## CHAPTER V

## RESULTS

## Part I. Cell culture for propagation of virus

Vero cells; a continuous cell line of African green monkey kidney, were subcultured, the number of the passages of 215 - 354 were used in this study. Cells were frozen in freezing media and kept at -70°C.

## Part II. Herpes simplex virus

Isolation of Herpes Simplex Virus

One hundred nineteen specimens were obtained from the patients with suspected genital herpes; 79 (66.4%) specimens were from Bangkok and 40 (33.6%) specimens were from Chonburi. All of specimens were from 51 men and from 68 women that divided to 46 prostitutes and 22 house-wife contacts. The age of patients were between 14-62 years, 92 patients had prior history of genital herpes and 21 patients with vesicle and 98 patients with ulcer appearances. Twenty-two patients had prior history of antiviral drug uptake. Thirty-eight (31.9%) specimens were cultured positive for CPE. Twenty-eight (73.7%) were collected from Bangkok and 10 (26.3%) were collected from Chonburi. There were 3 patients with HIV infection, one of them was cultured positive for CPE and one of 119 specimens was bisexual orientation (Table 3).

No significant differences in viral isolations ( $\alpha$ = .05) between place of collection ( $\chi^2$  = 0.56), sex ( $\chi^2$  = 2.15), female status ( $\chi^2$  = 0.5), age ( $\chi^2$  = 0.71) and prior history of antiviral drug uptake ( $\chi^2$  = 1.03), but were seen significantly different between prior history of genital herpes ( $\chi^2$  = 47.15) and genital herpes appearance ( $\chi^2$  = 47.15).

### **HSV** identification

The development of ELISA technique for HSV-2 identification, the procedure was 100  $\mu$  of the polyclonal rabbit anti-HSV-2 was diluted 2 fold, beginning, 1:1000 to 1: 8000 for coating the wells of microtiter plate overnight at 4°C, then the plate was washed 3 times with PBS-T and 150  $\mu$  of blocking solution was added for one hour at 37°C to protect the nonspecific binding. The plate was washed 3 times with PBS-T and added 100 ДІ of HSV-2, strain 186 which was 2 fold diluted with diluent starting, 5x10<sup>2</sup> to 1x10<sup>5</sup> PFU/ml, for one hour at 37°C. Then, the plate was washed 3 times with PBS-T and added 100 Ltl of 1:200 of peroxidase - conjugate rabbit anti-HSV-2 was added and incubated at 37°C for one hour. The plate was washed 3 times with PBS-T and 100  $\mu$  of substrate was added for one hour at 37°C and the reaction was stopped with 50 W of 4N H<sub>2</sub>SO<sub>4</sub>. OD at 492 nm was recorded. It was found that the appropriate dilution of rabbit anti-HSV-2 for coating was 1: 4000 (Table 4 and Figure 13). When the rabbit anti-HSV-2, 1:4000 was coated to the wells and the reaction was done with conjugate which was diluted 2 fold, beginning, 1:200 to 1:1000. The optimal conjugate dilution was 1:200 and the optimal viral concentration was 1 x 10<sup>4</sup> PFU/ml (Figure 14). Therefore, in this study, 100  $\mu$ l of the rabbit anti-HSV-2, 1: 4000 for coating, the viral concentration 1 x10<sup>4</sup> PFU/ml, and conjugate dilution 1: 200 were used for HSV identification of 38 HSV isolates. Results of the HSV-2 identification by ELISA were showed in Table 6. The average OD of reagent control (no virus) was 0.117. The cut off value that was 2 times of control OD, was 0.234. The range of HSV-2 isolate OD was 0.634 - 1.200.

The result in comparison of HSV plaque size between HSV-1 strain KOS, HSV-2 strain 186 and HSV isolates showed all plaques of HSV isolates were bigger than plaque of HSV-1 strain KOS and the plaque size of isolates were similar to that of HSV-2, strain 186.

#### Titration of virus

The thirty-eight HSV-2 isolates from primary culture were titrated and virus concentration as shown in Table 7 were used in this study. The range of HSV-2 isolates concentration was  $1.2 \times 10^8 - 4.2 \times 10^7$  PFU/ml.

## Part III. Study of antiviral activity of acyclovir against HSV strain 186.

Antiviral activity (ED<sub>so</sub>) of acyclovir against HSV strain 186 by pre-treatment, post-treatment and inactivation were 0.56, 0.72 and 0.5  $\mu$ I/mI, respectively (Table 8). There was significant difference among treatments. Table 9 showed the statistic using ANOVA which compared the mean of dual treatments. Pre-treatment / post-treatment ( $t_{.06}$ = 0.16) and post-treatment/ inactivation ( $t_{.06}$ = 0.22) were significantly different. No significant difference between inactivation and pre-treatment ( $t_{.06}$ = 0.06) and the least concentration of ACV (0.5  $\mu$ g/mI) in activation was observed. So, this treatment was used to study antiviral activity of acyclovir and medicinal plant extracts against HSV-2 strain 186 and HSV-2 isolates.

## Part IV. Study of antiviral activity of acyclovir against HSV-2 isolates.

Thirty-eight HSV-2 isolates showed the ED<sub>50</sub> of ACV in range 0.38 - 0.87  $\mu$ g/ml, the mean  $\pm$  SD was  $0.585 \pm 0.1$   $\mu$ g/ml (Table 10). The ED<sub>50</sub> of ACV against 38 HSV-2 isolates and HSV-2 strain 186 by inactivation activity were found significantly different (t = 5.31, P < .05).

# Part V. Study of antiviral activity of medicinal plant extracts against HSV-2 strain 186 and HSV-2 isolates

Thirty-five of medicinal plant extracts from Cerbera odollam Gaertn., Clausena excavata Burm.F., Coleus amboinicus Lour., Phyla nodiflora (L.) Greene, and Thevetia peruviana Schum were used. All of extracts were tested for cytotoxicity assay in Vero cell, the results were indicated in Table 11. The cytotoxic concentration of almost extracts were >50 µg/ml, except, fraction 5(F5) of Cerbera odollam Gaertn., fraction 2 (F2) and fraction 5 (F5) of Coleus amboinicus Lour. were >10 µg/ml. While, fraction 2 (F2) of Clausena excavata Burm.F., and fraction 2 (F2) and fraction 5 (F5) of Thevetia peruviana Schum. were >20 µg/ml.

Table 12 indicated the ED<sub>50</sub> of medicinal plant extracts against HSV-2 strain 186, the extracts from F1, F2, F4, and F5 elicited active antiviral activity against HSV-2 strain 186, therefore, these fraction-extracts were used to study antiviral activity against HSV-2 isolates by inactivation-activity.

Table 13 showed the range and mean  $\pm$  SD of ED<sub>50</sub> of medicinal plant extracts against all HSV-2 isolates (the raw data showed in Appendix III). The most active extracts of Cerbera odollam Gaertn., Clausena excavata Burm.F., Coleus amboinicus Lour., Phyla nodiflora (L.) Greene, and Thevetia peruviana Schum. were F5 (4.99 $\pm$ 0.91  $\mu$ g/ml), F2 (8.97 $\pm$ 0.78  $\mu$ g/ml), F2 (3.57 $\pm$ 0.77  $\mu$ g/ml), F4 (20.90 $\pm$ 3.14  $\mu$ g/ml) and F2 (3.04 $\pm$ 0.70  $\mu$ g/ml), respectively. There were significant differences between groups of plant in the same plant, likewise it was seen significantly different between groups of plant in the same fraction.

Results of the tests of significance for  $ED_{50}$  between groups of fraction in the same plant were shown in Table 14, the mean  $ED_{50}$  of fractions in 5 plants were significantly different and the mean could be ranked by Dancan test with significant

level .05 into 3 subsets, except P. nodiflora divided into 2 subsets. The tests of significance for  $ED_{50}$  between groups of plant in the same fraction were indicated in Table 15. All fractions were significantly different and the mean could be ranked by Dancan test with significant level .05 into 4 subsets.

Part VI. Study of antiviral activity of inhibiting viral entry into cell, growth inhibition of viral replication in the cell and direct viral demolishment of medicinal plant extracts against HSV-2 strain 186 and HSV-2 isolates

ACV 0.5 μg/ml, F1 extracts from 5 plants 50 μg/ml were used as drug control and medicinal plant extracts. HSV-2 strain 186 and 6 HSV-2 isolates chose by simple random sampling were used in this study. Six isolates were A6, A8, A13, A47, BC4 (isolate No. 3, 4, 5, 19, 27 in Appendix III) from Bangkok and C8 (isolate No. 33 in Appendix III) from Chonburi. Results of antiviral activity of ACV and F1 medicinal plant extracts against HSV-2 strain 186 and HSV-2 isolates were shown in Figure 15. It was found that the pre - treatment and post - treatment activity could not reduce the percent of plaque forming of HSV-2 strain 186 and 6 HSV-2 isolates. In spite of increasing exposure time of medicinal plant extracts in the reaction up to 3 hours, the extracts showed no more antiviral activity. On the other hand, the percent of plaque forming were reduced by ACV and all of 5 plants in inactivation.

Table 3. Patients and lesion characteristics

Characteristic	Total (N=119)	CPE + (N=38)	CPE - (N=81)	p valu
Place of collection				
Bangkok	79 (66.4)	28 (73.7)	51 (63.0)	p>.05
Chonburi	40 (33.6)	10 (26.3)	30 (37.0)	
Sex			<del></del>	
male	51 (42.9)	20 (52.6)	31 (36.3)	p>.05
female	68 (57.1)	18 (47.4)	50 (61.7)	
Female status				
prostitute	46 (38.7)	11 (28.9)	35 (43.2)	p>.05
house-wife contact	22 (18.5)	7 (18.4)	15 (18.5)	
Age				•
14-29	70 (58.8)	24 (36.2)	46 (56.8)	p>.05
30-49	46 (38.7)	14 (36.8)	32 (39.5)	
≥50	3 (2.5)	0 (0)	3 (3.7)	
Prior history of genital herpes	92 (77.3)	35 (92.1)	57 (70.4)	* p<.05
No history of genital herpes	27 (22.7)	3 (7.9)	24 (29.6)	
Genital herpes appearance	55000	100000		
vesicle	21 (17.6)	20 (52.6)	1 (1.2)	* p<.05
ulcer	98 (82.4)	18 (47.4)	80 (98.8)	
Prior history of antiviral drug up	otake 22 (18.5)	5 (13.2)	17 (21.0)	p>.05
No history of antiviral drug upta	ake 97 (81.50	33 (86.8)	64 (79.0)	
HIV status				
infected	3 (2.5)	1 (2.6)	2 (2.5)	
not infected	104 (87.4)	37 (97.4)	67 (82.7)	
unknown	12 (10.1)	0 (0)	12 (14.8)	
Sexual orientation			· · · · · · · · · · · · · · · · · · ·	
bisexual	1 (0.8)	0 (0)	1 (1.2)	
heterosexual	118 (99.2)	38 (100)	80 (98.8)	

Value are no. (%),  $p \le .05 = significance$ 

Table 4. Titration of dilution of rabbit anti-HSV-2 serum against viral concentration. (Conjugate dilution 1: 200 )

Viral concentration	O.D.ELISA of rabbit anti-HSV-2 serum dilution coated plate					
(PFU/mi)	1:1000	1:2000	1:4000	1:8000		
5x10 <sup>2</sup>	0.137	0.153	0.161	0.152		
1x10³	0.191	0.215	0.259	0.225		
5x10 <sup>3</sup>	0.459	0.589	0.674	0.555		
1x10 <sup>4</sup>	0.662	0.818	0.942	0.769		
5×10⁴	1.413	1.738	1.925	1.512		
1×10 <sup>5</sup>	1.481	1.845	1.992	1.622		

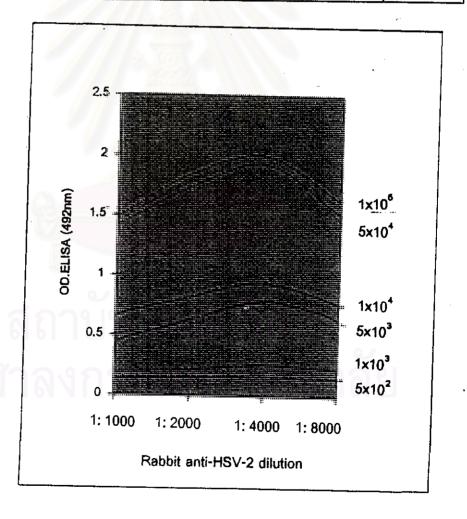


Figure 13. Titration curves of dilution of rabbit anti-HSV serum against viral concentration. (Conjugate dilution 1: 200)

Table 5. Titration of dilution of conjugate rabbit anti-HSV serum against viral concentration. (Rabbit anti-HSV-2 dilution 1: 4000)

Viral concentration	O.D.ELISA of conjugate dilution					
(PFU/ml)	1:200	1:400	1:800	1:1000		
5x10 <sup>2</sup>	0.135	0.115	0.121	0.126		
1x10 <sup>3</sup>	0.391	0.248	0.182	0.141		
5x10 <sup>3</sup>	0.612	1.369	0.224	0.119		
1x10 <sup>4</sup>	1.604	0.916	0.571	0.199		
5×10 <sup>4</sup>	2.162	1.222	0.766	0.242		
1x10 <sup>5</sup>	3.001	2.043	1.301	0.347		

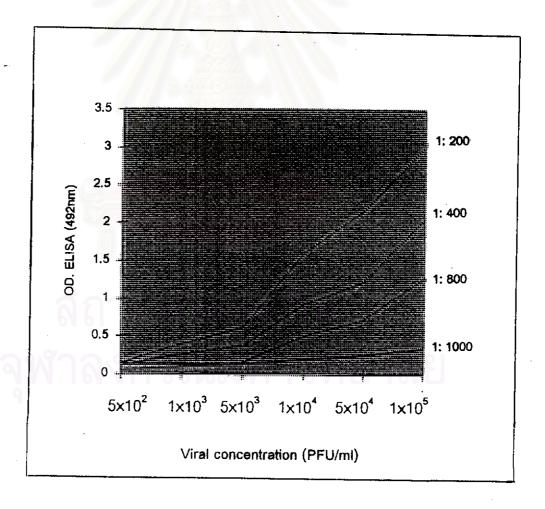


Figure 14. Titration of dilution of conjugate rabbit anti-HSV serum against viral concentration. (Rabbit anti-HSV-2 dilution 1: 4000)

Table 6. OD ELISA (492 nm) of HSV-2 identification.

Isolate	<b>O</b> D	Results	Isolate	OD	Results
No.	ELISA		No.	ELISA	
reagent 1	0.118	blank	МЕМ	0.126	- control
reagent 2	0.112	blank	MEM	0.134	- control
reagent 3	0.121	blank	HSV-2(188)	0.614	+ control
1.	0.766	+	20	1.030	+
2	0.944	+	21	0.871	+
3	0.987	+	22	0.858	+
4	0.654	+	23	0.871	+
5	0.887	+	24	0.916	+
6	0. <b>6</b> 75	+	25	0.785	+
7	0.897	+	26	1.124	+
8	1.010	+	27	0.634	+
9	0.753	+	28	0.671	+
10	1.102	+	29	1.120	+-
11	0.875	+	30	0.854	+
12	1.120	+	31	0.985	+
13	0.992	+	32	0.785	+
14	0.895	<b>+</b>	33	0.992	+
15	0.852	+	34	0.884	+
16	0.987	+	-35	1.200	+ 0
17	0.843	11-71	36	1.109	12+16
18	0.658	+	37	1. <b>19</b> 2	+
19	0.753	+	38	0.952	+

Cut Off Value (COV) =  $0.118 + 0.112 + 0.121 \times 2 = 0.234$ 

Table 7. Viral titration of HSV-2 strain 186 and HSV-2 isolates

Isolate No.	concentration	Isolate No.	Concentration
	(PFU/ml)		(PFU/mi)
1.	6x10 <sup>6</sup>	21	1.8×10 <sup>6</sup>
2	5.2x10 <sup>6</sup>	22	2.4x10 <sup>6</sup>
3	8x10 <sup>6</sup>	23	2.6x10 <sup>6</sup>
4	6.2×10 <sup>6</sup>	24	3.2x10 <sup>6</sup>
5	1x10 <sup>7</sup>	25	1x10 <sup>7</sup>
6	5x10 <sup>6</sup>	26	2.6x10 <sup>6</sup>
7	8.0x10 <sup>6</sup>	27	3.2×10 <sup>6</sup>
8	1.8x10 <sup>6</sup>	28	3.6×10 <sup>6</sup>
9	1.2x10 <sup>6</sup>	29	4.2×10 <sup>7</sup>
10	1.24x10 <sup>6</sup>	30	1.2×10 <sup>7</sup>
11	3.2x10 <sup>6</sup>	31	4.0x10 <sup>6</sup>
12	3.7x10 <sup>6</sup>	32	1.2×10 <sup>6</sup>
13	3.2×10 <sup>6</sup>	33	2.4x10 <sup>6</sup>
14	4.2x10 <sup>6</sup>	34	2.6x10 <sup>6</sup>
15	1.2x10 <sup>6</sup>	35	1.2×10 <sup>6</sup>
16	1×10 <sup>7</sup>	36	5.0x10 <sup>6</sup>
17	3.2x10 <sup>6</sup>	37	2.6x10 <sup>6</sup>
18	4.0x10 <sup>8</sup>	38	3.9x10 <sup>6</sup>
19	2.4×10 <sup>6</sup>	HSV-2(186)	5.0x10 <sup>6</sup>
9 20	2.6x10 <sup>6</sup>	Range	1.2x10 <sup>6</sup> -4.2x10 <sup>7</sup>

The average value from duplicated experiments of primary culture were done.

Table 8. Antiviral activity of ACV against HSV-2 strain 186

Experiments	ED <sub>so</sub> of ACV (μg/ml)				
	pre-treatment	post-treatment	inactivation	1	
1	0.52	0.65	0.5		
2	0.55	0.70	0.5	p<.05	
3	0.60	0.80	0.5		
total	1.67	2.15	1.5		
mean	0.5 <b>6</b>	0.72	0.5		

p < .05 = significance

Table 9. ANOVA table of antiviral activity of ACV against HSV-2 strain 186

source	degree of freedom(df)	sum of square (ss)	mean square  MS = ss/df	VR
among group	2	0.08	0.04	25
within group	6 10	0.01	0.0016	
total	8	0.09		p<.05

Compare mean with Least Significant Difference (LSD)

LSD 
$$t_{05} = 0.0976$$

Pre-treatment / Post-treatment = [0.56-0.72] = 0.16, p<.05

Post-treatment / Inactivation = [0.72-0.50] = 0.22, p<.05

Inactivation / Pre-treatment = [0.50-0.56] = 0.06, p>.05

Table 10. ED<sub>so</sub> of ACV against HSV-2 isolates

Isotate No.	μg/ml*	isolate No.	μg/mi*
1,	0.630	21	0.595
2	0.595	22	0.670
3	0.565	23	0.620
4	0.450	24	0.485
5	0.625	25	0.600
6	0.375	26	0.530
7	0.625	27	0.430
8	0.460	28	0.610
9	0.550	29	0.400
10	0.655	30	0.640
11	0.720	31	0.567
12	0.690	32	0.550
13	0.730	33	0.525
14	0.530	34	0.870
15	0.605	35	0.545
16	0.610	36	0. <b>6</b> 45
17	0.595	37	0.550
18	0.470	38	0.750
19	0.660	sum	22.227
20	0.505	mean <u>+</u> SD	0.585 <u>+</u> 0.1

<sup>&</sup>lt;sup>a</sup> The average value from duplicated experiments were done

Table 11. Cytotoxic concentration of medicinal plant extracts on Vero cell

Plants	Cytotoxic concentration of extracts (µg/ml)							
	F1	F2	F3	F4	F5	F6	F7	
Cerbera odollam Gaertn.	>50	>50	>50	>50	>10	>50	>50	
Clausena excavata Burm.F.	>50	>20	>50	>50	>50	>50	>50	
Coleus amboinicus Lour.	>50	>10	>50	>50	>10	>50	>50	
Phyla nodiflora (L.) Greene,	>50	>50	>50	>50	>50	>50	>50	
Thevetia peruviana Schum.	>50	>20	>50	>50	>20	>50	>50	

<sup>&</sup>lt;sup>a</sup> The average value from duplicated experiments were done.

F1 = methanol extract

F5 = aqueous- hexane extract

F2 = chloroform extract

F6 = butanol extract

F3 = aqueous-methanol extract F7 = aqueous-butanol extract

F4 = hexane extract

Table 12. ED<sub>so</sub> of medicinal plant extracts against HSV-2 strain 186

Plants	ED <sub>50</sub> of extracts (μg/mi) <sup>a</sup>						
	F1	F2	F3	F4	F5	F6	F7
Cerbera odollam Gaertn.	7.50	10.25	≥50	7.25	4.25	≥50	≥50
Clausena excavata Burm.F.	11.25	9.25	≥50	23.25	19.75	≥50	≥50
Coleus amboinicus Lour.	20.88	3.25	≥50	12.75	4.00	≥50	≥50
Phyla nodiflora (L.) Greene.	23.75	22.5	≥50	16.25	21.25	≥50	≥50
Thevetia peruviana Schum.	10.25	3.25	≥50	10.25	12.63	≥50	≥50

<sup>&</sup>lt;sup>a</sup> The average value from duplicated experiments were done.

F1 = methanol extract

F5 = aqueous-hexane extract

F2 = chloroform extract

F6 = butanol extract

F3 = aqueous-methanol extract

F7 = aqueous-butanol extract

F4 = hexane extract

Table 13. ED<sub>so</sub> of medicinal plant extracts against HSV-2 isolates (N=38)

Frac	tions	C. odollam (μg/ml)	C. excavata (µg/ml)	C. amboinicus	P.nodiflora (μg/ml)	T.peruviana (μg/ml)	p <b>value</b>
F1	range	7.50-15.75	11.00-18.25	7.50-15.75	16.25-26.10	8.25-13.25	72.00
	X±SD	10.30 <u>+</u> 1.87	13.78 <u>+</u> 2.05	18.64 <u>+</u> 1.95	22.00 <u>+</u> 2.36	10.64 <u>+</u> 1.47	p<.05
F2	range	8.63-12.70	7.50-11.00	2.13-5.12	21.50-35.00	2.13-5.00	
	X±SD	10.57 <u>+</u> 0.92	8.97 <u>+</u> 0.78	3.57 <u>+</u> 0.77	27.26 <u>+</u> 3.90	3.04 <u>±</u> 0.70	p<.05
F4	range	6.25-9.25	19.25-29.80	9.75-16.25	16.25-27.10	9.50-14.75	
	X±SD	7.44 <u>+</u> 0.76	22.98 <u>+</u> 2.93	12.93 <u>+</u> 1.88	20.90 <u>+</u> 3.14	12.17 <u>+</u> 1.10	p<.05
F5	range	3.25-6.63	17.00-27.50	2.50-5.25	13.25-27.13	10.00-13.75	
	X±SD	4.99 <u>+</u> 0.91	20.61 <u>+</u> 2.48	3.60 <u>±</u> 0.56	21.14 <u>+</u> 3.01	12.63 <u>+</u> 0.88	p<.05
р	value			p<.05		·	<u> </u>

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Table 14. Tests of significance for  $ED_{50}$  of medicinal plant extracts between groups of fraction in the same plant by ANOVA (Randomized Complete Block Design).

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df	SS	MS	F .
3	791.75	263.92	302.16
37	116.30	3.14	3.60
111	96,94	.87	<u>"-</u>
151	151 1004.99		
Danca	n test with a	significant (	O05)
F5	F4	F1	F2
4.99	7.44 <sup>b</sup>	10.34°	10.57°
	3 37 111 151 Danca F5	3 791.75 37 116.30 111 96.94 151 1004.99  Dancan test with a	3 791.75 263.92  37 116.30 3.14  111 96.94 .87  151 1004.99  Dancan test with significant (  F5 F4 F1

#### C. excevata

Source	df		SS	MS	F	
Treatment	3	46	612.12	1537.37	267.8	5
Block(isolate)	37	2	287.96	7.78	1.30	6
Residual	111	6	348.58	5.74	,	
Total	151	55	34.19			
	Danca	in te	st with s	ignificant	( <b>α</b> .05)	
Fraction	F2		F1	F5	i F	4
Mean	8.97ª		13.78	b 20.2	7° 22	.97°

#### C. amboinicus

Source	df		SS		MS		F	
Treatment	3	62	280,72	20	93.57	110	3.77	
Block(isolate)	37	00	91.21		2.47		1.30	6
Residual	111 210.54			1.90				
Total	151 6582.47		- "					
	Danca	ın te	st with si	ignific	cant ( C	χ.05	)	
Fraction	F2		F5		F4		F1	
Mean	3.57	á	3.60°		12.93 <sup>b</sup>	1	18.64°	- <b>-</b> -

## Table 14. (Continued)

## P. nodiflora

Source	df		SS		MS	F		
Treatment	3	10	21.58	34(	0.53	36.18		
Block(isolate)	37 423.86		11.46		1.22			
Residual	111 1044.86		9	9.41				
Total	151 2490.30							
Dancan test with significant ( 0.05)								
Fraction	F4		F5		F1	F2		
Mean	20.90°		21.14	a	22,00°	27.26 <sup>b</sup>		

## T. peruviana

Source	df	SS	MS	F				
Treatment	3	2276.88	758.96	1112.73				
Block(isolate)	37	96.23	2.60	3.81				
Residual	111	75,71	.68					
Total	151 2448.82							
Dancan test with significant ( 02 .05)								
Fraction	F2	F1	F4	F5				
Mean	3.04	10.63	3 <sup>b</sup> 12.1	7° 12.63°				

The same superscript shows no significantly different

F1 = methanol extract

F2 = chloroform extract

F4 = hexane extract

F5 = aqueous-hexane extract

Table 15. Tests of significance for ED<sub>50</sub> of medicinal plant extracts between groups of plant in the same fraction by ANOVA (Randomized Complete Block Design).

## Fraction 1 (F1)

Source	df	SS	MS	F
Treatment	4	3987.82	996.95	305.55
Block(isolate)	37	229.18	6.19	1.90
Residual	148	462.89	3.26	
Total	189	4699.89		

Dancan test with significant ( 04.05)

Plant	C.odollam	T.peruviana	C.excevate	C.amboinicus	P.nodiflora
Mean	10.30°	10.83 <sup>b</sup>	13.78°	18.64 <sup>d</sup>	22.00 <sup>d</sup>

## Fraction 2 (F2)

Source	df	SS	MS	F
Treatment	4	14698.95	3874.24	1118,18
Block(isolate)	37	167.69	4.53	1.38
Residual	148	488.32	3.29	
Total	189	15350.96		
	Danc	an toet with ei	anificant ( C	Y 05)

Plant	T.peruviana	C.amboinicus	C.excevata	C.odollam	P.nodiflora
Mean	3.04ª	3.57ª	8.97 <sup>b</sup>	10.57°	27.28 <sup>d</sup>

## Fraction 4(F4)

df	SS	MS	F
4	6358.35	1589,59	345.71
37	197.66	5.34	1,16
148	680.52	4.60	
189	7236,53		•
	4 37 148	4 6358.35 37 197.66 148 680.52	4     6358.35     1589.59       37     197.66     5.34       148     680.52     4.60

Dancan test with significant ( & .05)

Plant	C.odollam	T.peruviana	C.amboinicus	P.nodiflora	C.excavata
Mean	7.44 <sup>a</sup>	12.17 <sup>b</sup>	12.93 <sup>b</sup>	20.90°	22.97 <sup>d</sup>

## Table 15. (Continued)

	LINCAR	ns o(ro)		
Source	df	SS	MS	F
Treatment	4	10606.08	2651.52	876.37
Block(isolate)	37	163.36	4.96	1.64
Residual	148	447.79	3.03	
Total	189	11237,22		
	Danca	an test with sig	nificant ( OL	.05)

Plant	C.amboinicus	C.odollam	T.peruviana	C.excavata	P.nodiflora
Mean	3.60°	4.99 <sup>b</sup>	12 <b>.63</b> °	20.81 <sup>d</sup>	21.14 <sup>d</sup>

The same superscript shows no significant difference

F1 = methanol extract

F2 = chloroform extract

F4 = hexane extract

F5 = aqueous-hexane extract

