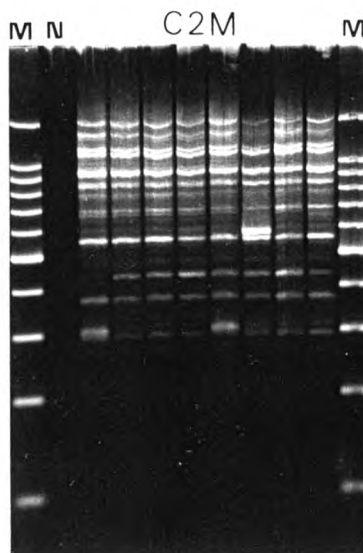
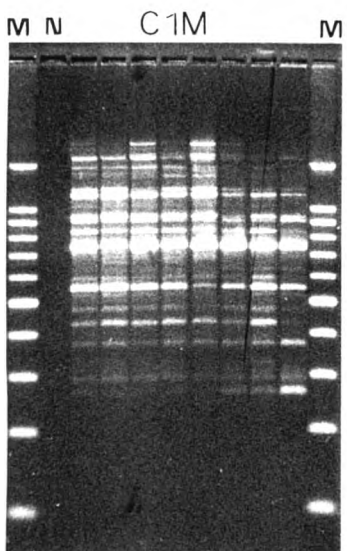
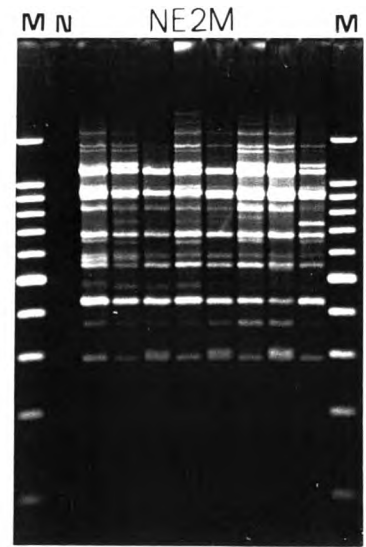
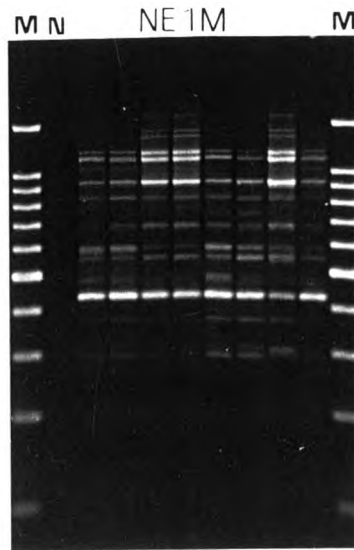
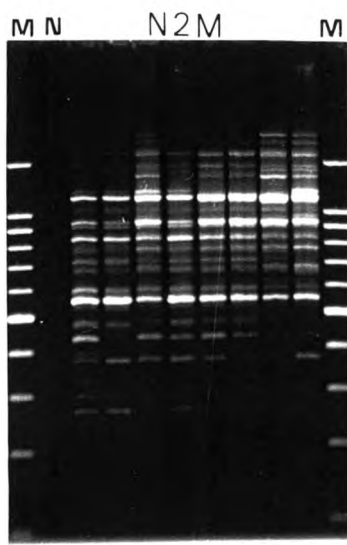


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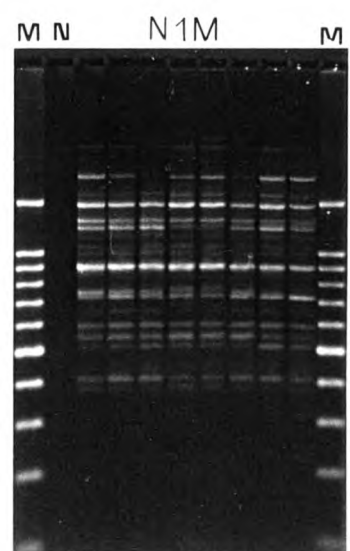
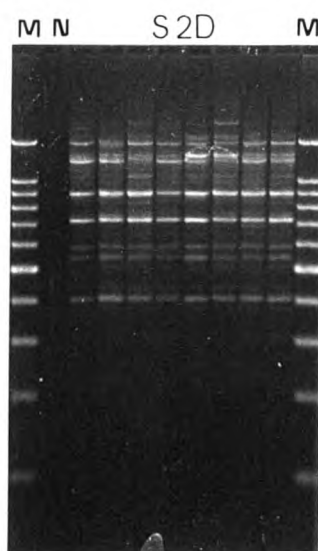
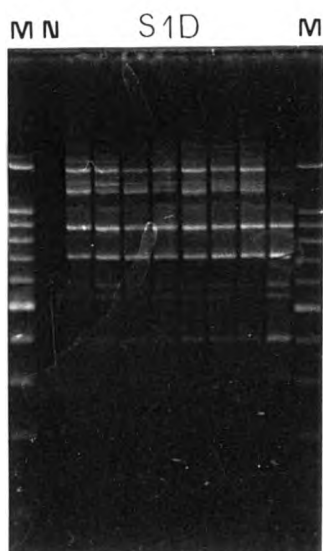
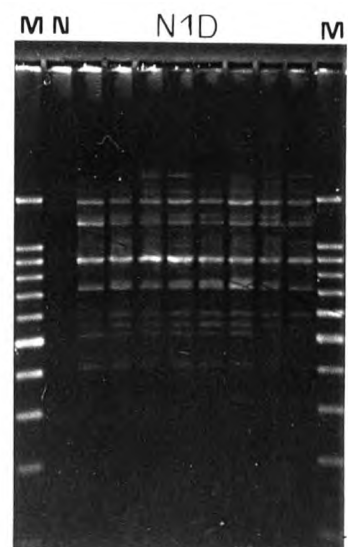
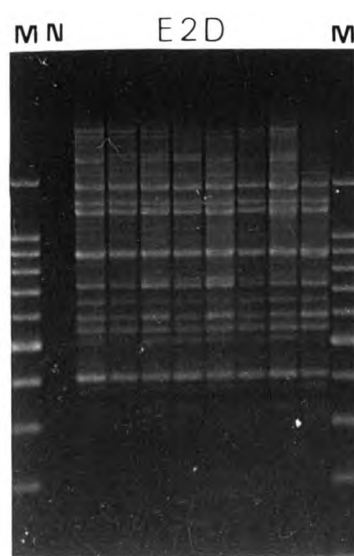
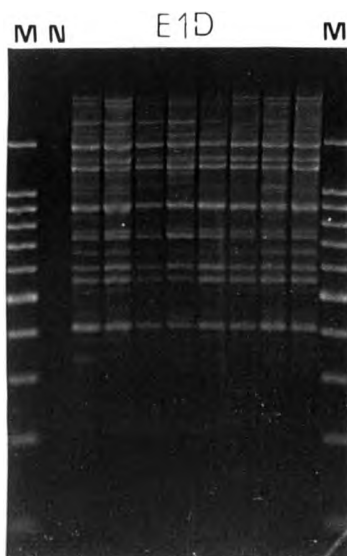
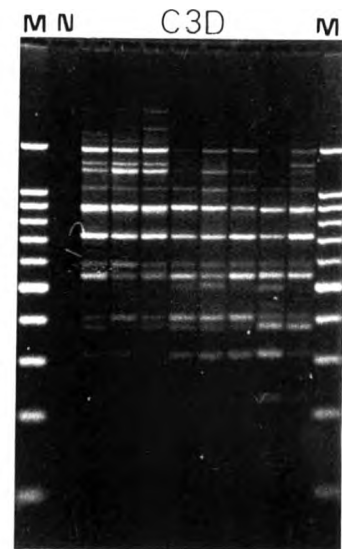
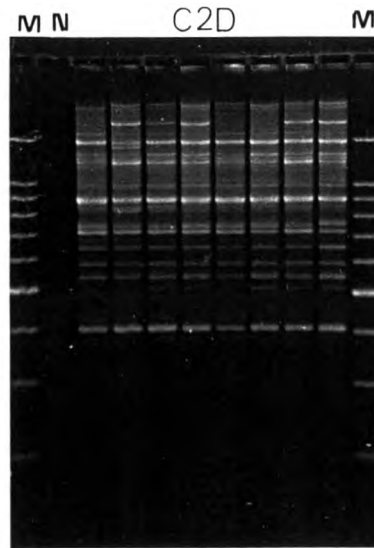
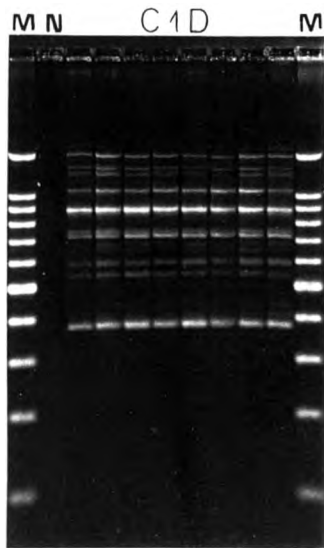


Primer OPA11

## Primer OPA11 (continue)

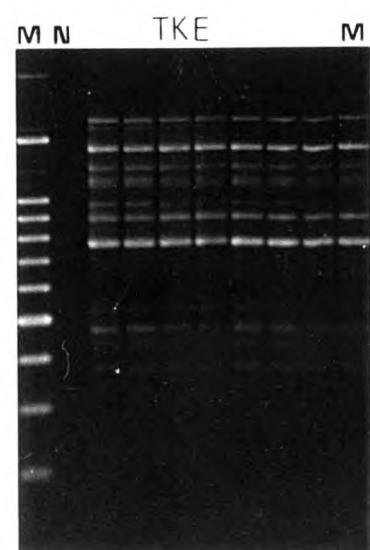
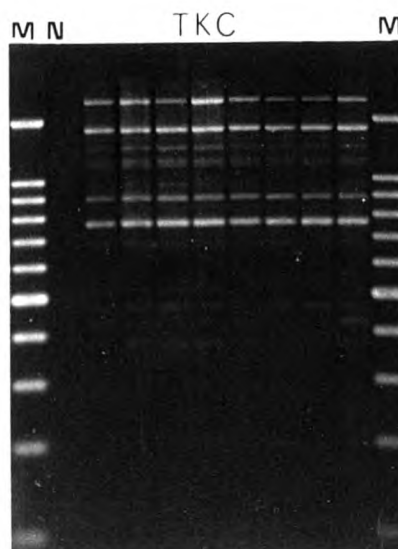
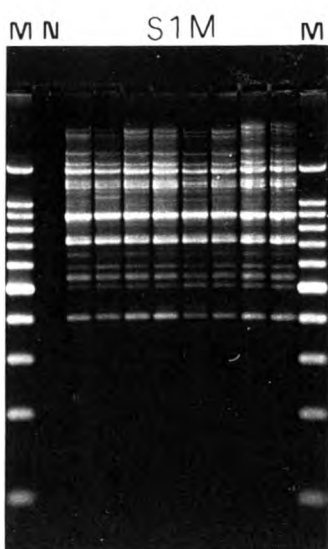
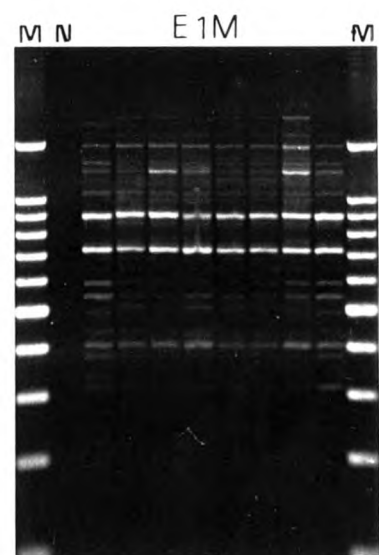
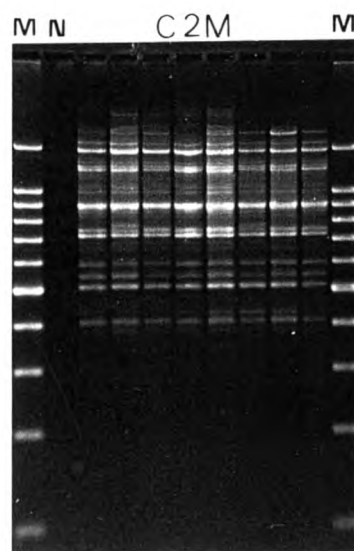
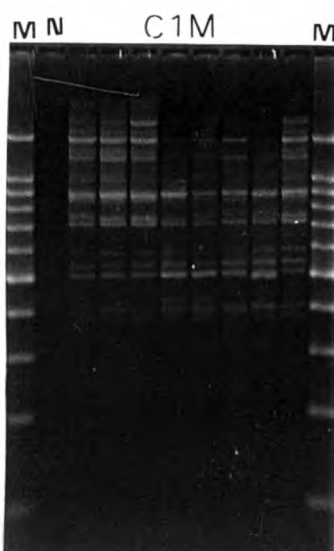
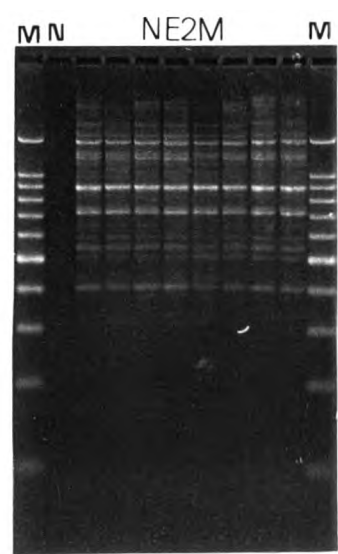
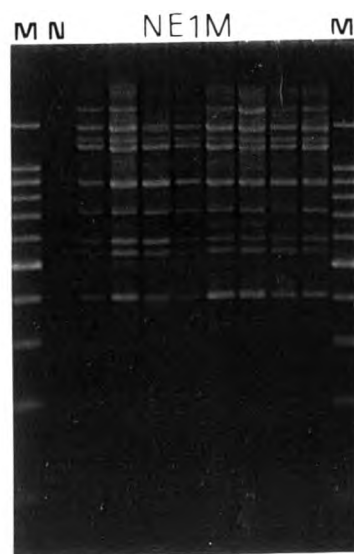
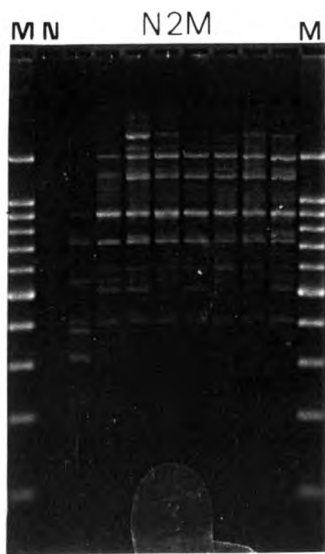


## Primer OPA12





## Primer OPA12 (continue)



Genotype for each primer in 16 sample groups of *T. clereae* and 2 of *T. koenigerum*.

OPA07 primer

size (bp)	Genotype																																					
	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	AA	AB	AC	AD	AE	AF	AG	AH	AI	AJ		
2040	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
2020	0	0	0	0	0	0	1	1	1	1	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0
2000	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1980	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1710	1	1	0	1	0	0	1	1	1	1	1	0	0	0	1	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1550	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1530	1	1	1	1	0	0	1	1	1	1	1	0	0	0	1	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1390	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1375	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1360	1	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1210	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1195	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1180	0	0	0	0	0	1	0	0	0	1	1	1	0	1	1	1	1	0	1	1	1	1	1	1	1	1	0	1	1	1	1	1	0	1	1	1	1	0
1160	0	0	0	0	0	0	0	1	0	0	0	0	1	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1110	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1070	1	1	1	1	1	0	0	1	1	1	1	1	1	1	1	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1050	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
950	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
880	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
850	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1	0	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
780	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	
760	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
960	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0	0	0	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
640	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1
620	0	0	0	0	0	0	1	0	0	0	1	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
610	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
590	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
550	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
510	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
480	0	0	0	0	0	0	1	0	1	1	1	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0	1	0	0	0	0	0	0	0	0	0
460	1	1	1	1	1	1	0	1	0	0	0	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1
430	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
410	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1
405	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0



OPA07 primer (Continue)

Size(bp)	Genotype																							
	AK	AL	AM	AN	AO	AP	AQ	AR	AS	AT	AU	AV	AW	AX	AY	AZ	BA	BB	BC	BD	BE	BF	BG	BH
2040	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
2020	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	1	0	0	0	0
2000	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
1980	1	1	1	1	1	1	0	1	1	0	1	1	0	1	0	0	0	0	1	1	1	0	0	0
1710	1	0	0	1	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	1	0	0	0
1550	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
1530	1	1	0	1	1	1	0	0	0	0	1	1	0	0	0	0	0	0	1	1	1	0	0	0
1390	1	1	1	1	1	1	1	0	1	1	1	1	0	1	0	1	1	1	1	1	1	0	0	0
1375	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
1360	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1210	1	0	0	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0
1995	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
1180	0	1	0	0	1	1	1	0	1	1	1	1	1	1	0	1	1	0	1	1	1	0	0	0
1160	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0
1110	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
1070	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0	0	0	0
1050	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
950	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1
880	1	1	1	1	1	0	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
850	1	1	1	1	1	1	1	0	0	0	1	1	1	1	1	1	1	1	1	1	1	0	0	0
780	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
760	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
690	1	1	1	1	1	1	1	1	1	1	1	1	0	0	1	1	1	1	1	1	1	0	0	0
640	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0
620	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0
610	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
590	0	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0
550	0	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	1	1	1
510	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	1	1	0	0	0
480	0	1	0	1	1	1	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1
460	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0
430	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
410	0	1	0	1	1	1	0	0	0	0	0	0	1	1	0	1	1	1	1	1	1	0	0	0
405	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0

OPA11 primer

size(bp)	Genotype																																					
	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	Y	Z	AA	AB	AC	AD	AE	AF	AG	AH	AI	AJ		
1650	1	0	1	0	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	1	1	1	
1490	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1440	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
1210	0	0	0	0	1	1	0	0	1	0	0	0	1	0	1	1	1	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
1140	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
920	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
910	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
820	1	1	1	1	1	1	1	1	0	0	0	0	0	1	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
810	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
770	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
740	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0	0	0	1	1	1	1	1	1	1	1	1	1
720	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
680	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1
640	0	0	0	1	1	0	0	1	0	0	0	0	0	0	0	0	0	1	1	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
590	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	
580	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	
540	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
480	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	0	1	0	0	0	0	0	1	0	
450	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	
430	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
395	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
370	1	1	1	1	1	1	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
305	1	1	1	1	1	1	1	1	0	0	1	1	0	0	1	1	1	1	0	0	1	0	0	1	1	0	1	1	0	1	1	0	1	1	0	1	0	0
290	1	1	1	0	0	0	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
265	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1	0	0	0	0	0	0	0	0	0	

OPA11 primer (Continue)

Size (bp)	Genotype																					
	AK	AL	AM	AN	AO	AP	AQ	AR	AS	AT	AU	AV	AW	AX	AY	AZ	BA	BB	BC	BD	BE	BF
1650	1	1	0	1	1	1	1	1	0	1	0	0	1	1	0	0	1	0	0	0	0	0
1490	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	1
1440	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	0
1300	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1
1210	1	1	1	1	1	1	0	0	1	1	1	1	0	1	1	1	1	0	0	0	0	0
1140	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0
920	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	1	1	0	0	0	0	0
910	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1	1	1	1
820	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	1	0	0	0	0	0
810	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1
770	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0	0
740	1	1	0	0	0	0	0	0	0	0	1	1	0	0	0	1	0	0	0	0	0	0
720	0	0	1	1	0	1	1	0	0	0	0	1	0	0	1	0	0	0	0	0	0	0
680	0	0	1	1	1	1	1	1	1	1	1	0	1	1	0	1	1	0	0	0	0	0
640	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	1	1	1	1	1
590	0	1	1	1	1	1	1	1	1	1	1	0	1	1	0	1	0	0	0	0	0	0
580	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1	0	0	0	0	0	0	0
540	1	1	1	1	1	1	1	1	1	1	1	0	1	1	1	1	1	0	0	0	0	0
480	1	0	1	1	0	1	1	0	1	0	1	0	1	1	0	0	0	0	0	0	0	0
450	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0	0	0	1	1	1	1
430	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0	0
395	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
370	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0
305	0	0	0	0	0	0	0	0	0	0	1	1	0	0	1	0	0	0	0	0	0	0
290	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0
265	0	0	1	1	1	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0



## Biography

Miss Warisa Tangjingjai was born on April 1, 1972 in Prachuap Khiri Khan. She graduated with the degree of Bachelor of Science from Department of Biology at Chulalongkorn University in 1994. In 1995, she has studies Master degree of Science at the program of Biotechnology, Chulalongkorn University.

