

REFERENCES

- Adlercreutz, H., Honju, H., Higashi, A., Fotsis, T., Hamallainen, E., Hasegawa, T. and Okada, H. 1991. Urinary excretion of lignans and isoflavonoid phytoestrogens in Japanese men and women consuming a traditional Japanese diet. *Am. J. Clin. Nutr.* 54: 1093-1100
- Akiyama, T., Ishida, J., Nakasawa, S., Ogawara, H., Watanabe, S., Itoh, N., Shibuya, M. and Fukami Y. 1987. Genistein, a specific inhibitor of tyrosine-specific protein kinases. *J. Biol. Chem.* 262: 5592-5595
- Awad, A. B., Downie, A. and Fink, C. S. 2000. Inhibition of growth and stimulation of apoptosis by beta-sitosterol treatment of MDA-MB-231 human breast cancer cells in culture. *Int J Mol Med.* 5: 541-545
- Awad, A. B., Downie, A., Fink, C. S. and Kim, U. 2000. Dietary phytosterol inhibits the growth and metastasis of MDA-MB-231 human breast cancer cells grown in SCID mice. *Anticancer Res.* 20: 821-824
- Awad, A. B. and Fink, C. S. 2000. Phytoesterol as anticancer dietary components: Evidence and mechanism of action. *J. Nutr.* 130: 2127-2130
- Awad, A.B., Von Holtz R., Conne, J. P., Fink C. S. and Chen Y. C. 1998. β -sitosterol inhibits the growth of HT-29 human colon cancer cells by activating the sphingomyelin cycle. *Anticancer Res.* 18: 471-479
- Awad, A. B., Williams, G. and Fink, C. S. 2001. Phytoestrols reduce in vitro (SCID mice) effects of phytoesterols on the growth and dissemination of human prostate cancer PC-3 cells. *Eur J Cancer Prev.* 10: 507-513
- Barnes, S., Kirk, M., and Coward, L. 1994. Isoflavones and their conjugates in soy foods: extraction conditions and analysis by HPLC-mass spectrometry. *J. Agric. Food Chem.* 42: 2466-2474
- Barnes, S., Sfakianos, J., Coward, L., and Kirk, M. 1996. Soy isoflavonoids and cancer prevention. Underlying biochemical and pharmacological issues. In Back, N., Cogen, I.R., Kritchevsky, D., Lajtha, A., and Paoletti., R.(eds) *Dietary phytochemical in cancer prevention and treatment*. New York. Plenum Press. 87-99
- Barnes, S., Coward, L., Kirk, M., and Sfakianos, J. 1998. HPLC-Mass spectrometry analysis of isoflavones (44230). *Proc. Soc. Exp. Biol. Med.* 217: 253-262

- Benson,G.K., Cowie, A.T. and Hosking, Z.D. 1961. Mammogenic activity of miroestrol. *J. Endocri.* 21: 401-409
- Bingham, S. A., Atkinson, C., Liggins, J., Bluck, L., and Coward, A. 1997. Phytoestrogens: where are we now?. *British Journal of Nutrition.* 79: 393-406
- Bound and Pope, G.S. 1960. Light absorption and chemical properties of miroestrol, the estrogenic substance of *Pueraria mirifica*. *J. Chem. Soc.*: 3196-3205
- Brandis, D. 1990. *Indian trees*. London: Archibald Constable & CO.LTD.
- Cain, J.C. 1960. Miroestrol: an estrogen from the plant *Pueraria mirifica*. *Nature*.158: 774-777
- Carrao-panizzi, C.M., Pino-Beleia, A. D., Kitamura, K., and Neves-Oliveira, M. C. 1999. Effects of genetics and environment on isoflavone content of soybean from different regions of Brazil. *Pesq. Agropec. Bras., Brasilia.* 34: 1787-1795
- Cheewasopit, W. 2001. *Antiproliferative effects of Pueraria mirifica, Butea superba and Mucuna collettii on human mammary carcinoma MCF-7 and cervical carcinoma HeLa* . Master's Thesis Chulalongkorn University.
- Chandra, A., Rana, J., and Li, Y. 2001. Separation, identification, quantification, and method validation of anthocyanins in botanical supplement raw materials by HPLC and HPLC-MS. *J. Agric. Food Chem.* 49: 3515-3521
- Chansakaow, S., Ishikawa, T., Seki, H., Sekine, K. (nee Yoshizawa), Okada, M., and Chaichantopyuth, C. 2000. Identification of deoxymiroestrol as the actual Rejuvenating principle of "Kwao Keur", *Pueraria mirifica*. The known miroestrol may be an artifact. *J. Nat. Prod.* 63: 173-175
- Chansakaow, S., Ishikawa, T., Seki, H., Sekine, K. (nee Yoshizawa), Okada, M., and Chaichantopyuth, C. 2000. Isoflavonoids from *Pueraria mirifica* and their estrogenic activity. *Planta Med.* 66: 572-575
- Dixon, R.A., Canovas, P., Guo, Z-J, He, X-Z, Lamb, C. and McAlister, F. 1999. Molecular controls for isolavonoid biosynthesis in relation to plant and human health . In Romeo, J. T. (ed.). *Phytochemicals in human health Protection, Nutrition and Plant Defense*. vol. 33 New York. Kluwer Academic/Plenum Publishers. 133-153 pp

- Dweck, A. C. 2003. The Pueraria family with special interest in *Pueraria mirifica*. *Personal care.* 7-8
- Eldridge, A. C., and Kwolek, W. F. 1983. Soybean isoflavones: effect of environment and variety on composition. *J. Agric. Food Chem.* 31: 394-396
- Eldridge, A. C. 1982. High-performance liquid chromatography separation of soybean isoflavones and their glucosides. *J. Chromatogr.* 234: 494-496
- Fanti P., Monier-Faugere M.C., Gerg Z., Schmidt J., Morris P.E., Cohem D. and Malluche H.H. 1998. The phytoestrogen genistein reduces bone loss in short-term ovariectomized rats. *Osterporos Int.* 8: 274-281
- Franke, A. A., Custer, L. J., Wang, W., and Shi, C. Y. 1998. HPLC analysis of isoflavonoids and other phenolic agents from foods and from human fluids (44231). *Proc. Soc. Exp. Biol. Med.* 217: 263-273
- Harborne, J.B. 1999. *Classes and Function of Secondary Products From Plants*. In Walton, N. J. and Brown, D. E., (eds.) *Chemical from Plants: Perspectives on Plant Secondary Products*. London. Imperial collage Press. 1-23 pp.
- Hoyodom, M. 1971. *Constituents of the tuberous roots of Pueraria mirifica*. Master's Thesis, Chulalongkorn University. 33 p. (in Thai)
- Ingham, J. L., Markham, K.R., Dziedzic, S. Z. and Pope, G.S. 1986. Puerarin 6-o- β -apiofuranoside, a c-glycosylisoflavone o-glucoside from *Pueraria mirifica*. *Phytochemistry.* 25: 1772-1775
- Ingham, J. L., Tahara, S., and Dziedzic, S. Z. 1986. A chemical investigation of *Pueraria mirifica* roots. *Z. Naturforsch.* 41c: 403-408
- Ingham, J. L., Tahara, S., and Dziedzic, S. Z. 1988. Coumestan from the roots of *Pueraria mirifica* roots. *Z. Naturforsch.* 43c: 5-10
- Ingham, J. L., Tahara, S., and Dziedzic, S. Z. 1989. Minor isoflavones from the roots of *Pueraria mirifica* roots. *Z. Naturforsch.* 44c(9/10): 724-726
- Ishida H., Uesugi T., Hirai K., Toda T., Nukaya H., Yokotsuka K. and Tsuji K. 1998. Prevention effects of the plant isoflavones, daidzin and genistin on bone loss in ovariectomized rats fed with a calcium-deficient diet. *Biol Pharm Bull.* 21: 62-66

- Izumi, T., Piskula, M. K., Osawa, S., Obata, A., Tobe, L., Saito, M., Kataoka, A., Kubota, Y., and Kikuchi, M. 2000. Soy isoflavone aglycones are absorbed faster and in higher amounts than their glucosides in humans. *J. Nutr.* 130: 1695-1699
- Jones, H. E. H., and Pope, G. S. 1960. A study of the action of miroestrol and other oestrogens on the reproductive tract of the immature female mouse. *J. Endocrin.* 20: 229-235
- Jones, H. E. H., and Pope, G. S. 1961. A method for the isolation of miroestrol from *Pueraria mirifica*. *J. Endocrin.* 22: 303-312
- Kaizuka, H., and Takahashi, K. 1983. High-performance liquid chromatographic system for a wide range of naturally occurring glucosides. *J. Chromatogr.* 258: 135-146
- Katagiri, Y., Ibrahim, R. K., and Tahara, S. 2000. HPLC analysis of white Lupin isoflavonoids. *Biosci. Biotechnol. Biochem.* 64: 1118-1125
- Kaufman, P. B., Duke, J. A., Briemann, H., Boik, J., and Hoyt, J. E. 1997. A comparative survey of leguminous plants as sources of the isoflavones, Genistein and Daidzein: implications for human nutrition and health. *J Altern Complement Med.* 3: 7-12
- Kinochita, E., Sugimoto, T., Wzawa, Y., and Aishima, T. 1998. Differentiation of soy sauce produced from whole soybeans and defatted soybeans by pattern recognition analysis of HPLC profiles. *J. Agric. Food Chem.* 46: 877-883
- Kitamura, K., Igita, K., Kikuchi, A., Kudou, S. and Okubo, K. 1991. Low isoflavone content in early maturing cultivars, so called summer-type soybeans (*Glycine max* (L.) Merrill). *Japanese Journal of Breeding.* 41: 651-654
- Kudou, S., Fleury, Y., Welto, D., Magonlato, D., Uchida, T., Kitamura, K., and Okubo, K. 1991. Malanyl isoflavone glycosides in soybean seeds (*Glycine max* Merrill). *Agric. Biol. Chem.* 55: 2227-2233
- Kurz, Z. 1877. *Forest Flora of Britsh Burma* Vol. 1. Calcutta. Office of the superintendent of government printing.
- Markaverich, B.M., Webb B., Densmore C.L. and Gregory, R.R. 1995. Effects of coumestrol on estrogen receptor function and uterine growth in ovariectomized rats. *Environ Health Perspect.* 103: 574-589

- Menon, L.G., Kuttan, R., Nair, M.G., Chang Y.C. and Kuttan G. 1998. Effect of isoflavones genistein and daidzein in the inhibition of lung metastasis in mice induced by B16F-10 melanoma cells. *Nutr. Cancer.* 30: 74-7
- Morris, P.F., Ward, E.W.B. 1992. Chemoattraction of zoospores of the soybean pathogen, *Phytophthora sojae* by isoflavones. *Physiol. Mol. Plant Pathol.* 40: 17-22
- Muangman, V. and Cherdshewasart, W. 2001 . Clinical trial of the phytoestrogen-rich herb, *Pueraria mirifica* as a crude drug in the treatment of symptoms in menopausal women. *Siriraj Hosp. Gaz.* 53: 300-309
- Murkies, A. L., Wilcox, G., and Davis, S. R. 1997. Clinical review 92 : Phytoestrogens. *J. Clin. Endocrin. Met.* 83: 297-303
- Nilandihi, T., Kamthong, B., Isarasena, K. and Shiengthong, D. 1957. Constituents of the tuberous roots of *Pueraria mirifica*. *Z. Naturforsch.* 5c: 41
- Ohshima, Y., Okuyama, T., Takahashi, K., Takizawa, T., and Shibata, S. 1987. Isolation and high performance liquid Chromatography (HPLC) of isoflavonoids from the *Pueraria* root. *Planta Med.* 54: 250-254
- Ohyama, M., Tanaka, T., Ito, T., Inuma, M., Bastow, K. F. and Lee, H. K. 1999. Antitumor agents 200. Cytotoxicity of naturally occurring resveratololigomers and their acetate derivative. *Bioorg. Med. Chem. Lett.* 20: 3057-3060
- Panriansaen, R. 2000 . *Characterization of Pueraria mirifica populations from various parts of Thailand*. Master's Thesis, Chulalongkorn University.7-8pp.
- Pengklai, C. 1977. *Mucuna collettii*. In Smitinand, T. (ed). *Flora of Khao Yai National Park*. Thailand. 70p
- Peterson, G. and Barnes, S. 1991. Genistein inhibition of the growth of human breast cancer cells: Independence from estrogen receptors and multi-drug resistance gene. *Biochem Biophys Res Commun.* 179: 661-667
- Picherit, C., Coxam. V., Pelissero, C. B., Coulibaly, S. K., Davicco, M. J., Lebecque, P. and Barlet, J. P. 2000. Daidzein is more efficient than genistein in preventing ovariectomy-induced bone loss in rats. *J. Nutr.* 130:1675-1681
- Regwald, A., Meier, B., and Sticher, O. 1994. Qualitative and quantitative reversed-phase high-performance liquid chromatography of flavonoids in *Crataegus* leaves and flowers. *J. Chromatogr. A.* 677: 25-33

- Ridley, H. N. 1967. *The flora of the Malay Peninsula I.* A. Asher&Co. Amsterdam, Holland: 555-556
- Roengsumran, S., Petsom, A., Ngamrojanavanich, N., Rugsilp, T., Sittiwichewong, P., Khorphueng, P., Cherdshewasart, W., and Chaichantipyuth, C. 2000. Flavonoid and flavonoid glycoside from *Butea superba* Roxb. and their cAMP phosphodiesterase inhibitory activity. *J. Sci. Res. Chula. Univ.* 25(1): 170-176
- Roengsumran, S., Sookkongwaree, K., Petsom, A., Pornpakakul, S. and Sangvanich P. 2001. Cyclic AMP phosphodiesterase inhibitor from tubers of *Mucuna collettii* Lace. *27th Congress on Science and Technology of Thailand*: 184 p. (abstract)
- Schoeller, W., Dohrn, M. and Hohweg, W. 1940. An estrogenic substance from the tubers of the Siamese vine, *Butea superba*. *Naturwissenschaften*. 28: 252
- Setchell, K. D. R. and Welsh, M. B. 1987. High - performance liquid chromatographic analysis of phytoestrogens in soy protein preparations with ultraviolet, electrochemical and thermospray mass spectrometric detection. *J. Chromatogr.* 386: 315-323
- Smitinand, T. (revised by The Forest Herbarium, Royal Forest Department) 2001. *Thai plant name*. Royal Forest Department
- Song, T., Barua, K., Buseman, G., and Murphy, P. A. 1998. Soy isoflavone analysis: quality control and a new internal standard. *Am J Clin Nutr.* 68: 1474s-9s
- Smitasiri, Y. and Wunghai, C. 1986. Some biological aspects of *Pueraria mirifica*: 1) flower, pod and seed, *J. Sci. CMU.*,14 (1) : 67-74
- Suntara, L. A. 1931. *The Kwo Krue Tuber Pamplet*. Upatipong Printing. Chiangmai:18 pp. (in Thai)
- Suvatti, C. 1978. *Flora of Thailand*. Kurusapha Ladprao press, Thailand : 680.
- Tsukamoto, C., Shimada, S., Igita, K., Kudou, S., Kokubun, M., Okubo, K., and Kitamura, K. 1995. Factors affecting isoflavone content in soybean seeds: changes in isoflavones, saponins, and composition of fatty acids at different temperatures during seed development. *J. Agric. Food Chem.* 43: 1184-1192
- Tsusumi, N. 1995. Effect of coumestrol on bone metabolism in organ culture. *Bio Pharm Bull* 18: 1012-1015

- Van beek, T. A. 1999. Modern methods of secondary product isolation and analysis. In Walton, N. J. and Brown, D. E. (eds.) *Chemical from Plants: Perspectives on Plant Secondary Products*. London. Imperial collage Press and World Scientific Publishing Co., Pte, Ltd. 91-177 pp.
- Verma, S. P., Salomone, E. and Goldin, B. 1997. Curcumin and genistein plant natural products show synergistic inhibitory effects on the growth of human breast cancer MCF-7 cells induced by estrogenic pesticides. *Biochem Biophys Res Commun.* 233: 692-696
- Verpoorte, R. and Alfermann, A.W., eds. 2000. *Metabolic engineering of Plant Secondary Metabolism*. Netherlands. Kluwer Academic Publishers
- Viana, G. S. B., Matos, F. F., Araujo, W. L., Matos, F. J. A., and Craveiro, A.A. 1981. Essential oil of *Lippia grata*: Pharmacological effects and main constituents. *Quart. J. Crude Drug Res.* 19: 1-10
- Von Holtz, R. L., Fink, C. S. and Awad, A.B. 1998. β -sitosterol activates the sphingomyelin cycle and induces apoptosis in LNCaP Human Prostate Cancer Cells. *Nutr. and Cancer.* 32: 8-12
- Wagner, H. 1999. Phytomedicine Research in Germany. *Environ. Health Perspect.* 107: 779-781
- Wang, C. and Kurzer, M. S. 1997. Phytoestrogen concentration determines effects on DNA synthesis in human breast cancer cells. *Nutr. Cancer.* 28: 236-247
- Wang, H., and Murphy, P. A. 1994. Isoflavone composition of American and Japanese soybeans in Iowa: effects of variety, Crop Year, and Location. *J. Agric. Food Chem.* 42: 1874-1877
- Weisberger, J.H., Dolan, L. and Pitman, B. 1998. Inhibition of PhIP mutagenicity by caffeine, lycopene, daidzein and genistein. *Mutat Res.* 416: 125-128
- Westrate, J.A. and Meijer, G.W. 1998. Plant sterol-enriched margarines and reduction of plasma total- and LDL-cholesterol concentration in mildly hypercholesterolaemic subjects. *Eur. J. Clin Nutr.* 52: 334-343
- Wutteeraphon, S., Kawewat, K., Saenphet, S. and Luangpai, R. 2001. Effect of *Mucuna coletti* on reproductive organs and its toxicity on male rat. *27th Congress on Science and Technology of Thailand*: 424pp.(Abstract)

- Xu, Z., Wu, Q., and Godber, J. S. 2002. Stabilities of Daidzin, Glycitin, Genistin, and generation of derivatives during heating. *J. Agric. Food Chem.* 50: 7402-7406
- Yadava, R. N. and Reddy K. I. S. 1998. A novel flavone glycoside from the stems of *Butea superba*. *Fitoterapia*. Vol LXIX . 3: 269-270
- Zhang R., Li Y. and Wang W. 1988. Enhancement of immune function in mice fed high doses of soya daidzein. *Nutr Cancer*. 29: 24-48





APPENDICES

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

APPENDIX I

HIGH PERFORMANCE LIQUID CHROMATOGRAPHY

ศูนย์วิทยทรัพยากร
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HPLC

Chromatography was the most useful technique for the separation of phytochemicals. The chromatographic analysis were distributed between two phases, one of which was a stationary one while the other moved. The separation occurred because, under an optimum set of condition, each component in the mixture would interacted with the two phases differently relative to the other components in the mixture. The methods including thin layer chromatography (TLC), gas chromatography (GC) and high-performance liquid chromatography (HPLC). TLC is basically and widely used chromatographic technique available for the analysis of plant constituents, especially for preparation purposes. It is very simple, no special apparatus needed. GC and HPLC are more convenience, efficiency and resolution technique than TLC. The mobile phase of TLC and HPLC are liquid while the mobile phase of GC is gaseous. TLC and HPLC are thus the main analytical techniques for analysis of non-volatile compounds whereas GC is a technique of choice for volatile compounds. GC was limited to secondary metabolites which had boiling point below 450 °C and which were stable at the temperature of the separation. Therefore, HPLC had the advantage over GC in that it was not necessary to prepare volatile derivatives.

Chromatography is described and measured in terms of four major concepts: capacity, efficiency, selectivity and resolution. The capacity and selectivity of the column are mostly variable controlled by the column manufacturer, whereas efficiency and resolution could be controlled, to some extent, by the chromatographer. Important parameters in chromatographic are the retention time of a solute (t_R) and an unretained compound (t_0), the corrected time of a solute ($t_{R'}$), the volume of solute (V_R), the volume of solvent or void volume (V_0), the peak width at the baseline (w) and halfway of the peak ($w_{0.5}$).

The capacity factor (k'_R) of a column is a direct measure of the strength of the interact of the sample with the packing material. It was mostly function of packing material but could be manipulated to a degree by varying the solvent strength.

$$k'_R = \frac{(t_R - t_0)}{t_0} = \frac{(V_R - V_0)}{V_0}$$

The selectivity of the chromatographic system is a measure of the difference in retention times (or volumes) between two given peaks and described how effectively a chromatographic system could separate two compounds.(Figure 2) Selectivity is controlled using mobile phase and temperature and defined in term of α , where

$$\alpha = \frac{t_2 - t_0}{t_1 - t_0} = \frac{V_2 - V_0}{V_1 - V_0} = \frac{k'_2}{k'_1}$$

The resolution is a term used to describe the degree of separation between neighboring solute bands or peaks. It was affected by the selectivity (α), efficiency (N) and capacity (k') of column.

$$R_s = \frac{1}{4} \frac{\alpha - 1}{\alpha} (N^{1/2}) \frac{k'}{1 + k'}$$

The efficiency of column is a number that described peak broadening as a function of retention and it is defined in term of number of theoretical plates, N and N is defined in term of the retention time (t_r) of the solute and halfway of the peak. ($w_{0.5}$) The greater the number of theoretical plates, the more efficient the column is considered to be. The movement of a solute along the column is viewed as a stepwise transfer from one theoretical plates, the greater number of theoretical plate to the next. The thinner the theoretical plates, the greater the number that could be envisaged within a given length of column.

$$H = L / N$$

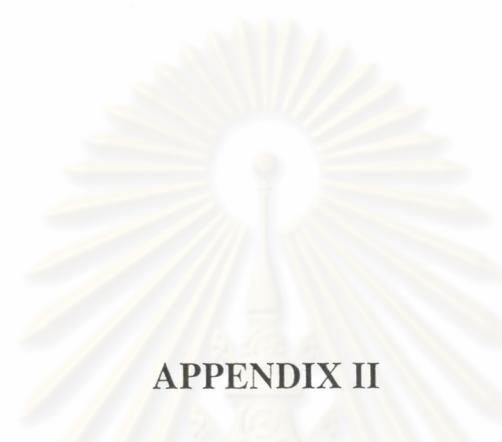
$$N = 5.54 (t_r / w_{0.5})^2$$

where L is the length of the column (millimeters). Thus, the smaller the height equivalent to a theoretical plate (HETP, or H), the greater is the efficiency of the column. In general, the H value is smaller for small stationary phase particle sizes, low mobile phase flow rates, less viscous mobile phases, higher separation temperature and smaller solute molecular sizes.

The most effective way to alter resolution is to change the selectivity or the capacity of the column. The effect of increasing the efficiency of the column by increasing the column length or flow-rate velocity is less significant, as resolution increase proportionally as the square root of the number theoretical plates. If increased resolution is required, a column with a higher capacity factor is the best choice. However, increasing the capacity factor would increase the analysis time, so a compromise must be reached between resolution and analysis time.



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APPENDIX II

ISOFLAVONE HPLC FINGERPRINTS OF REFERENCE STANDARD AND *P. mirifica*

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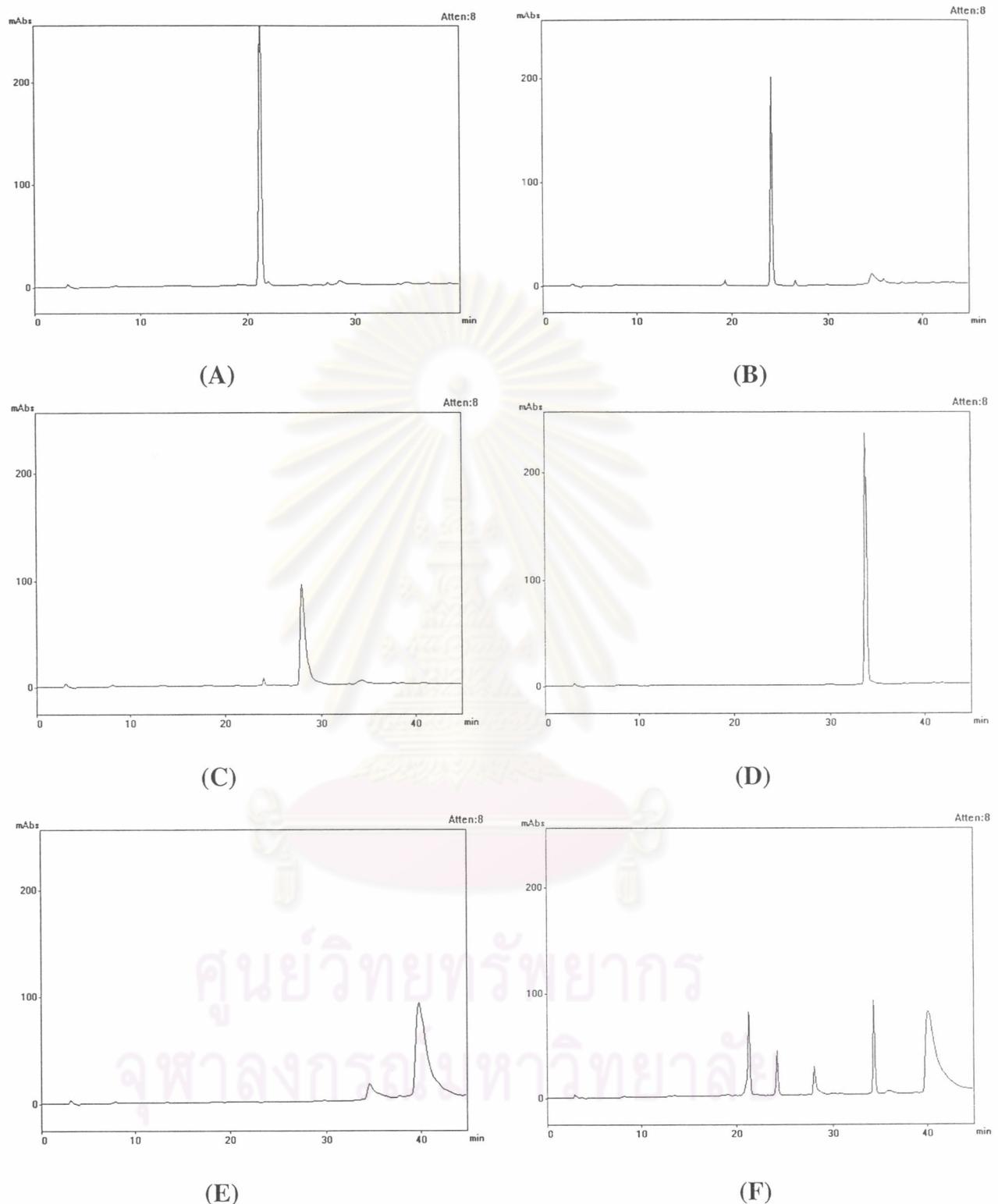


Figure 43 Isoflavone HPLC fingerprint of reference standard (A) Puerarin (B) Daidzin (C) Genistin (D) Daidzein (E) Genistein (F) mixed standard
1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

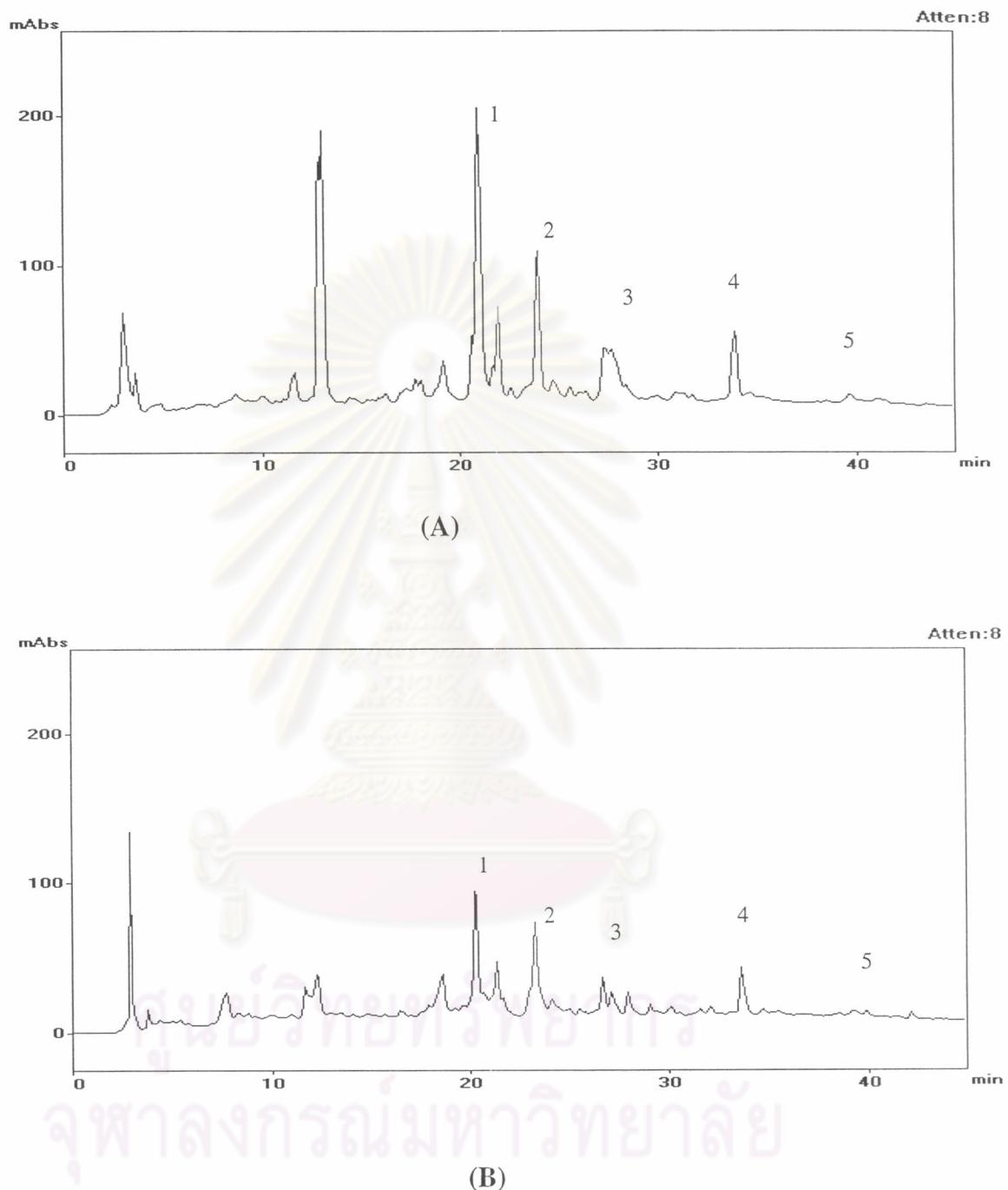


Figure 44 Isoflavone HPLC fingerprint of wild *P. mirifica* clone Chiang Dao at wavelength (A) 254 and (B) 280 nm

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

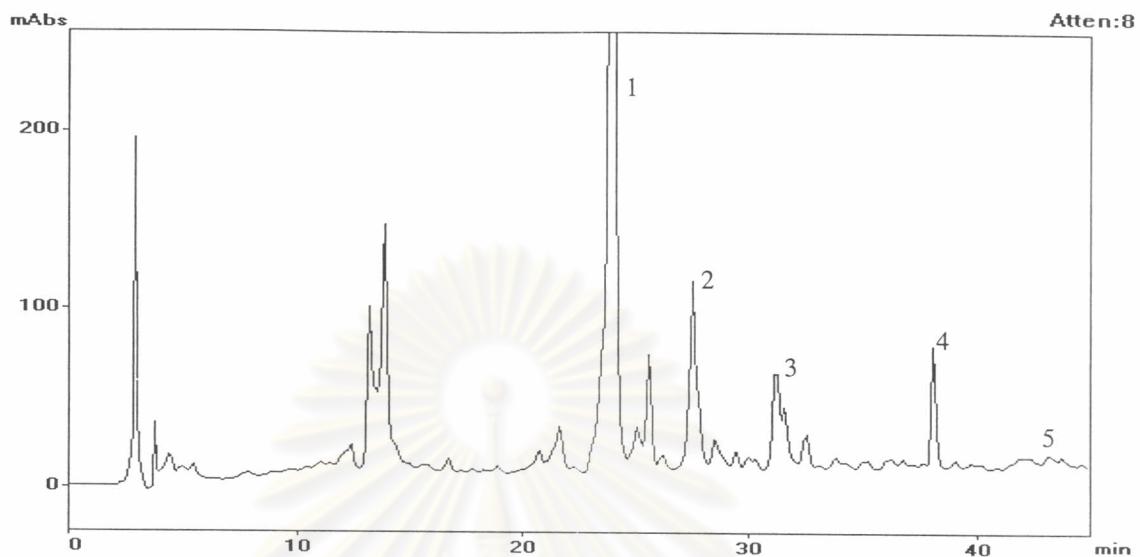


Figure 45 Isoflavone HPLC fingerprint of *P. mirifica* clone Chiang Dao spike with puerarin

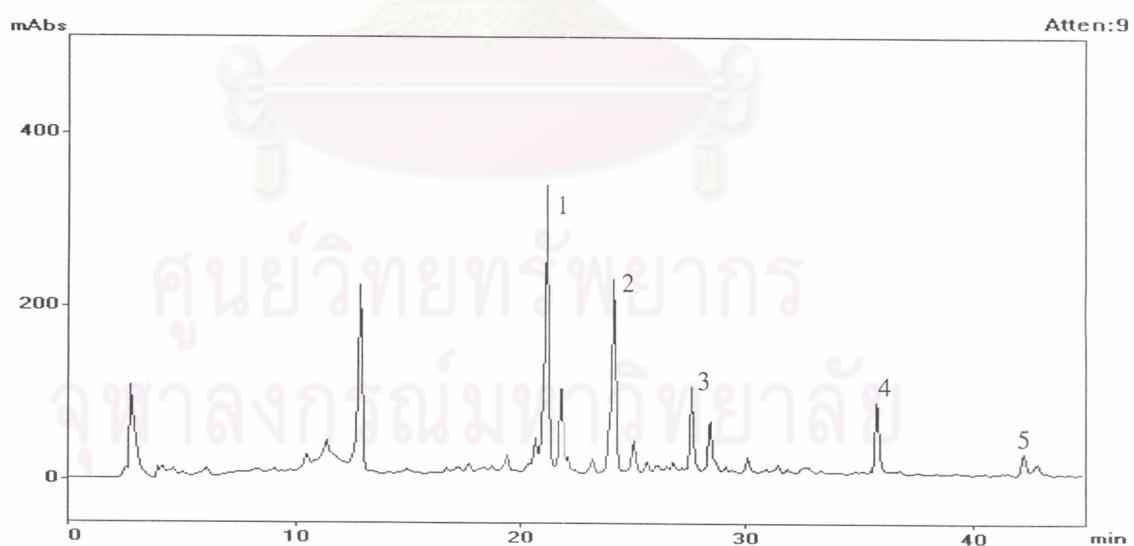


Figure 46 Isoflavone HPLC fingerprint of *P. mirifica* clone Chiang Dao spike with daidzin

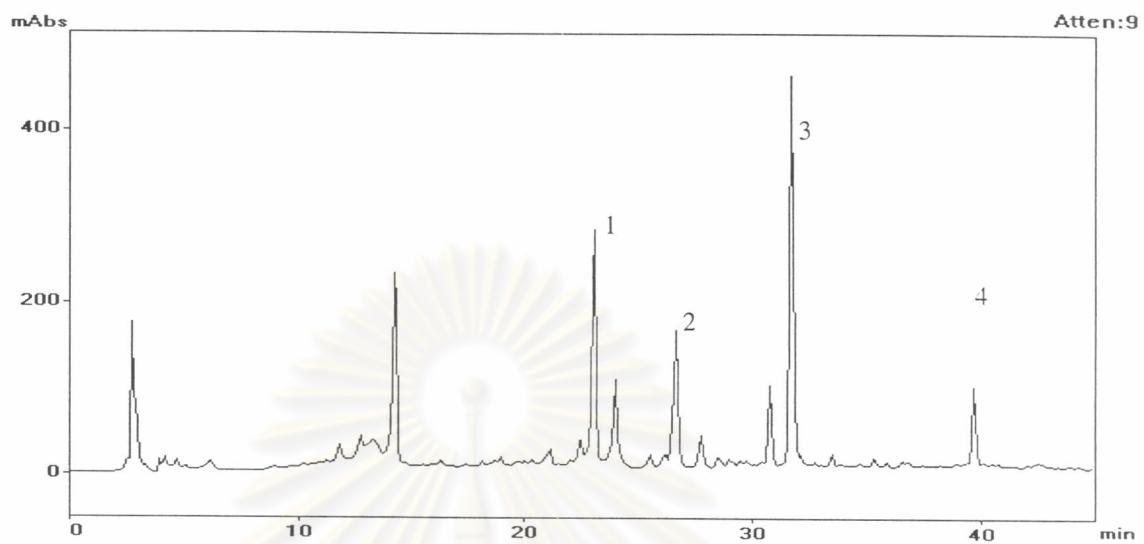


Figure 47 Isoflavone HPLC fingerprint of *P. mirifica* clone Chiang Dao spike with genistin

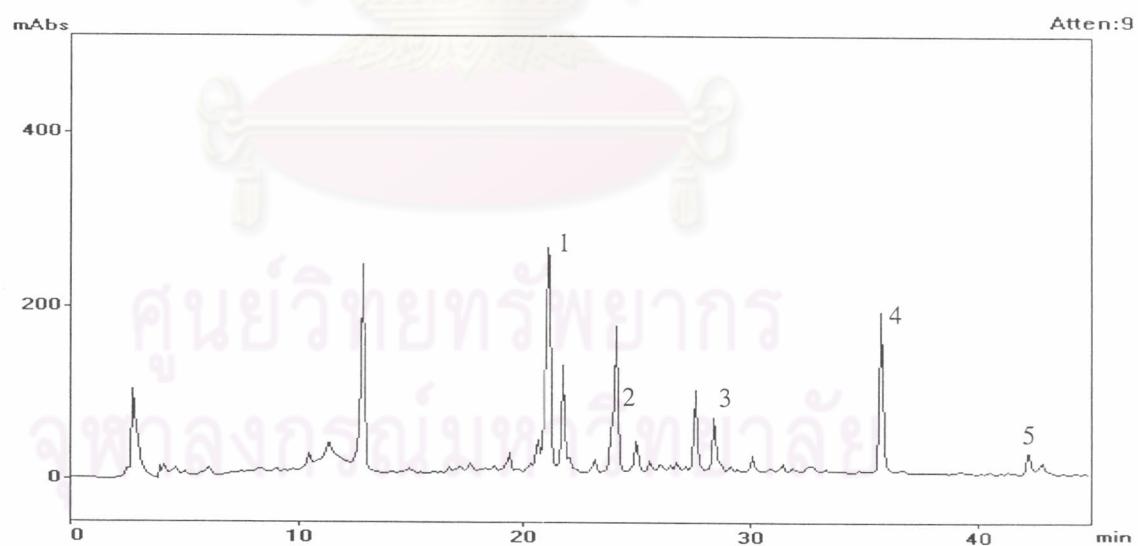


Figure 48 Isoflavone HPLC fingerprint of *P. mirifica* clone Chiang Dao spike with daidzein

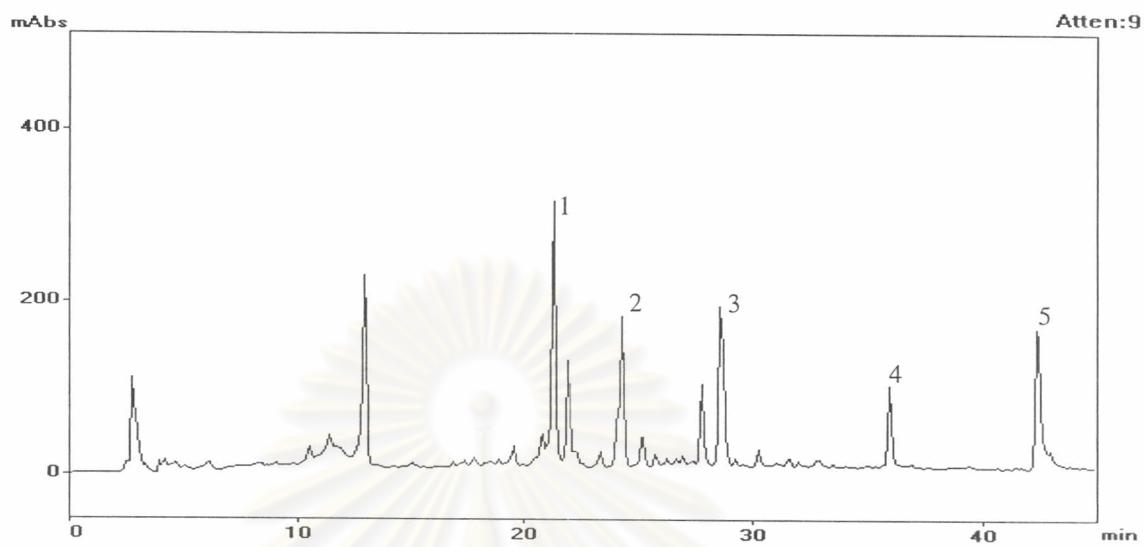


Figure 49 Isoflavone HPLC fingerprint of *P. mirifica* clone Chiang Dao spike with genistein

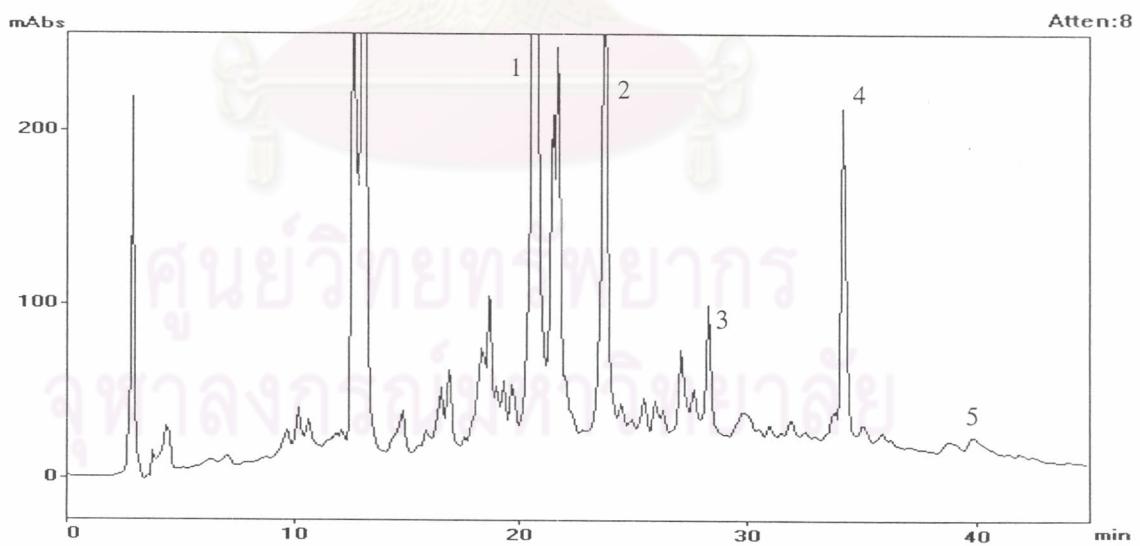


Figure 50 Isoflavone HPLC fingerprint of wild *P. mirifica* clone Chiang Dao, Spray Dry preparation

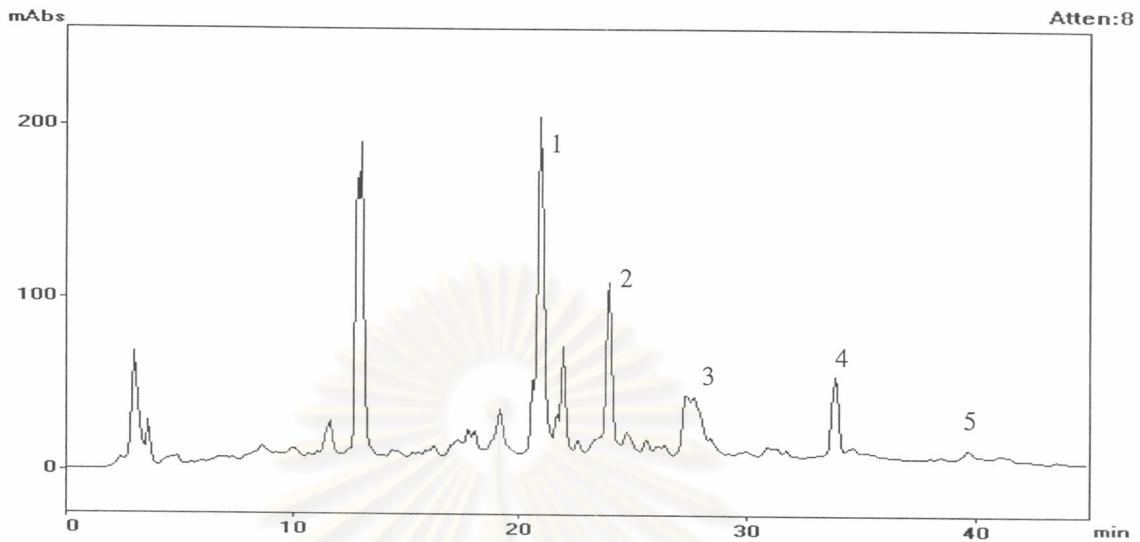


Figure 51 Isoflavone HPLC fingerprint of wild *P. mirifica* clone Chiang Dao extracted in methanol

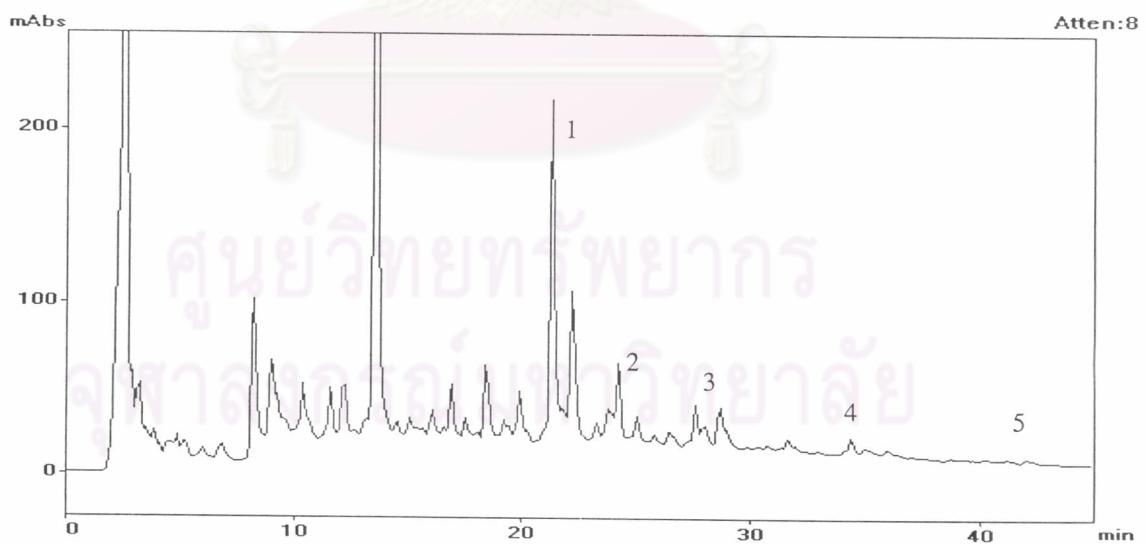


Figure 52 Isoflavone HPLC fingerprint of wild *P. mirifica* clone Chiang Dao extracted in water

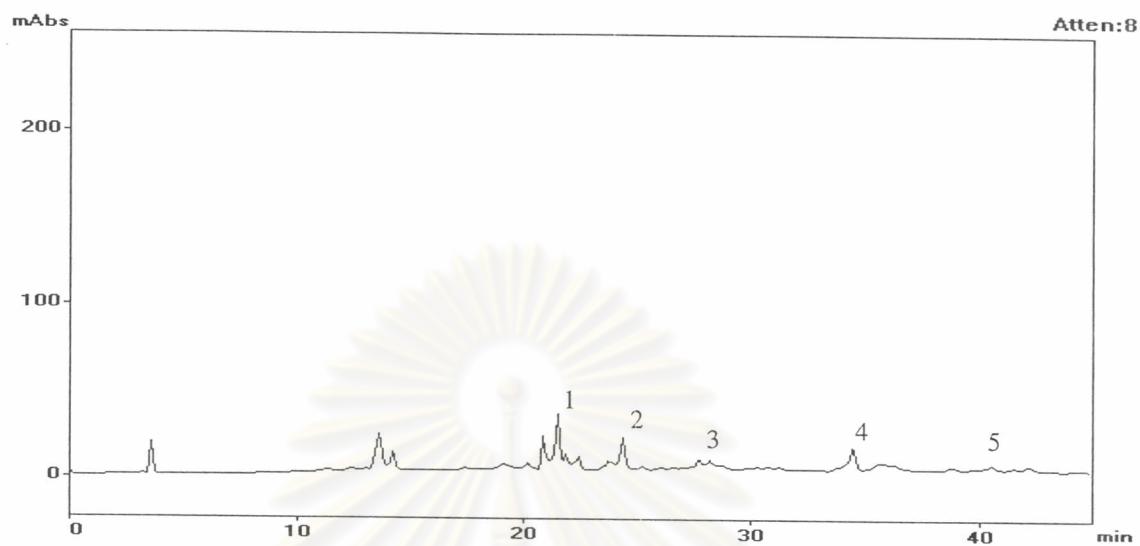


Figure 53 Isoflavone HPLC fingerprint of wild *P. mirifica* clone Chiang Dao extracted in ethanol

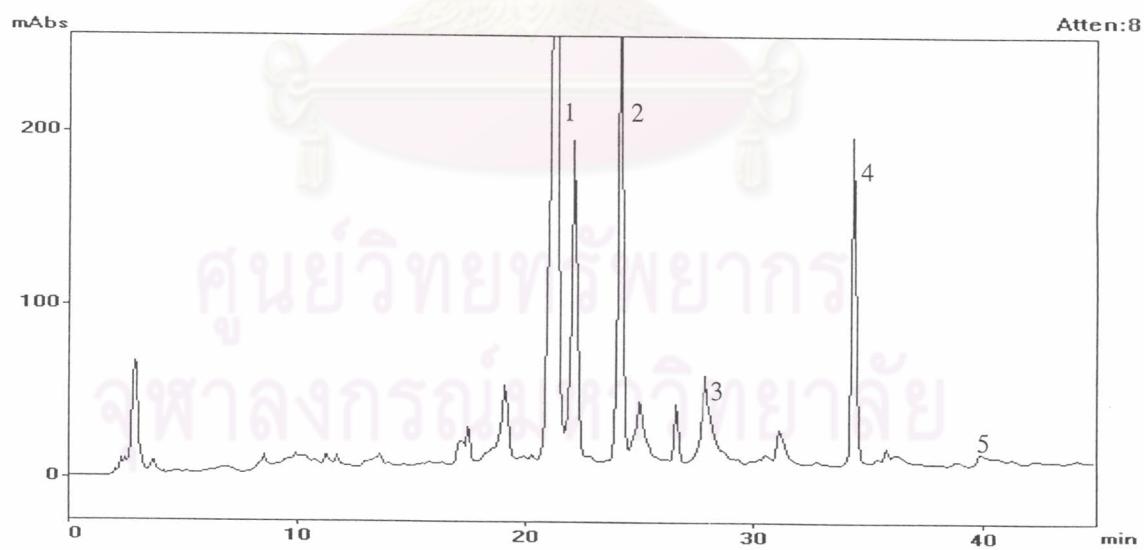


Figure 54 Isoflavone HPLC fingerprint of *P. lobata* collected from China

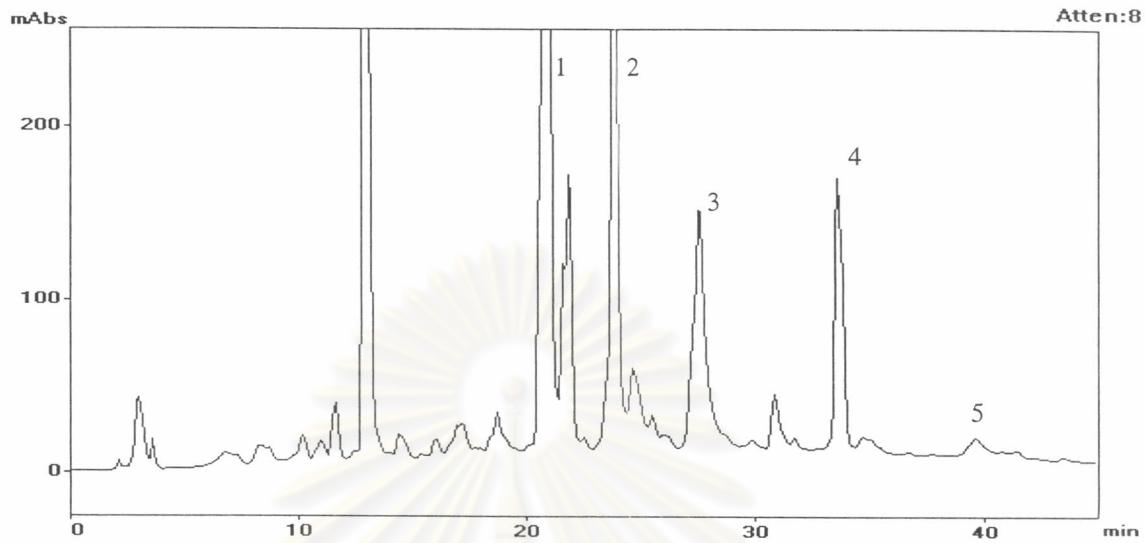


Figure 55 Isoflavone HPLC fingerprint of *P. mirifica* collected in Kanchanaburi province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

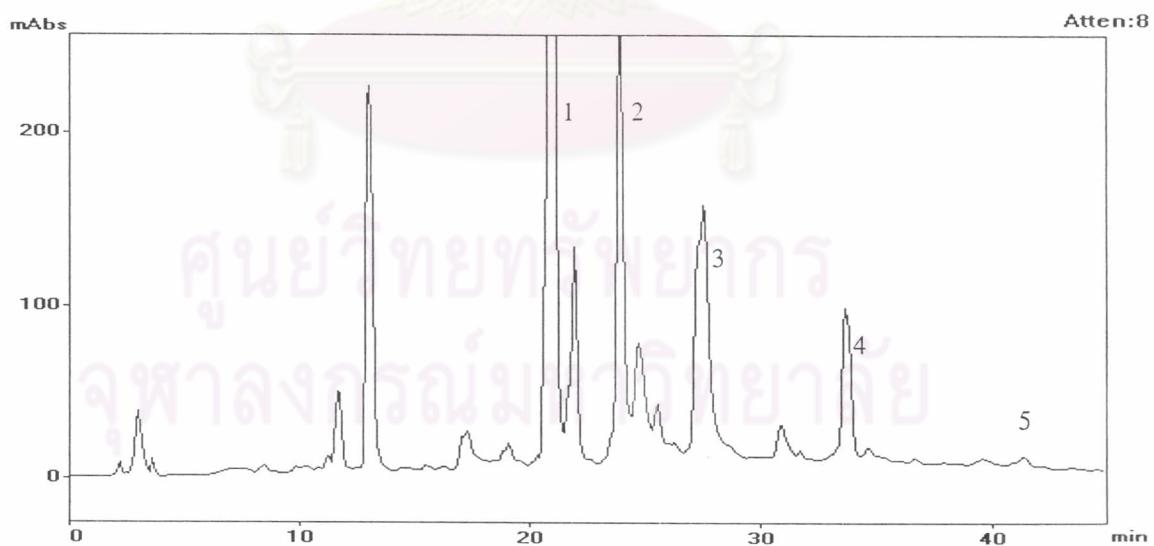


Figure 56 Isoflavone HPLC fingerprint of *P. mirifica* collected in Lamphun province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

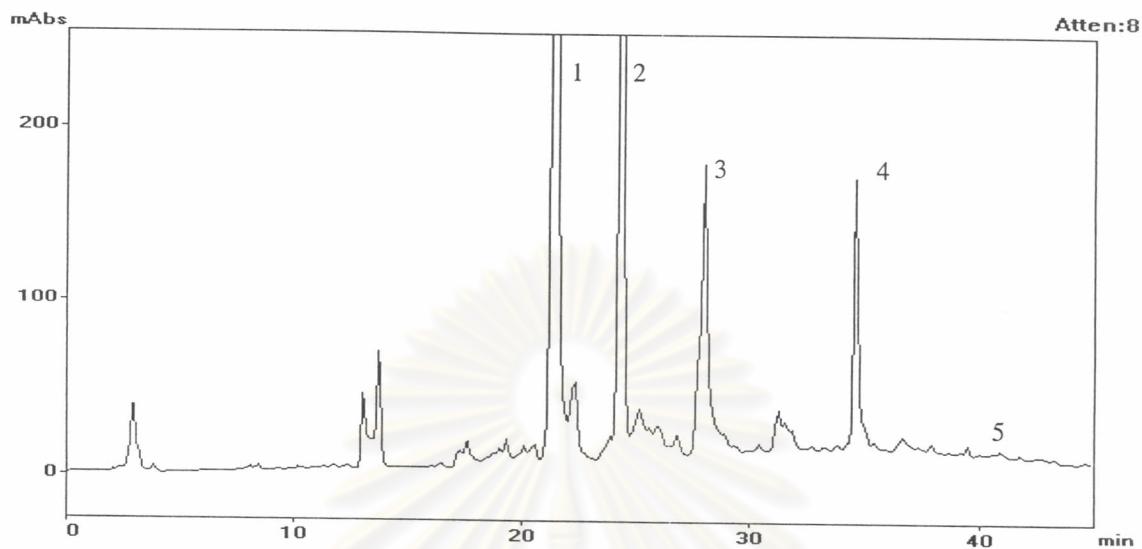


Figure 57 Isoflavone HPLC fingerprint of *P. mirifica* collected in Chiang Mai province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

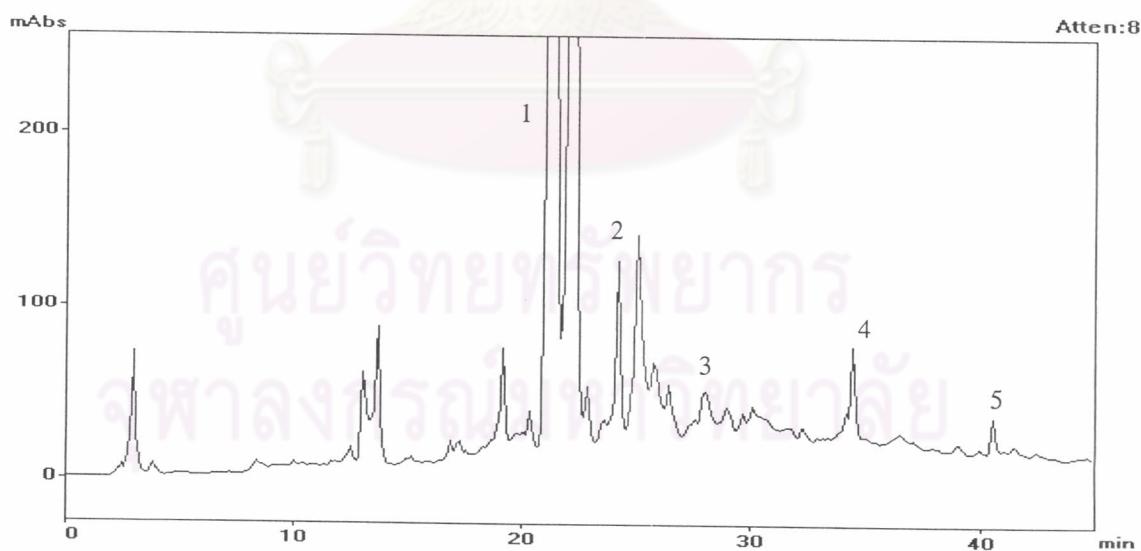


Figure 58 Isoflavone HPLC fingerprint of *P. mirifica* collected in Sakon Nakhon province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

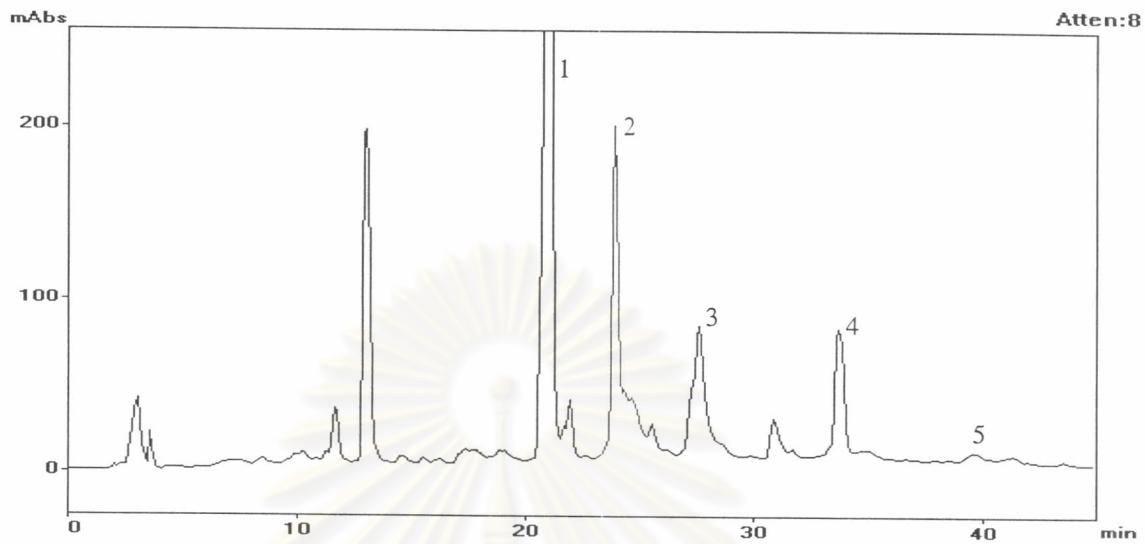


Figure 59 Isoflavone HPLC fingerprint of *P. mirifica* collected in Mae Hong Son province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

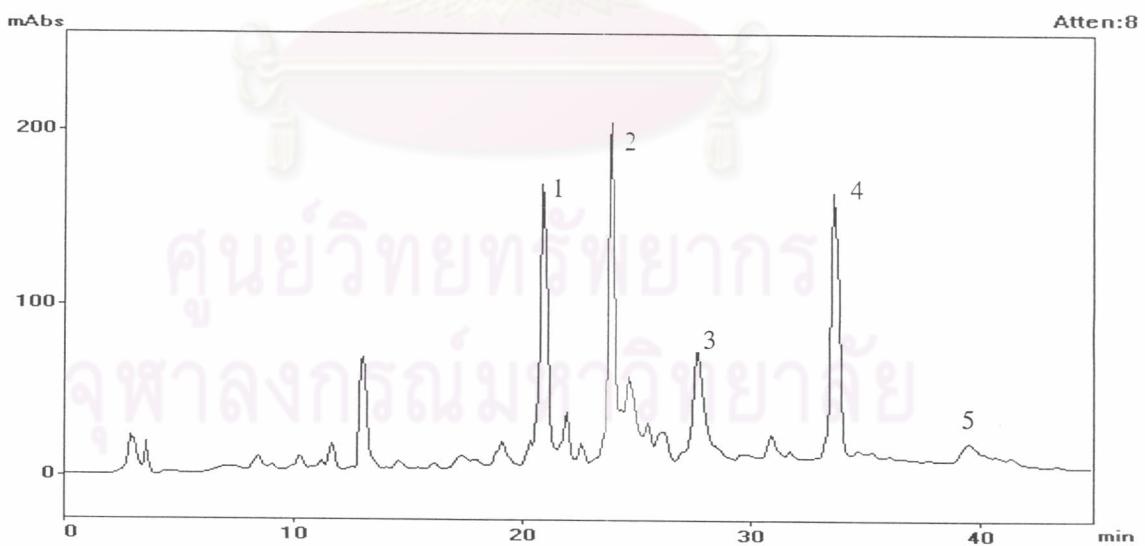


Figure 60 Isoflavone HPLC fingerprint of *P. mirifica* collected in Uthai Thani province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

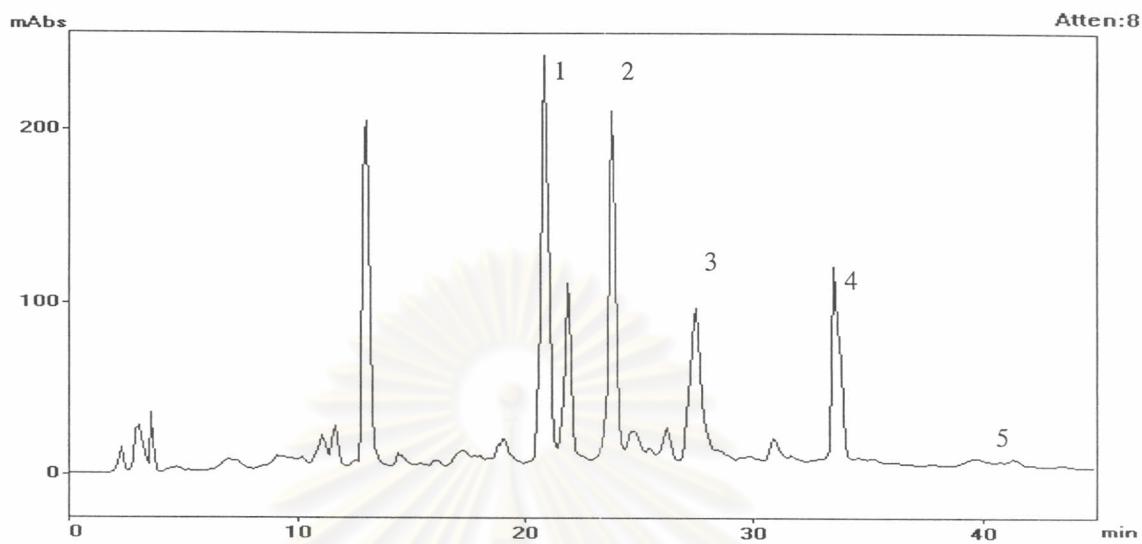


Figure 61 Isoflavone HPLC fingerprint of *P. mirifica* collected in Sukhothai province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

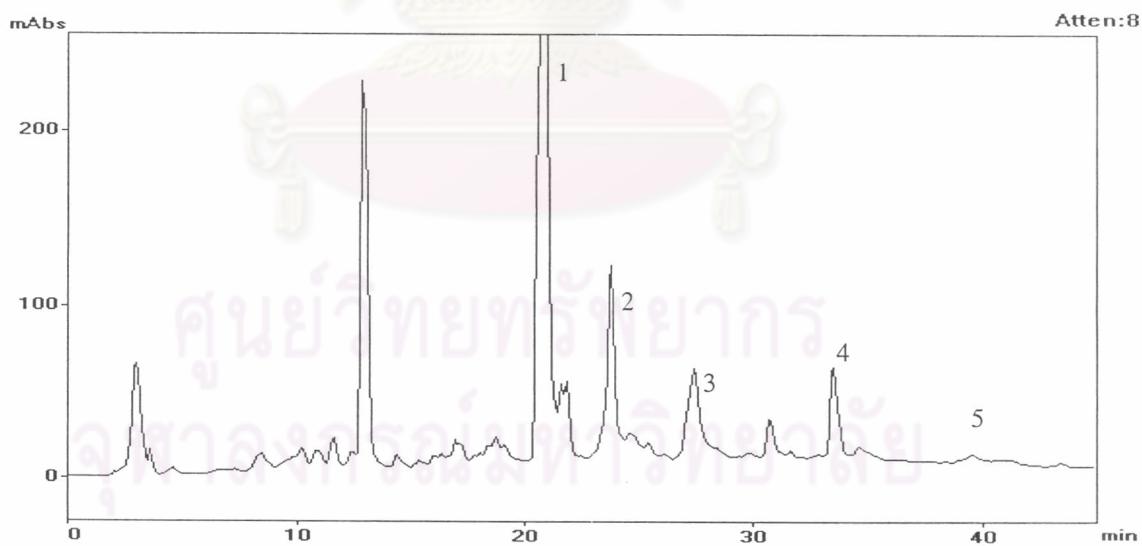


Figure 62 Isoflavone HPLC fingerprint of *P. mirifica* collected in Lampang province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

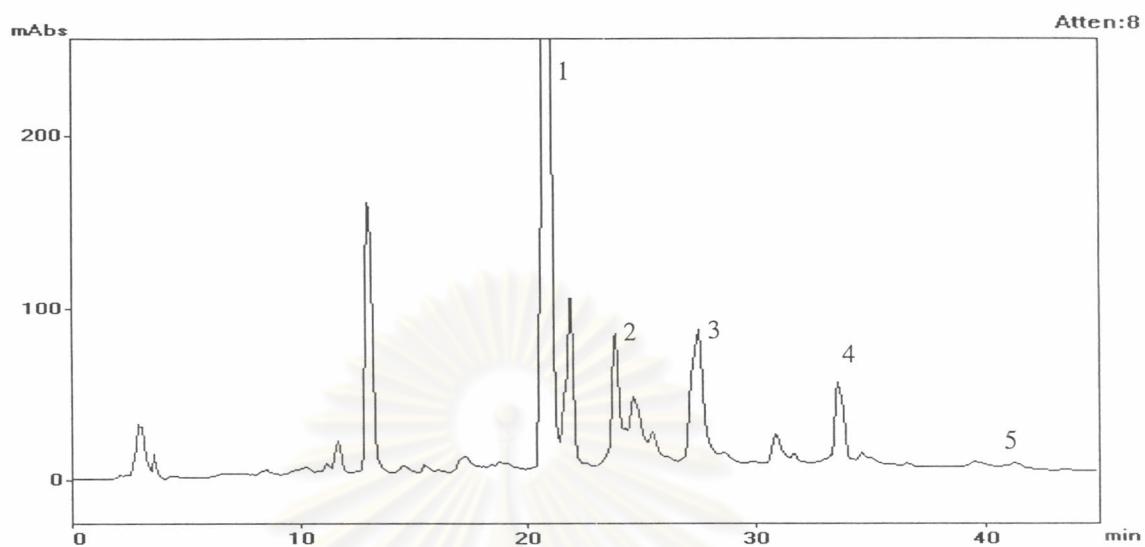


Figure 63 Isoflavone HPLC fingerprint of *P. mirifica* collected in Tak province
 1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

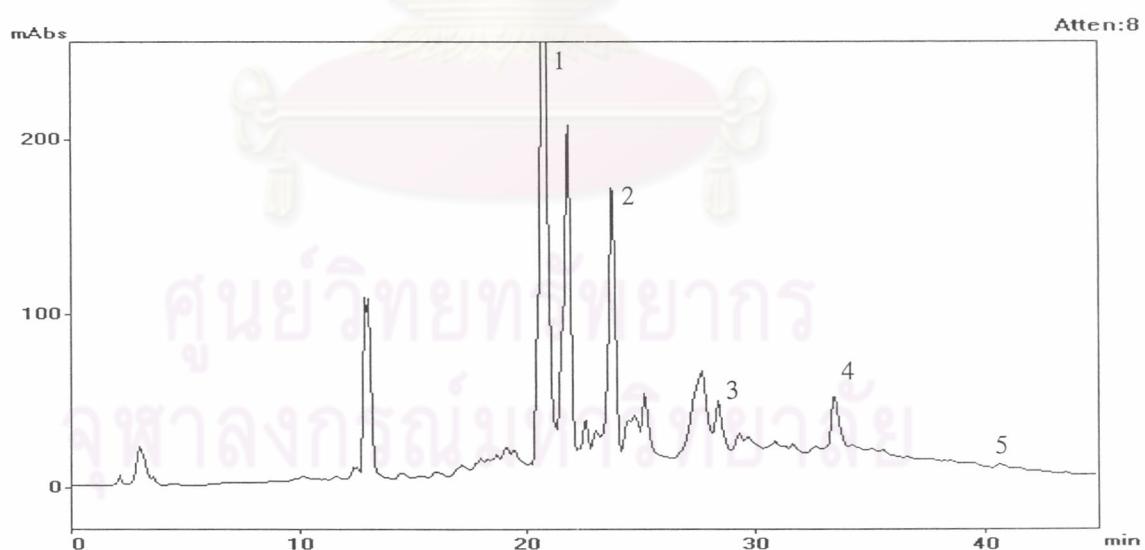


Figure 64 Isoflavone HPLC fingerprint of *P. mirifica* collected in Saraburi province
 1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

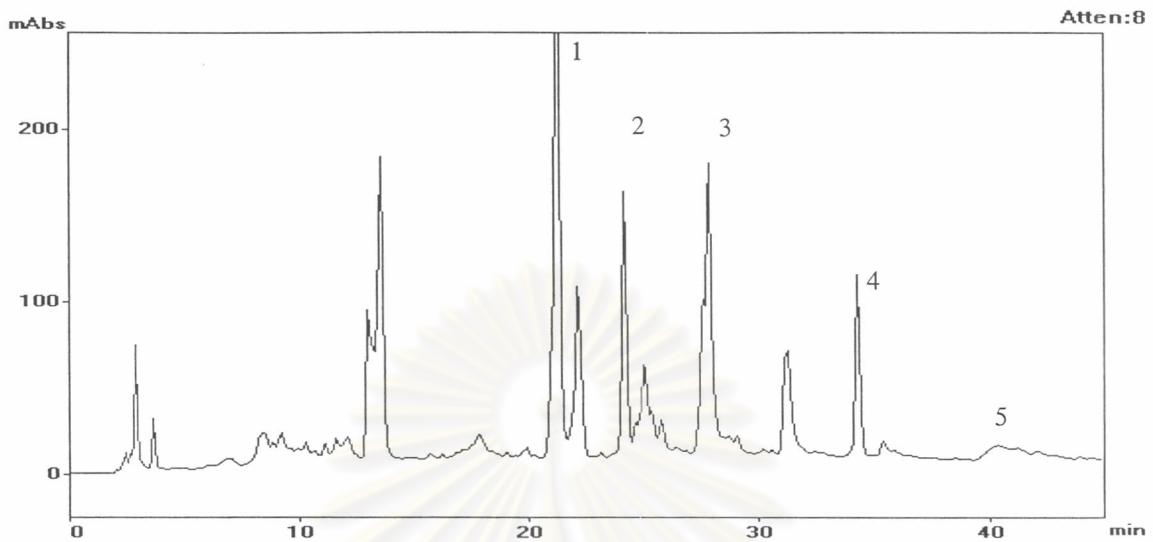


Figure 65 Isoflavone HPLC fingerprint of *P. mirifica* collected in Ratchaburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

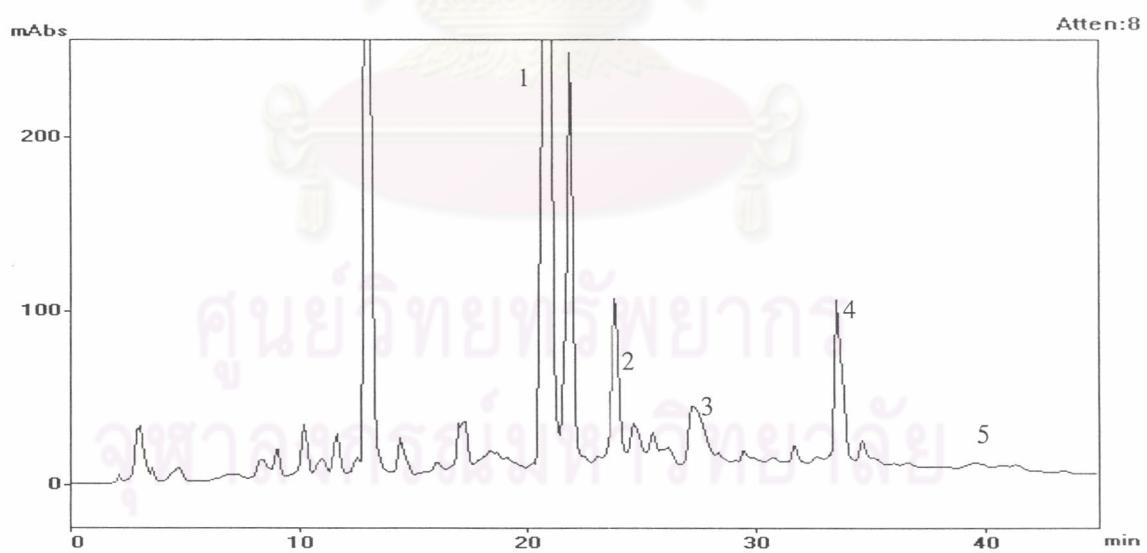


Figure 66 Isoflavone HPLC fingerprint of *P. mirifica* collected in Phitsanulok province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

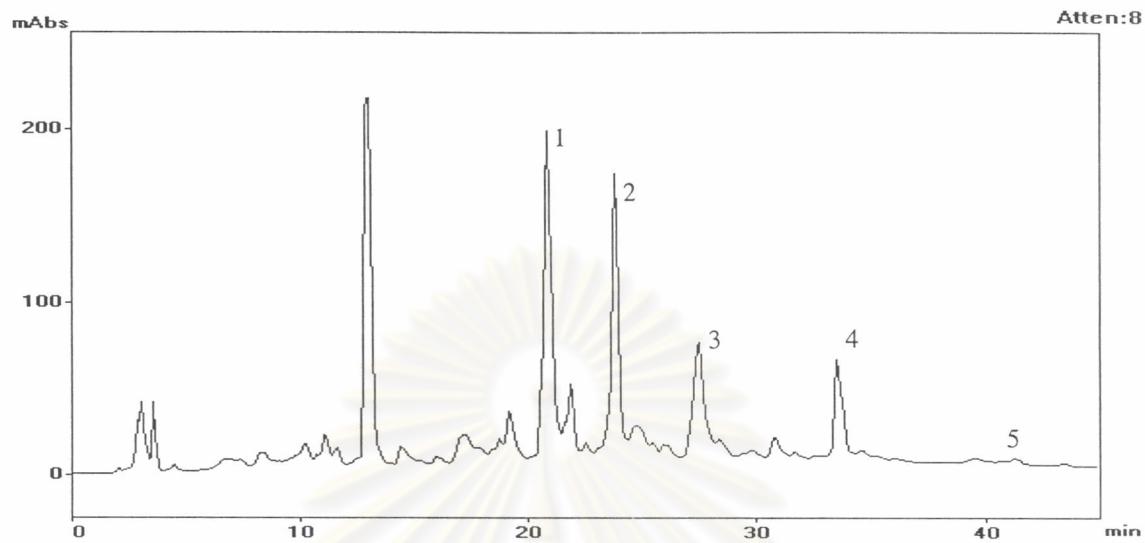


Figure 67 Isoflavone HPLC fingerprint of *P. mirifica* collected in Phetchaburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

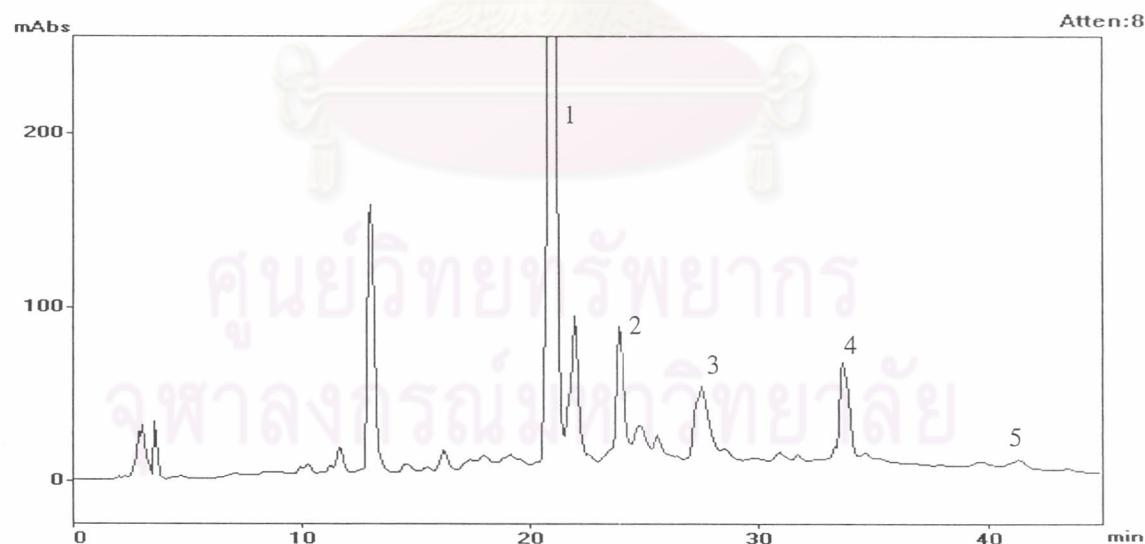


Figure 68 Isoflavone HPLC fingerprint of *P. mirifica* collected in Phrae province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

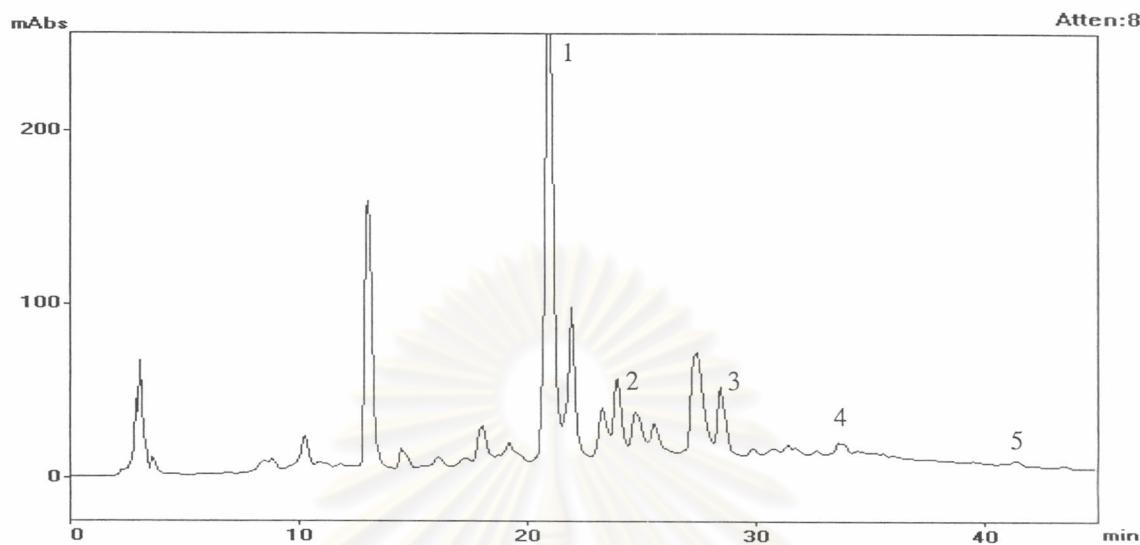


Figure 69 Isoflavone HPLC fingerprint of *P. mirifica* collected in Lop Buri province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

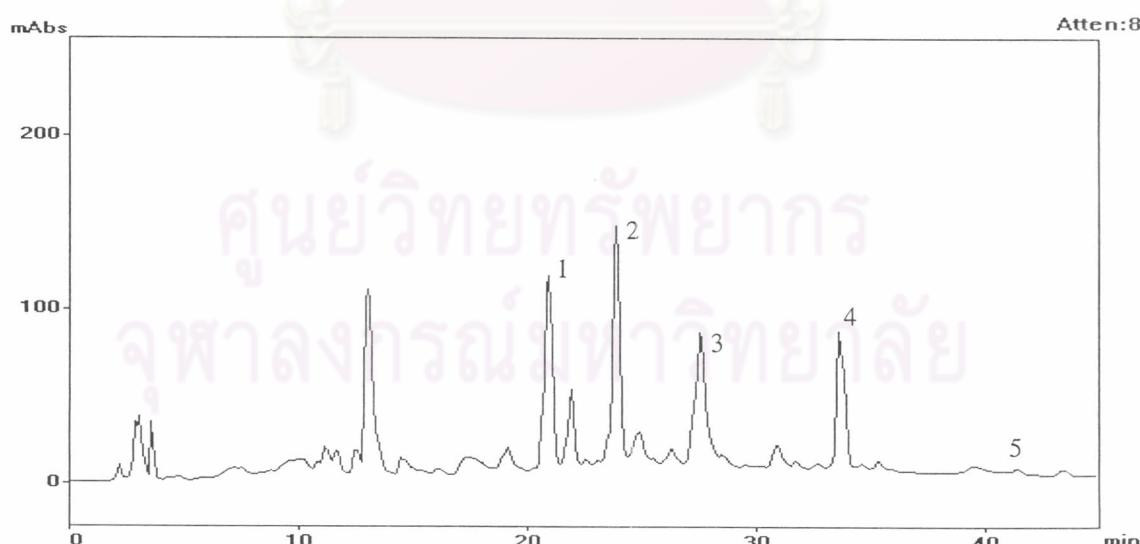


Figure 70 Isoflavone HPLC fingerprint of *P. mirifica* collected in Loei province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

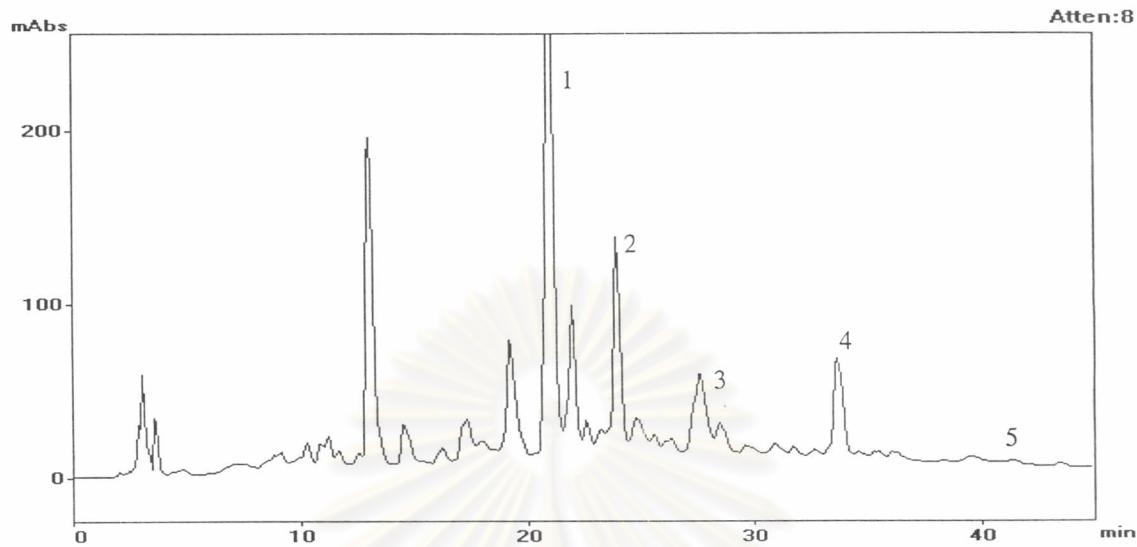


Figure 71 Isoflavone HPLC fingerprint of *P. mirifica* collected in Chaiyaphum province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

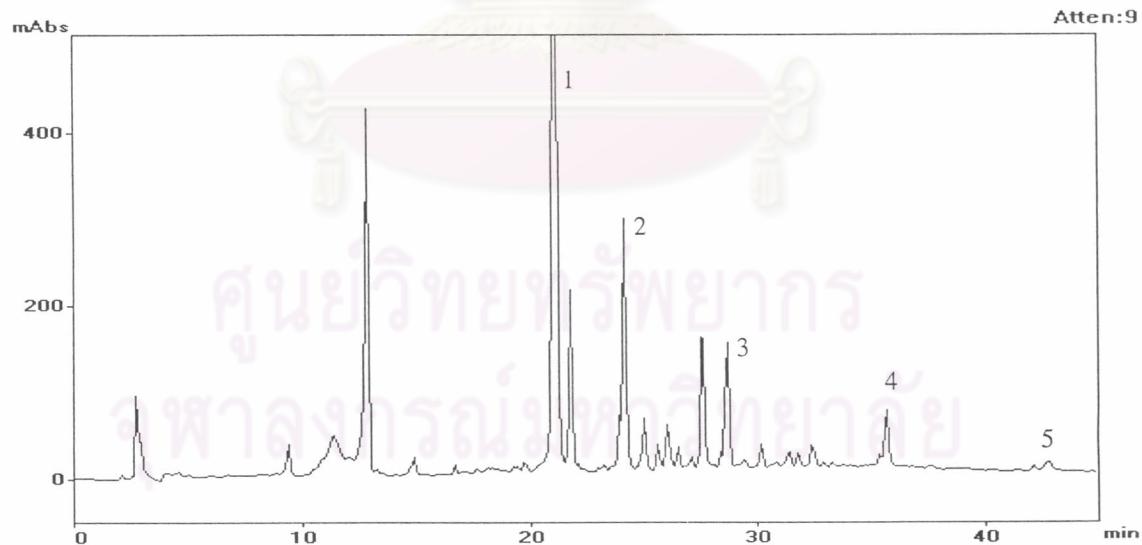


Figure 72 Isoflavone HPLC fingerprint of *P. mirifica* collected in Uttharadith province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

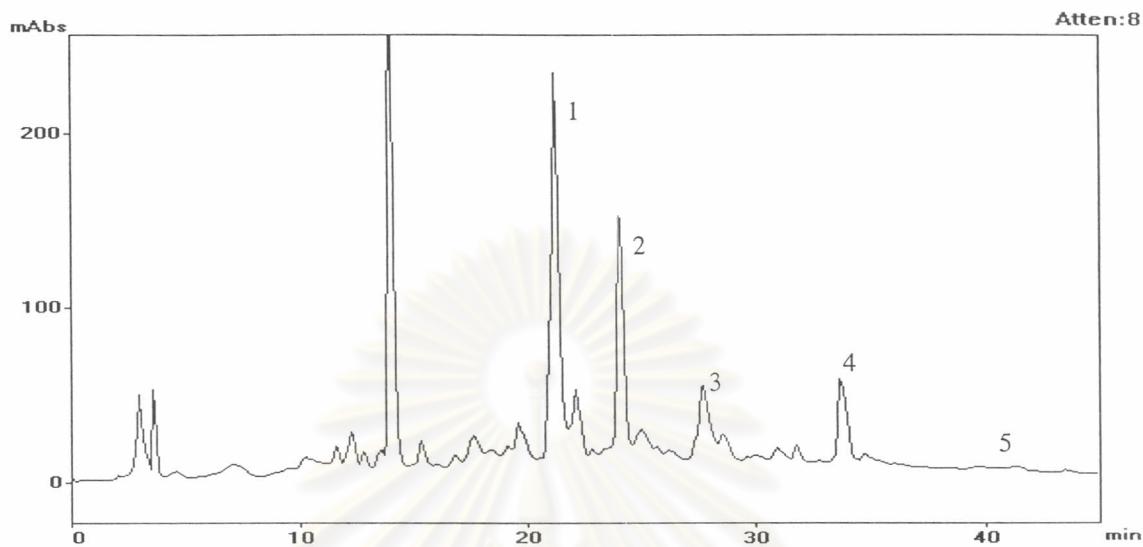


Figure 73 Isoflavone HPLC fingerprint of *P. mirifica* collected in Nakhon Sawan province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

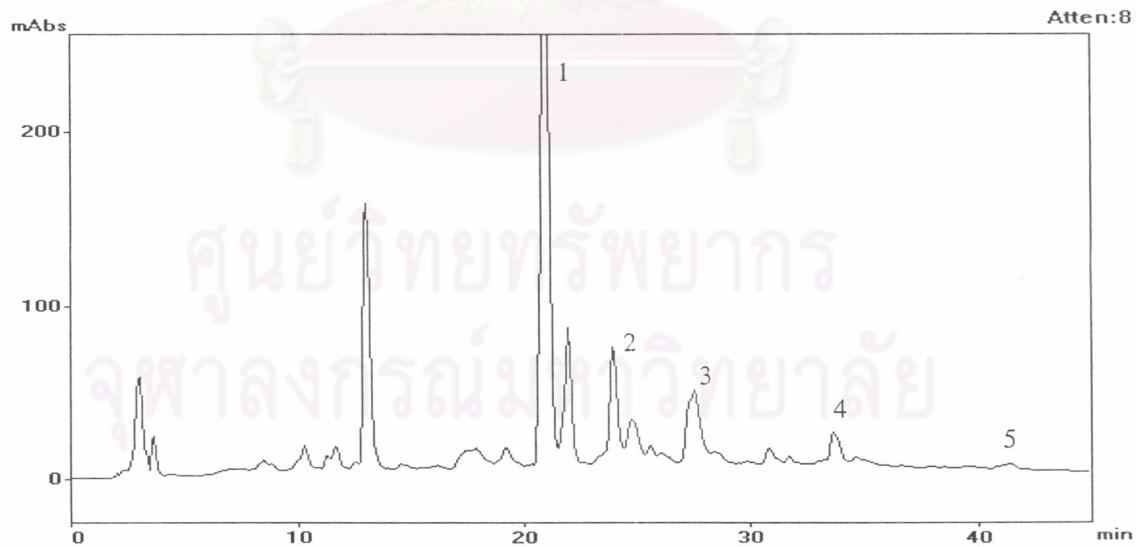


Figure 74 Isoflavone HPLC fingerprint of *P. mirifica* collected in Chiang Rai province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

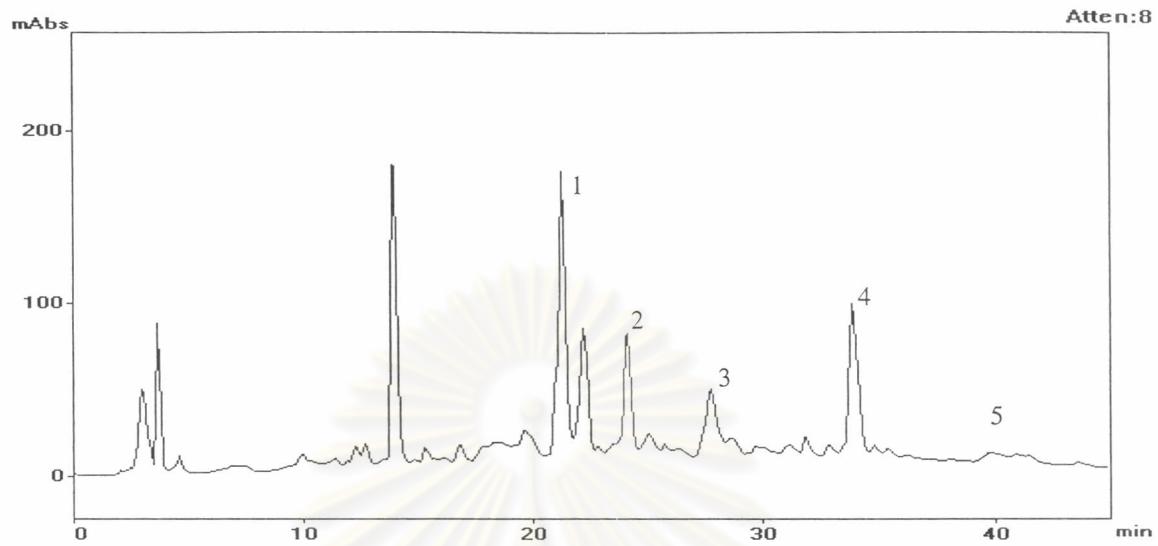


Figure 75 Isoflavone HPLC fingerprint of *P. mirifica* collected in Nong Bua - Lam Phu province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

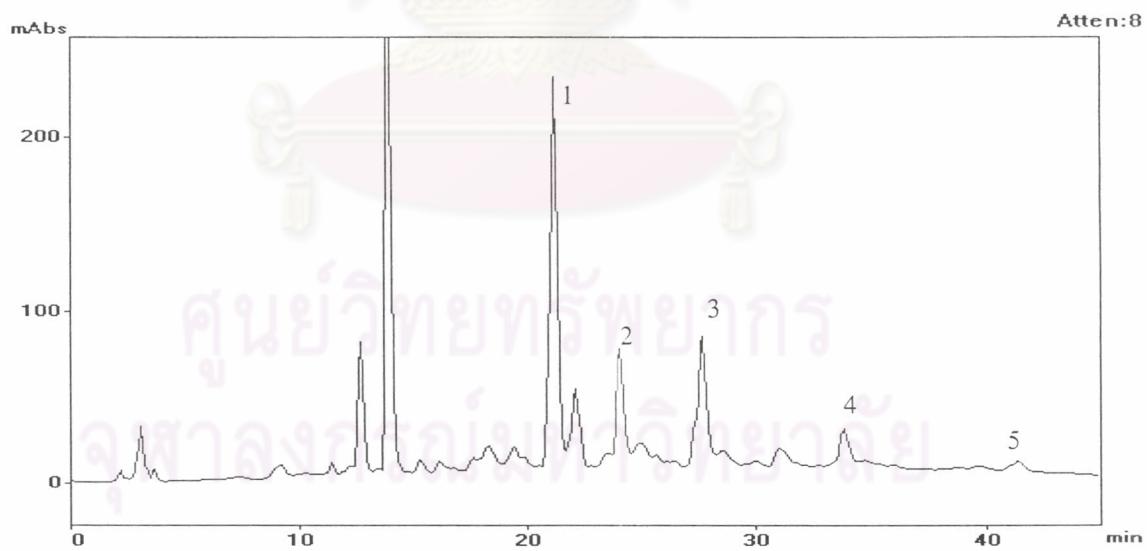


Figure 76 Isoflavone HPLC fingerprint of *P. mirifica* collected in Phayao province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

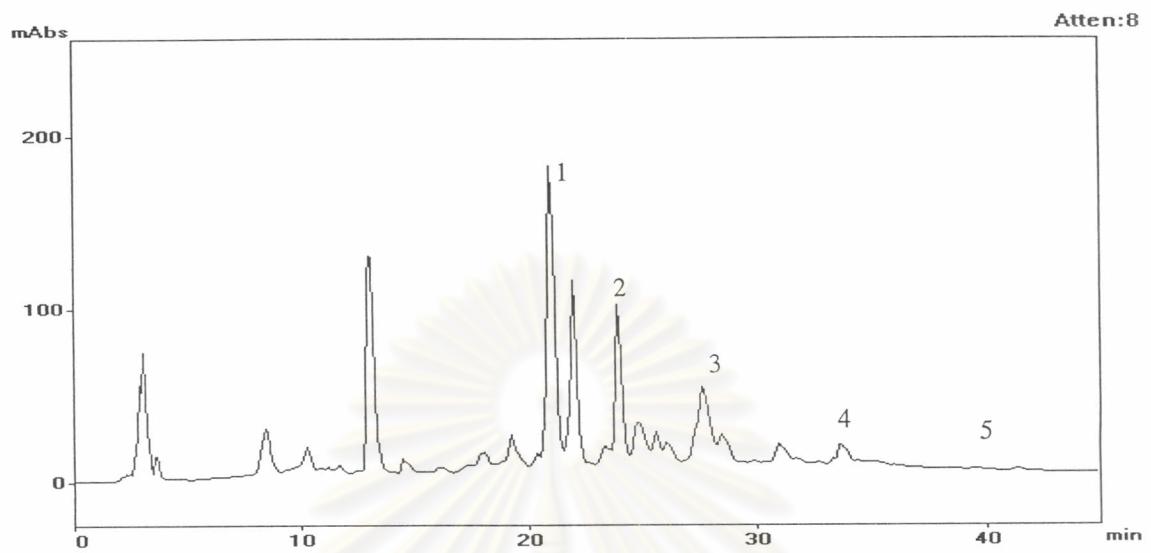


Figure 77 Isoflavone HPLC fingerprint of *P. mirifica* collected in Prachuab - Khiri Khan province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

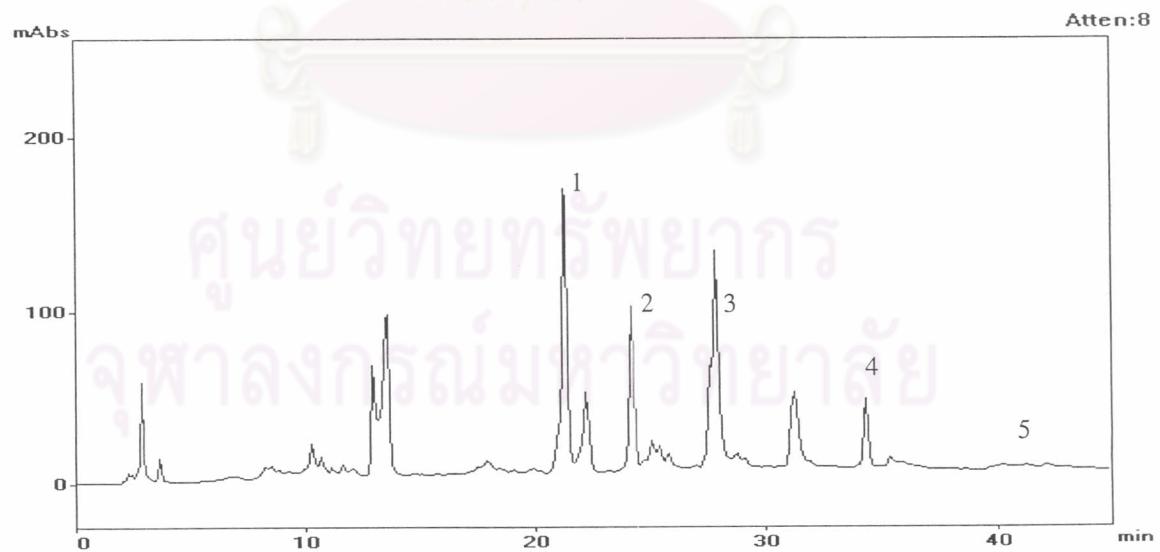


Figure 78 Isoflavone HPLC fingerprint of *P. mirifica* collected in Chumphon province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

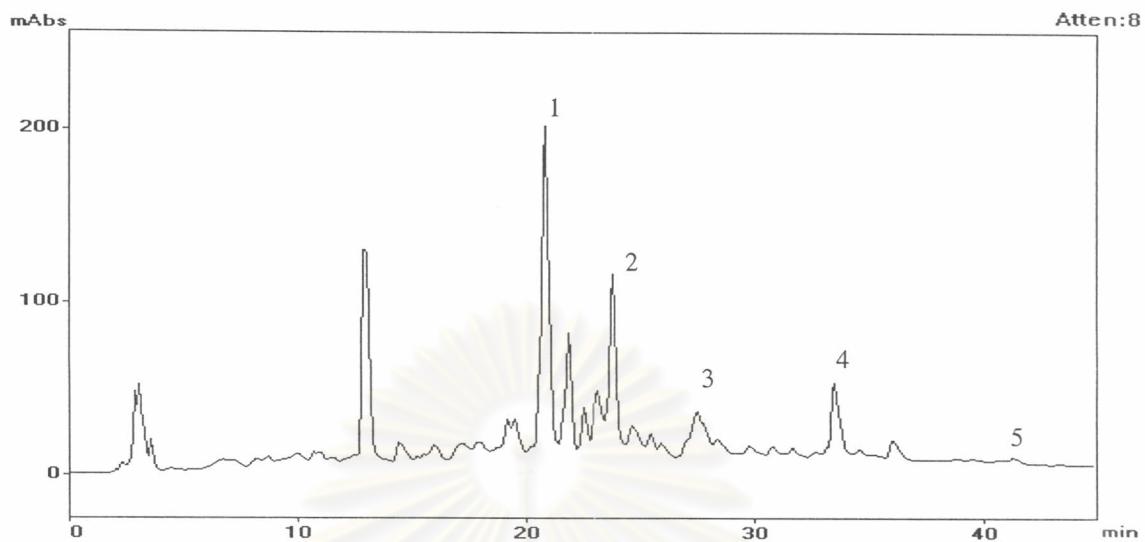


Figure 79 Isoflavone HPLC fingerprint of *P. mirifica* collected in Prachin Buri province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

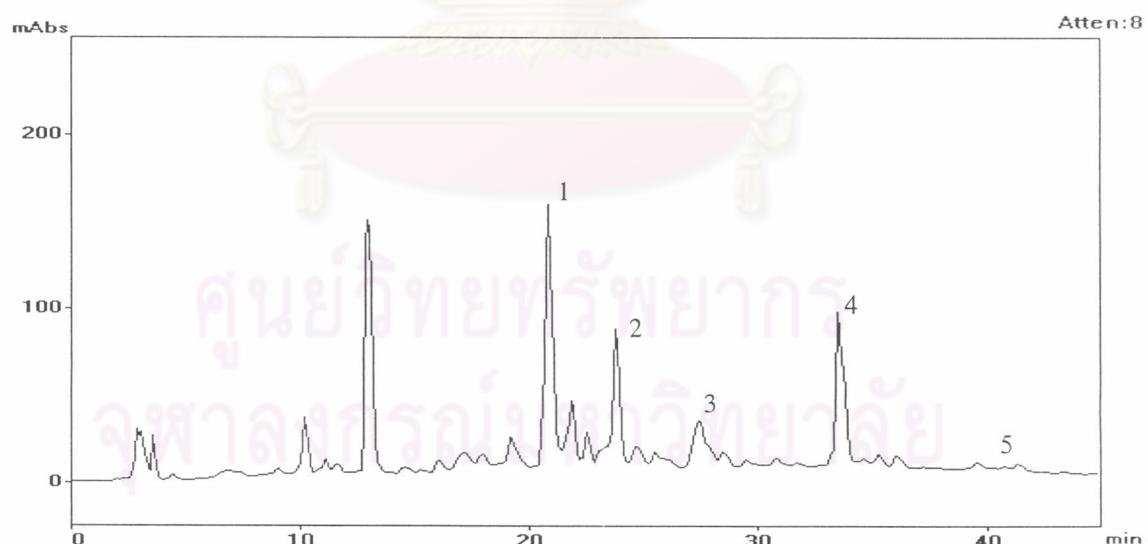


Figure 80 Isoflavone HPLC fingerprint of *P. mirifica* collected in Phetchabun province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

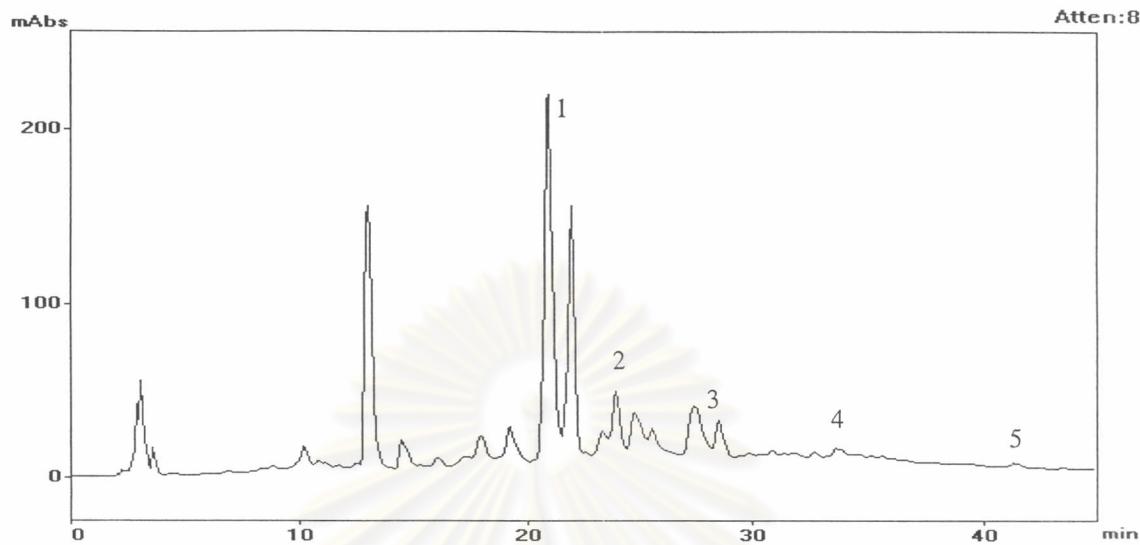


Figure 81 Isoflavone HPLC fingerprint of *P. mirifica* collected in Nakhon Ratchasima province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

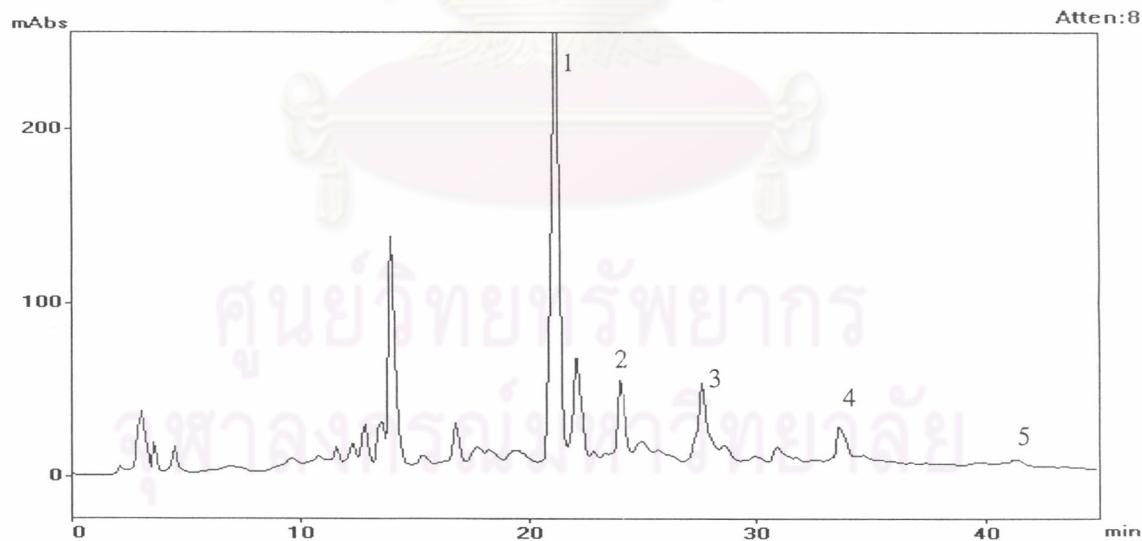


Figure 82 Isoflavone HPLC fingerprint of *P. mirifica* collected in Kamphaeng Phet province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

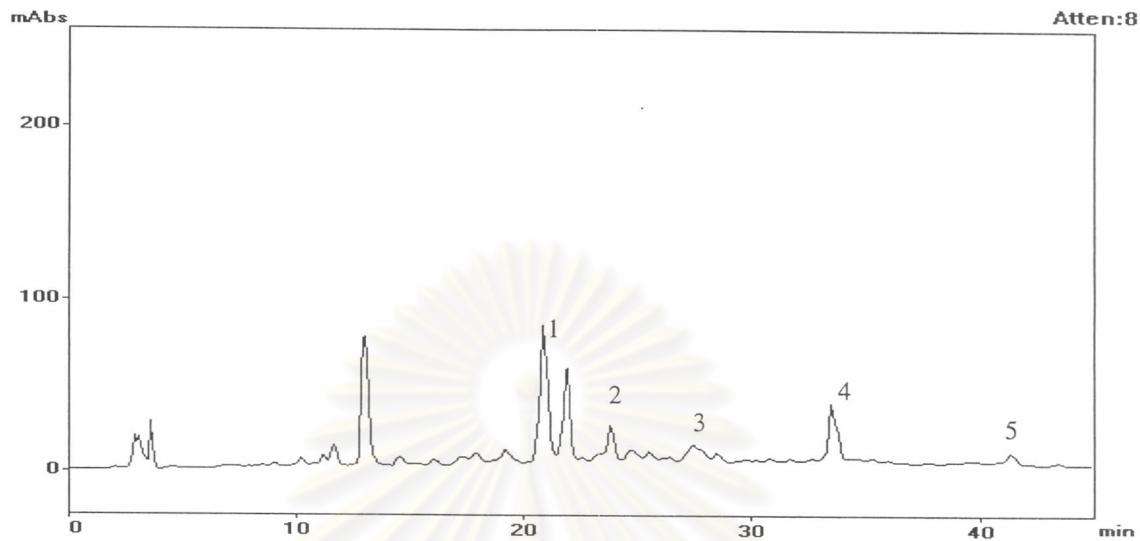


Figure 83 Isoflavone HPLC fingerprint of *P. mirifica* collected in Nan province
 1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

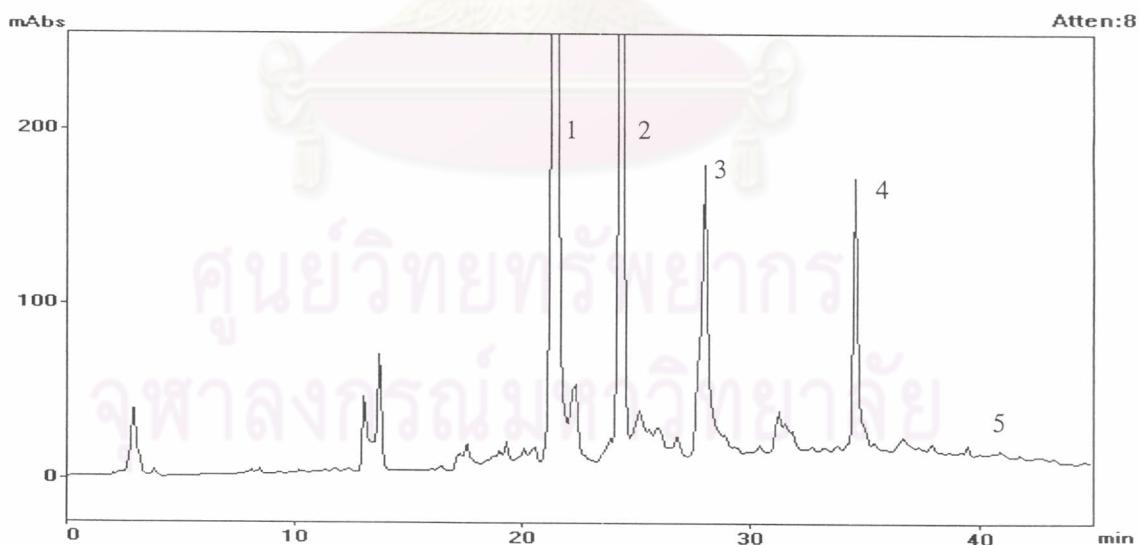


Figure 84 Isoflavone HPLC fingerprint of *P. mirifica* from Hod district in Chiang Mai province
 1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

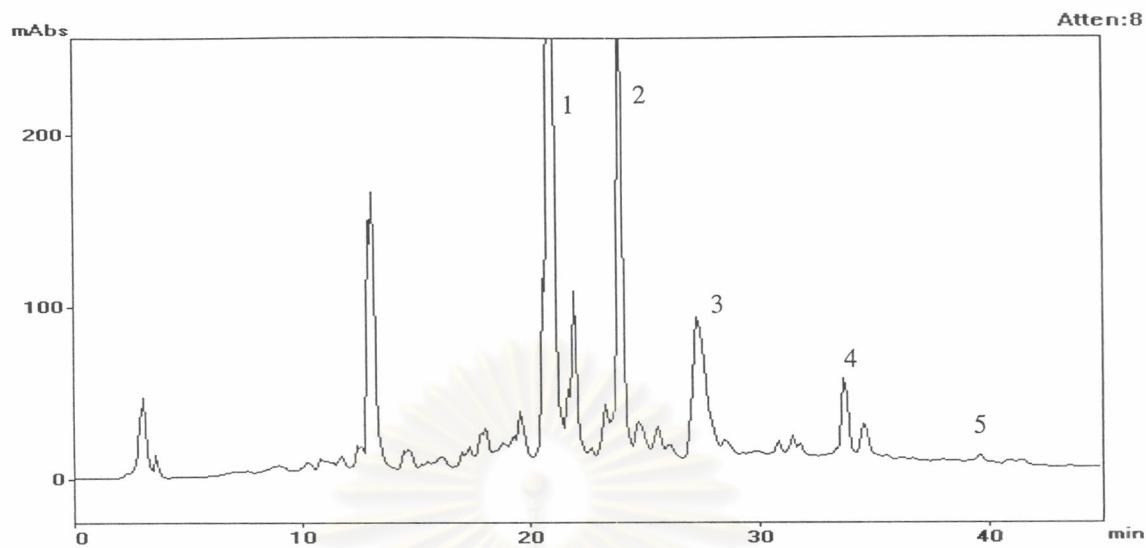


Figure 85 Isoflavone HPLC fingerprint of *P. mirifica* from Doi Tao district in Chiang Mai province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

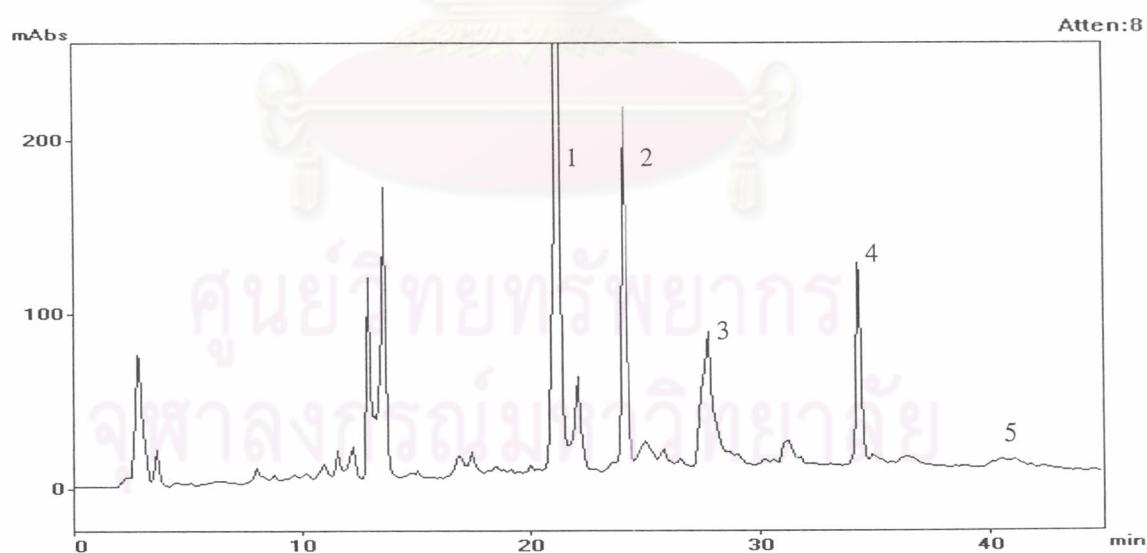


Figure 86 Isoflavone HPLC fingerprint of *P. mirifica* from Doi Saket district in Chiang Mai province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

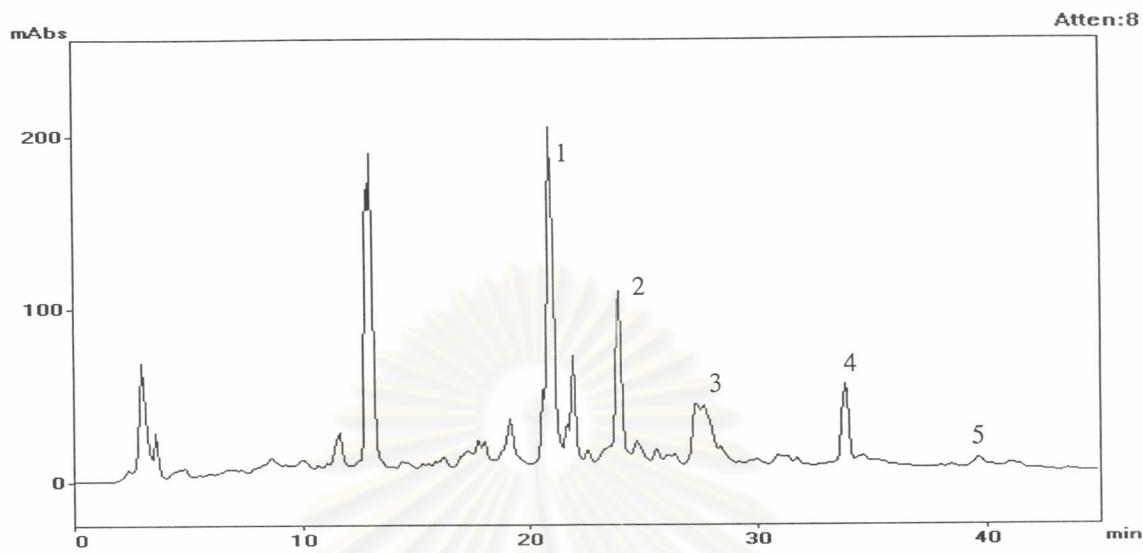


Figure 87 Isoflavone HPLC fingerprint of *P. mirifica* from Chiang Dao district in Chiang Mai province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

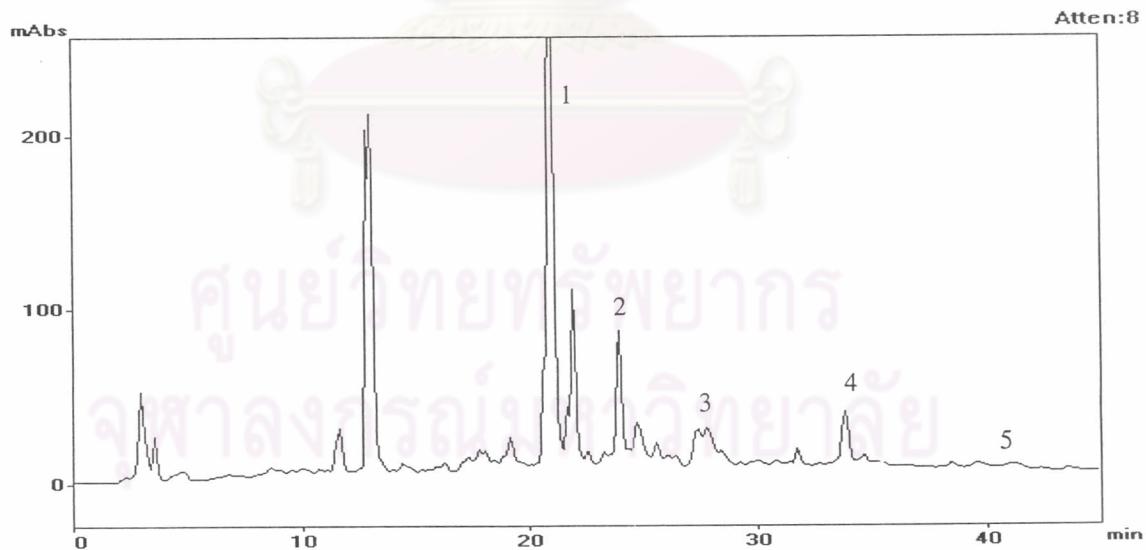


Figure 88 Isoflavone HPLC fingerprint of *P. mirifica* from Chaiprakarn district in Chiang Mai province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

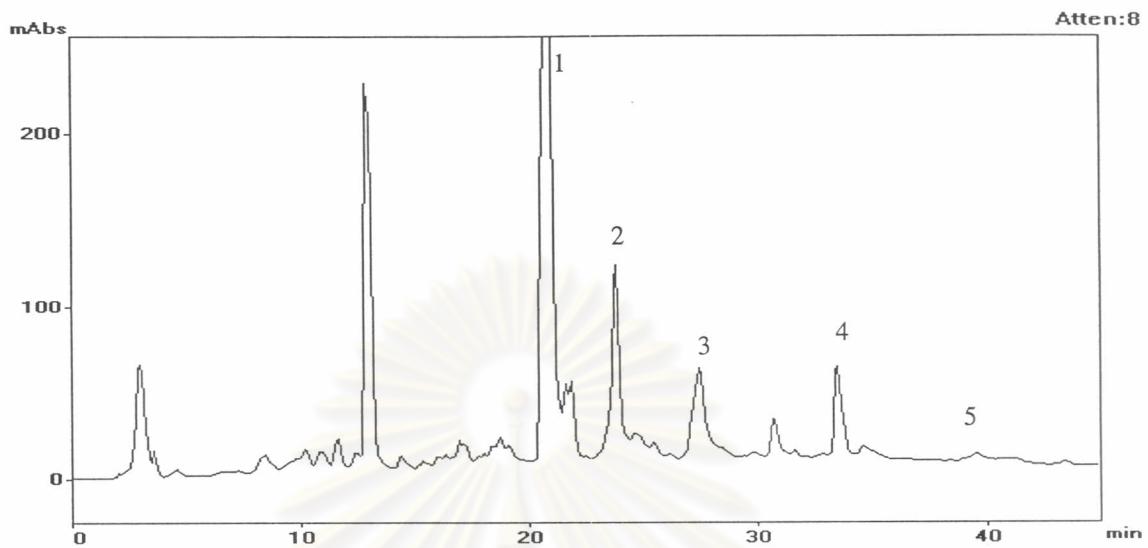


Figure 89 Isoflavone HPLC fingerprint of *P. mirifica* from Koh Ka district in Lampang province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

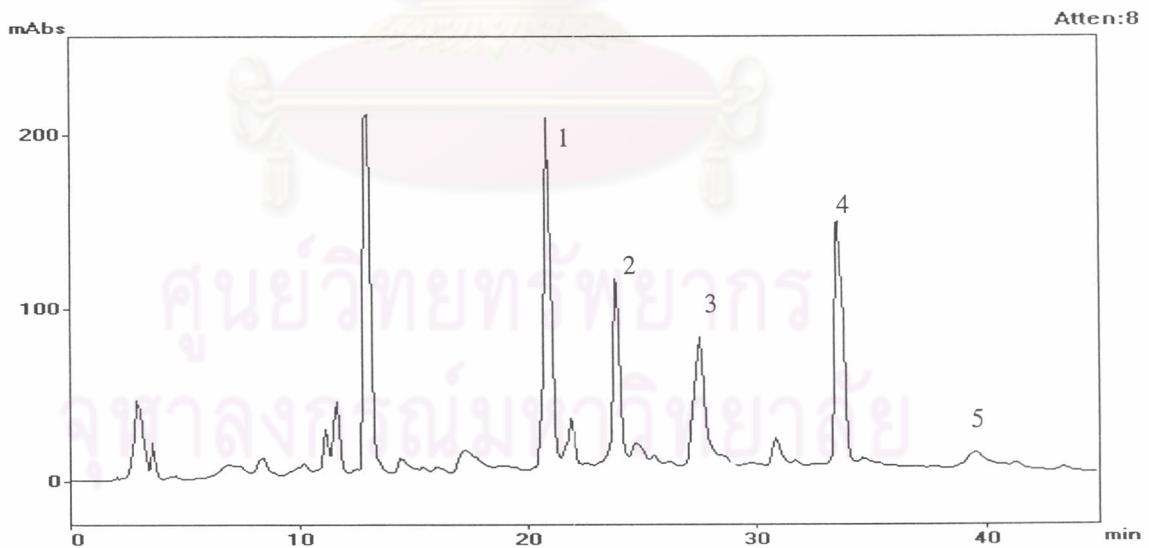


Figure 90 Isoflavone HPLC fingerprint of *P. mirifica* from Hang Chat district in Lampang province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

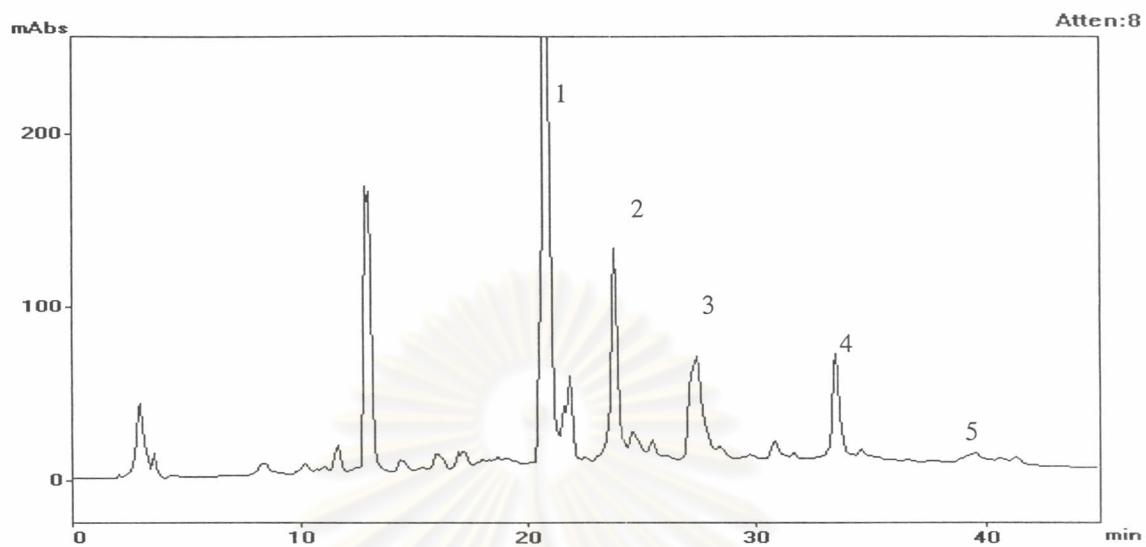


Figure 91 Isoflavone HPLC fingerprint of *P. mirifica* from Thern district in Lampang province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

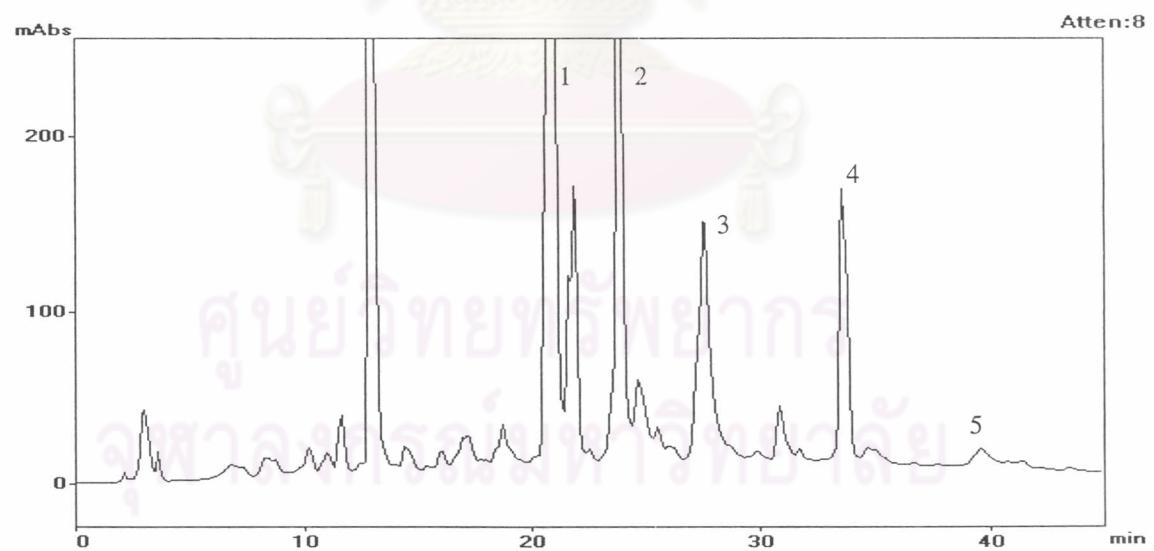


Figure 92 Isoflavone HPLC fingerprint of *P. mirifica* from Srisawat district in Kanchanaburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

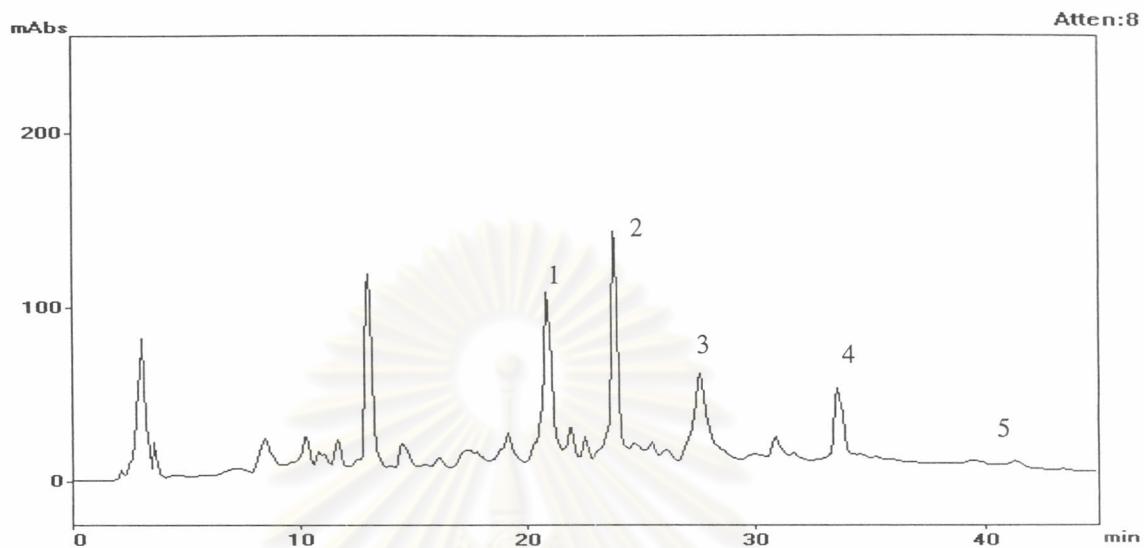


Figure 93 Isoflavone HPLC fingerprint of *P. mirifica* from Sai Yoke district in Kanchanaburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

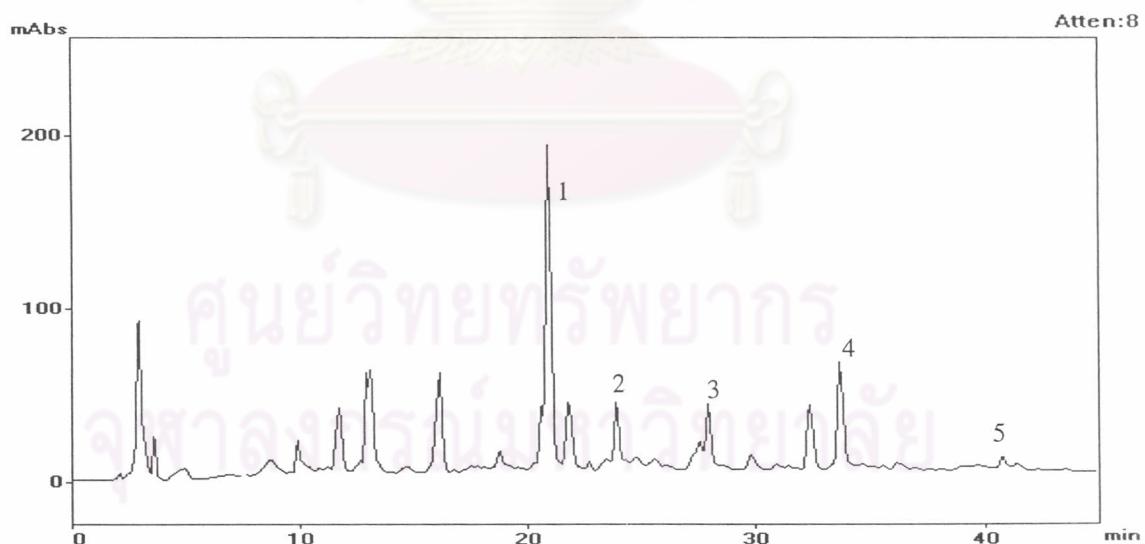


Figure 94 Isoflavone HPLC fingerprint of *P. mirifica* from Thongphaphum district in Kanchanaburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

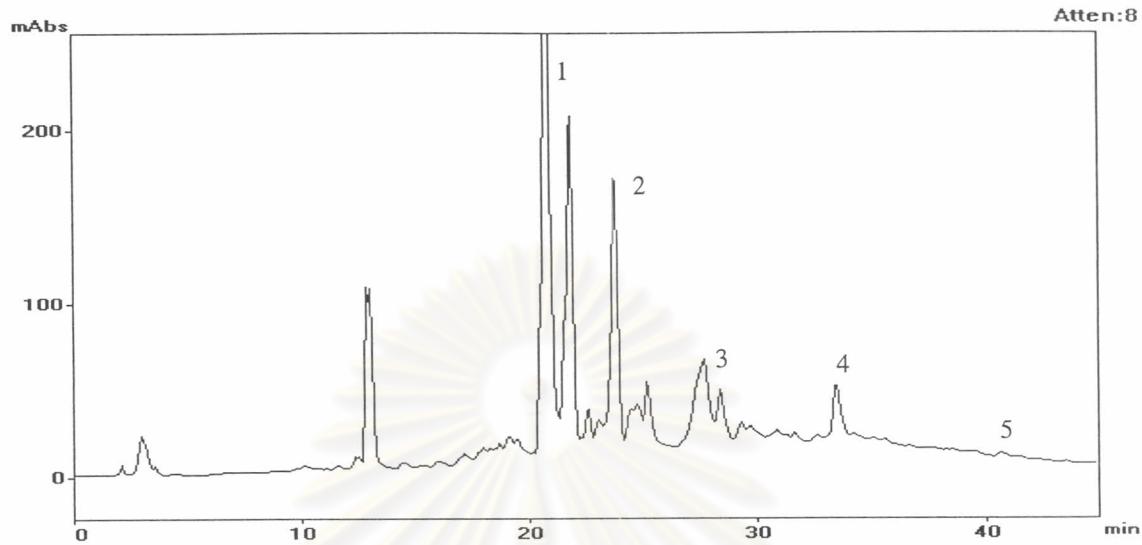


Figure 95 Isoflavone HPLC fingerprint of *P. mirifica* from Muak Lek district site I in Saraburi province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

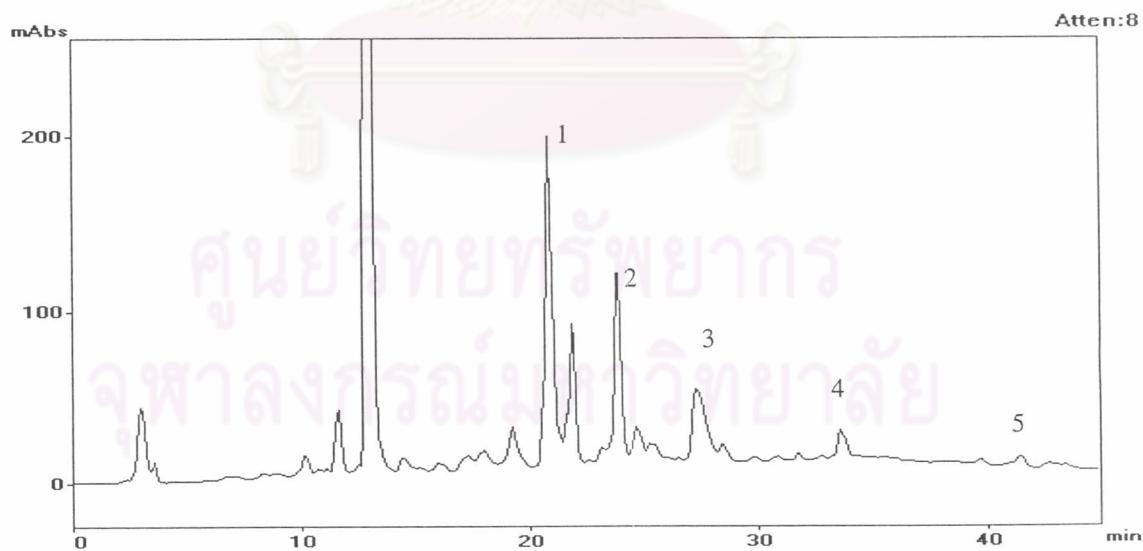


Figure 96 Isoflavone HPLC fingerprint of *P. mirifica* from Phra Putthabat district in Saraburi province

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

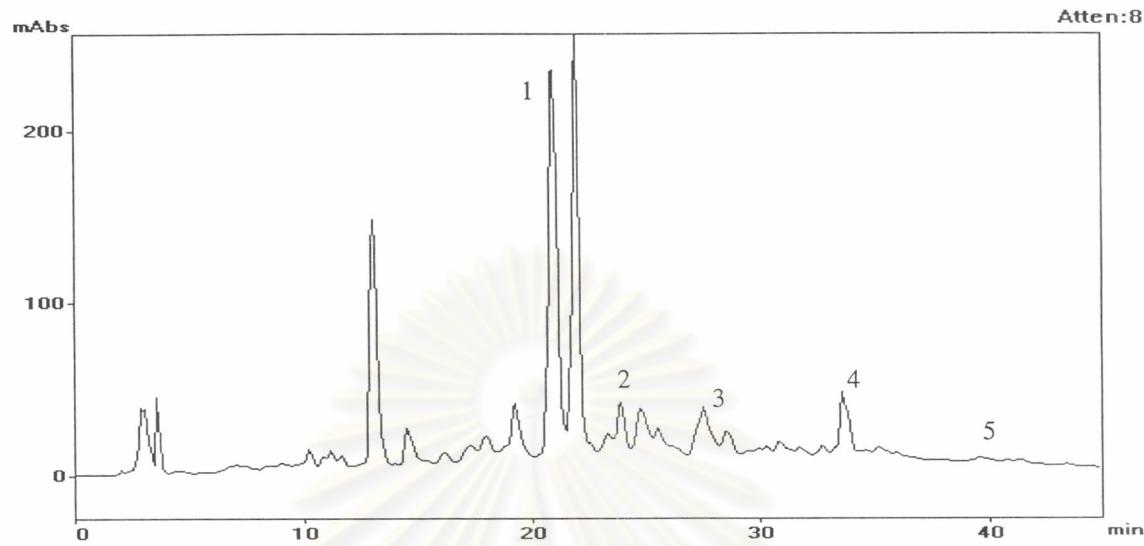


Figure 97 Isoflavone HPLC fingerprint of *P. mirifica* from Muak Lek district site II in Saraburi province

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

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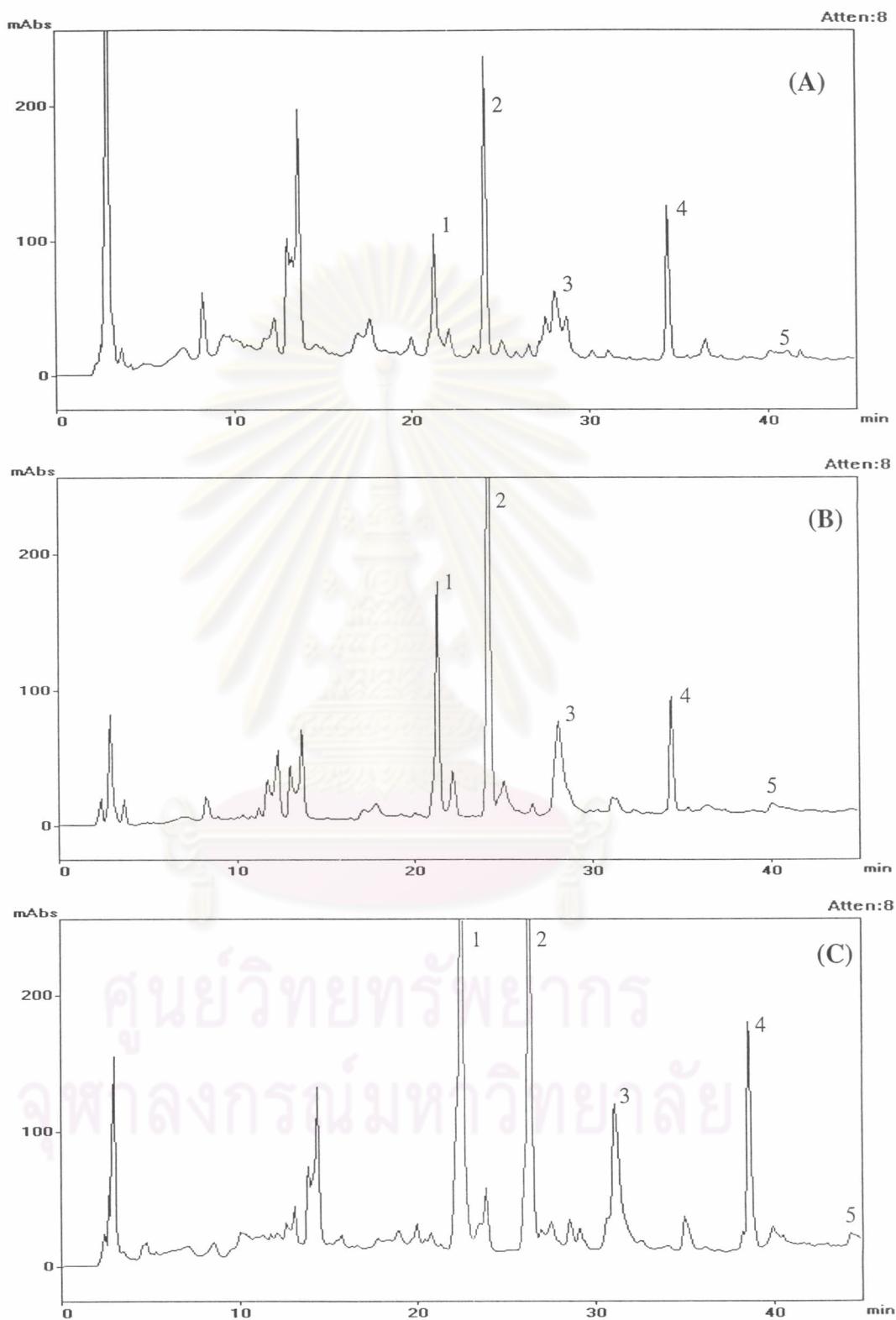


Figure 98 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao cultivated in different location
 (A) Chaing Rai (B) Bangkok (C) Ratchaburi
 1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

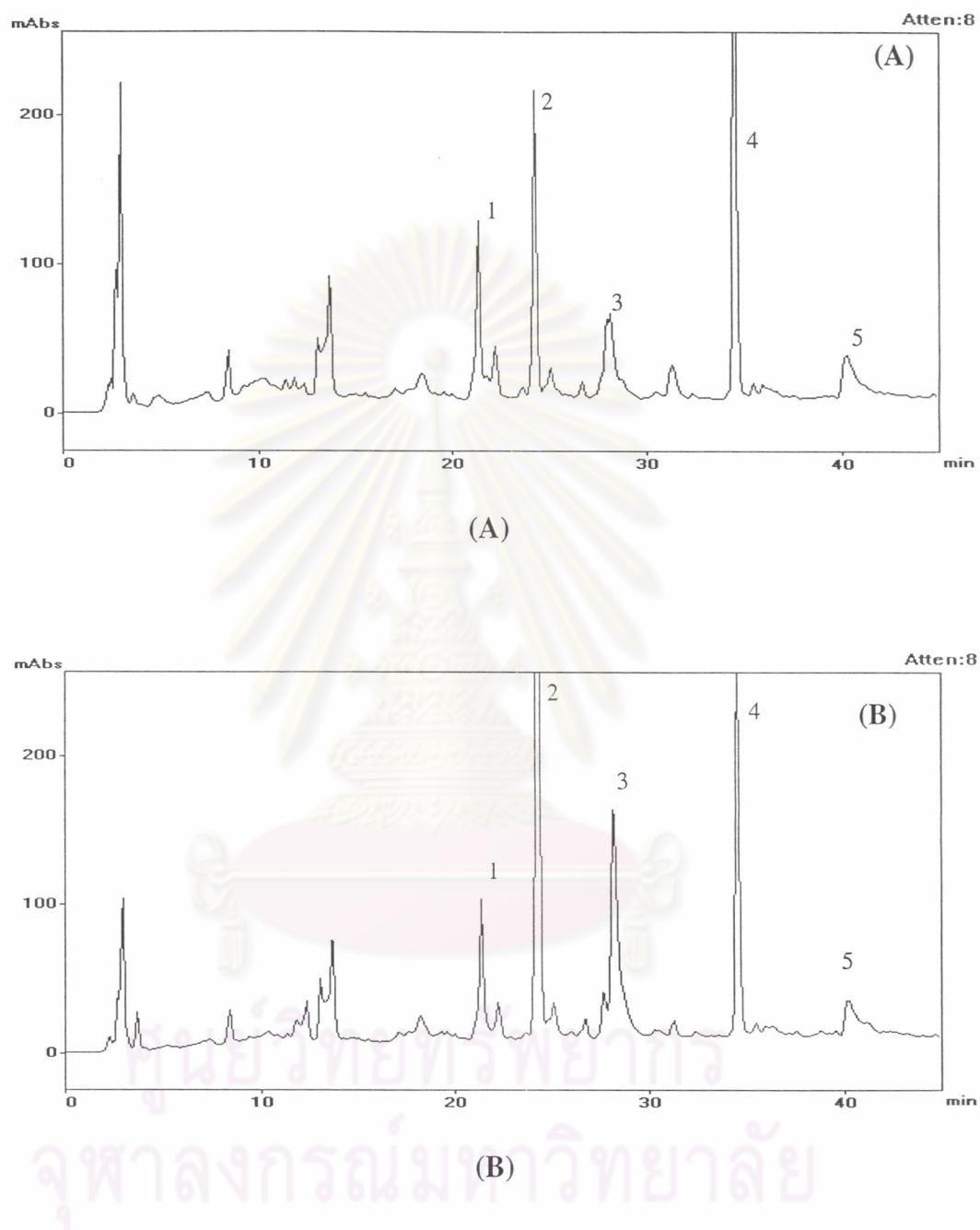


Figure 99 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Chaiprakarn cultivated in different location
 (A) Chaing Rai (B) Ratchaburi
 1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

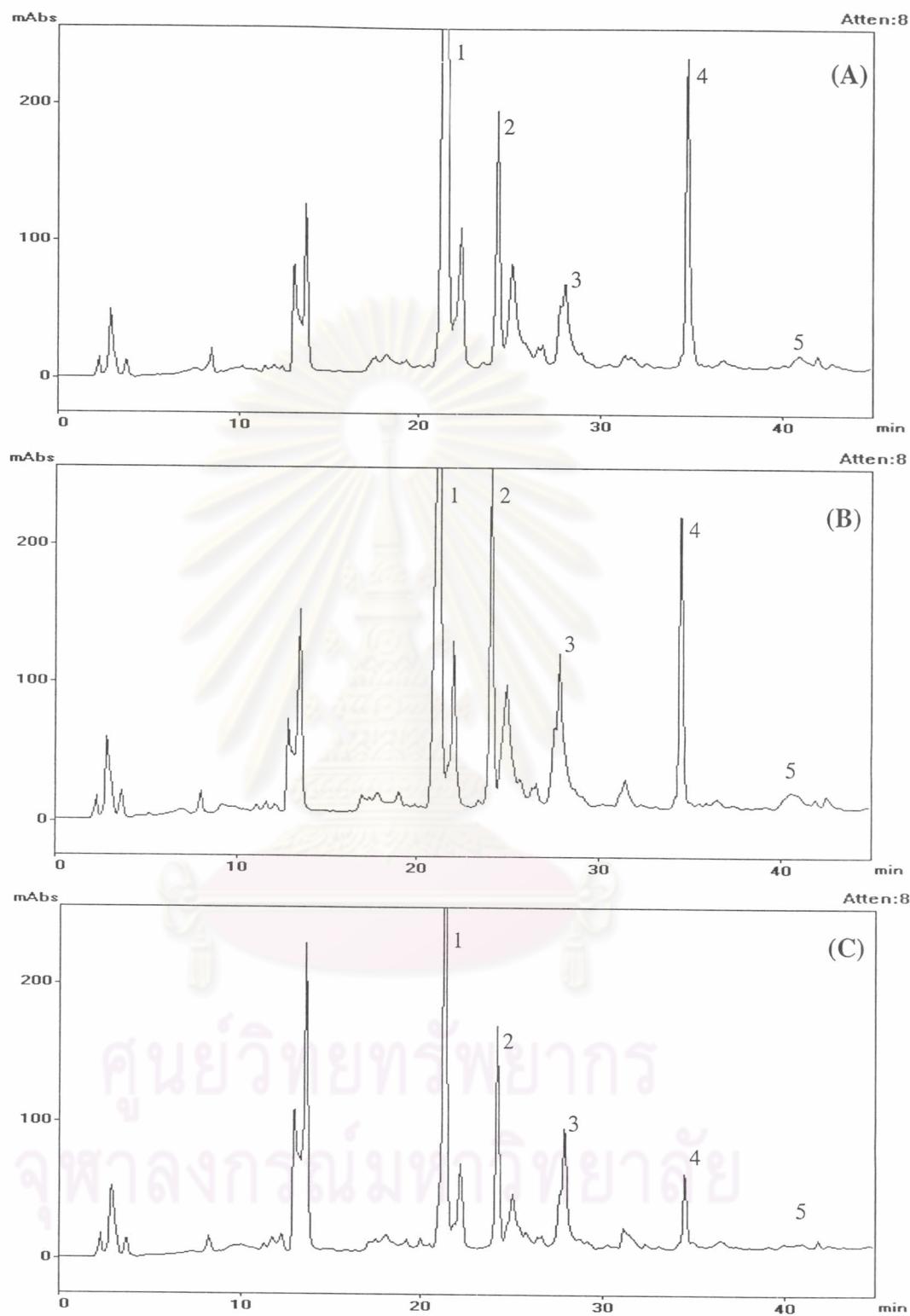


Figure 100 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao cultivated in different season

(A) Rainy (B) Winter (C) Summer

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

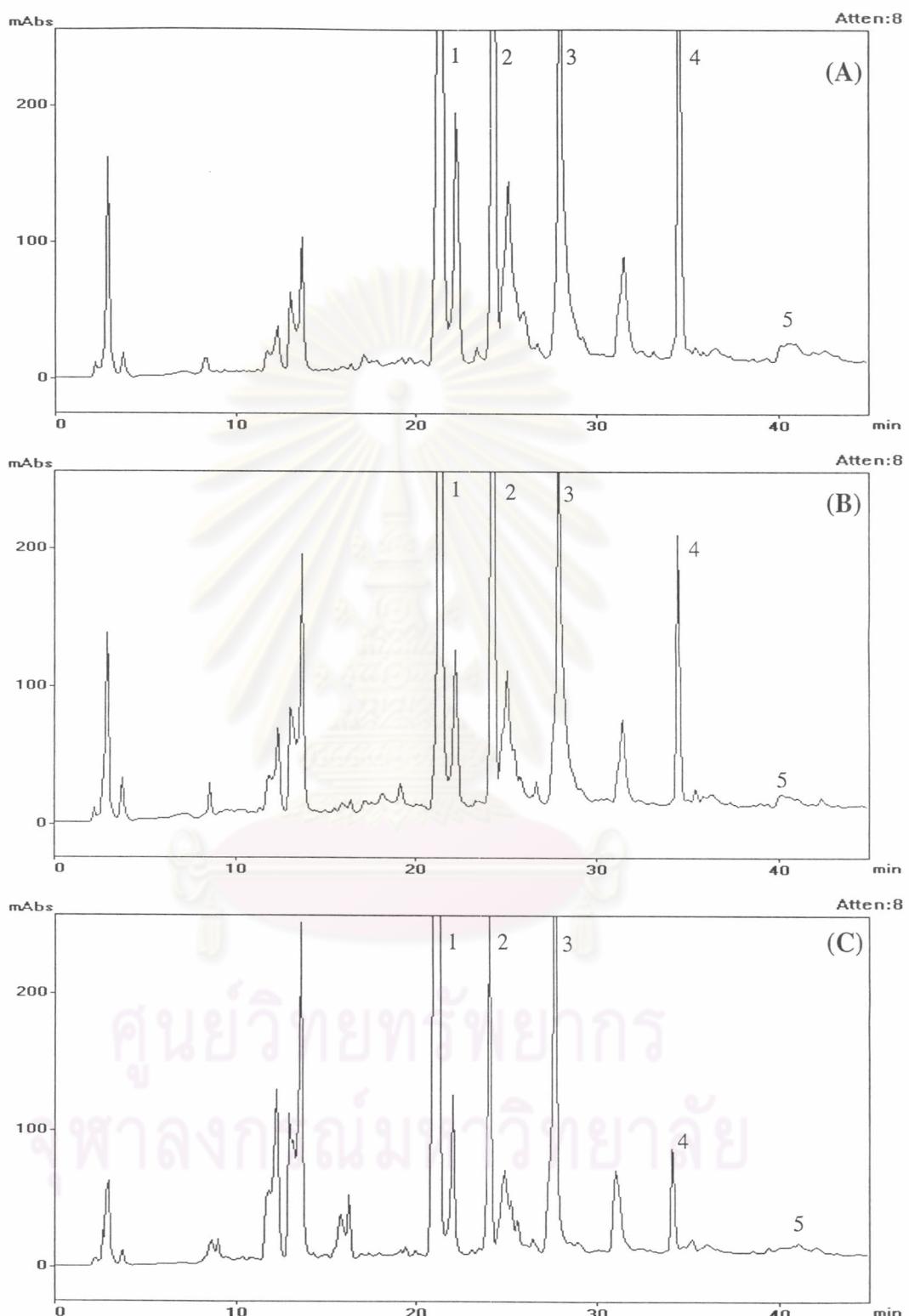


Figure 101 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Chaiprakarn cultivated in different season
 (A) Rainy (B) Winter (C) Summer
 1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

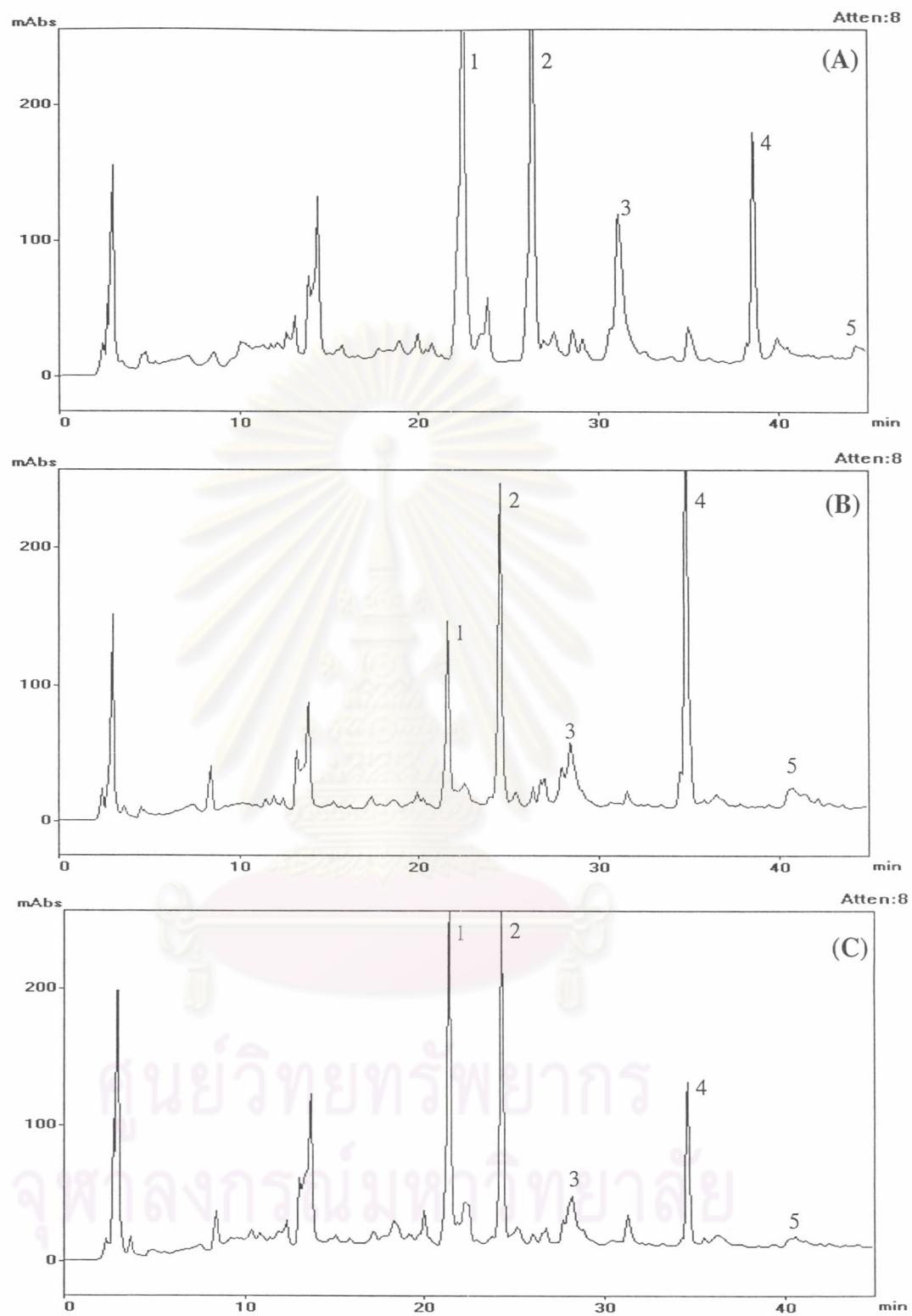


Figure 102 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao (F1) cultivated in Ratchaburi province
 (A) sub-clone I (B) sub-clone II (C) sub-clone III
 1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

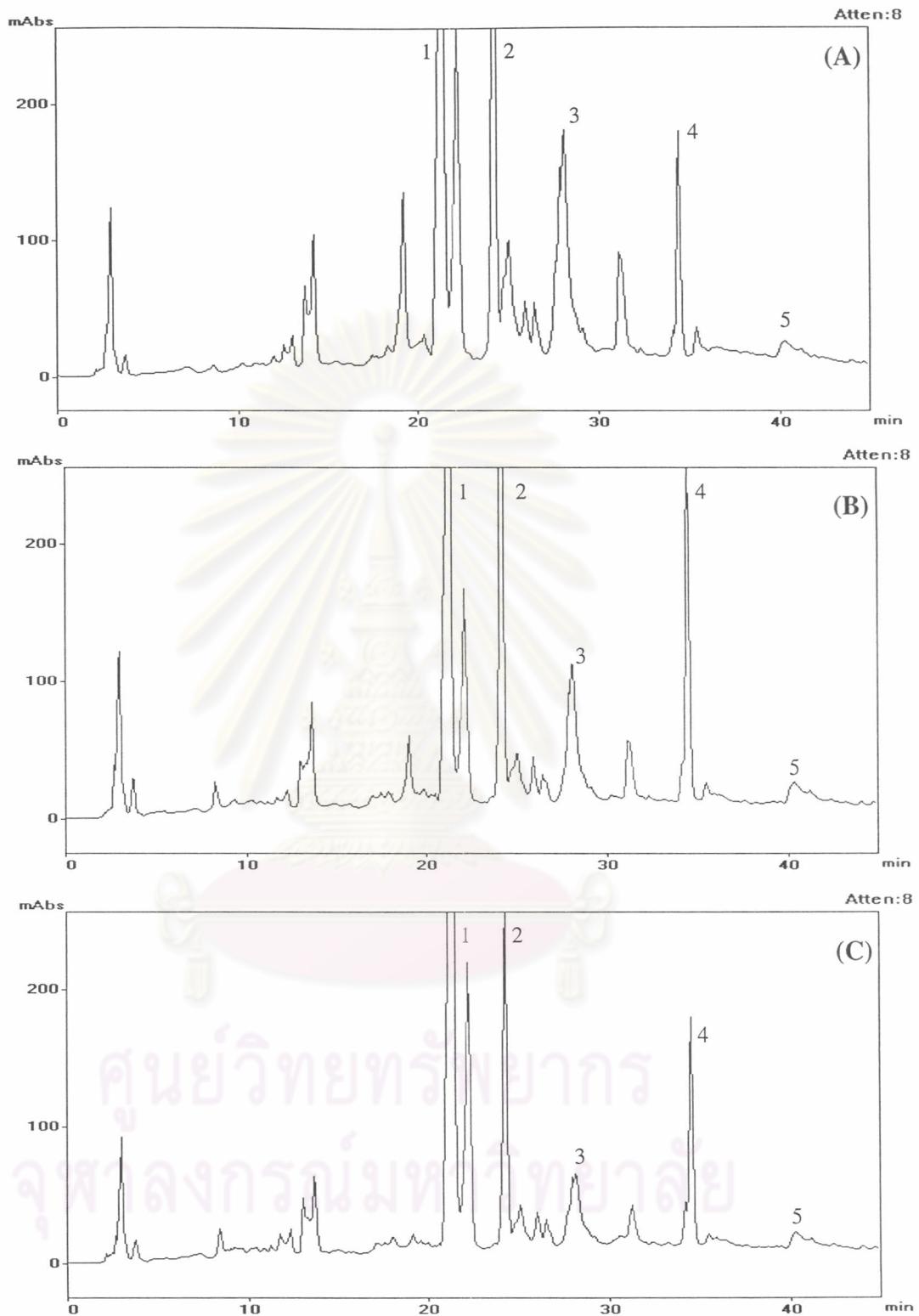


Figure 103 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Sai Yoke (F₁) cultivated in Ratchaburi province
 (A) sub-clone I (B) sub-clone II (C) sub-clone III
 1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

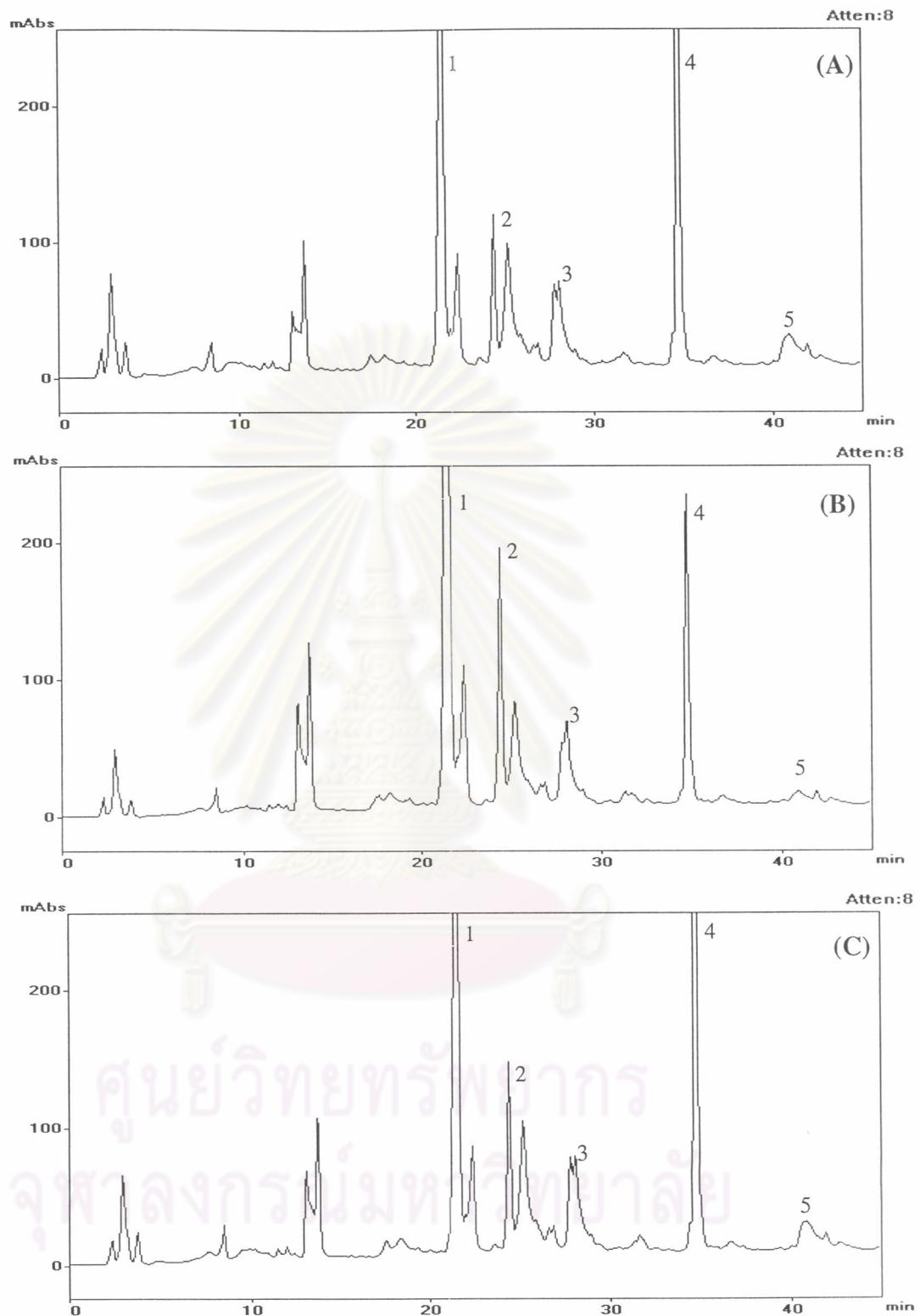


Figure 104 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao cultivated in the rainy season

(A) sub-clone I (B) sub-clone II (C) sub-clone III

1 = Puerarin 2= Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

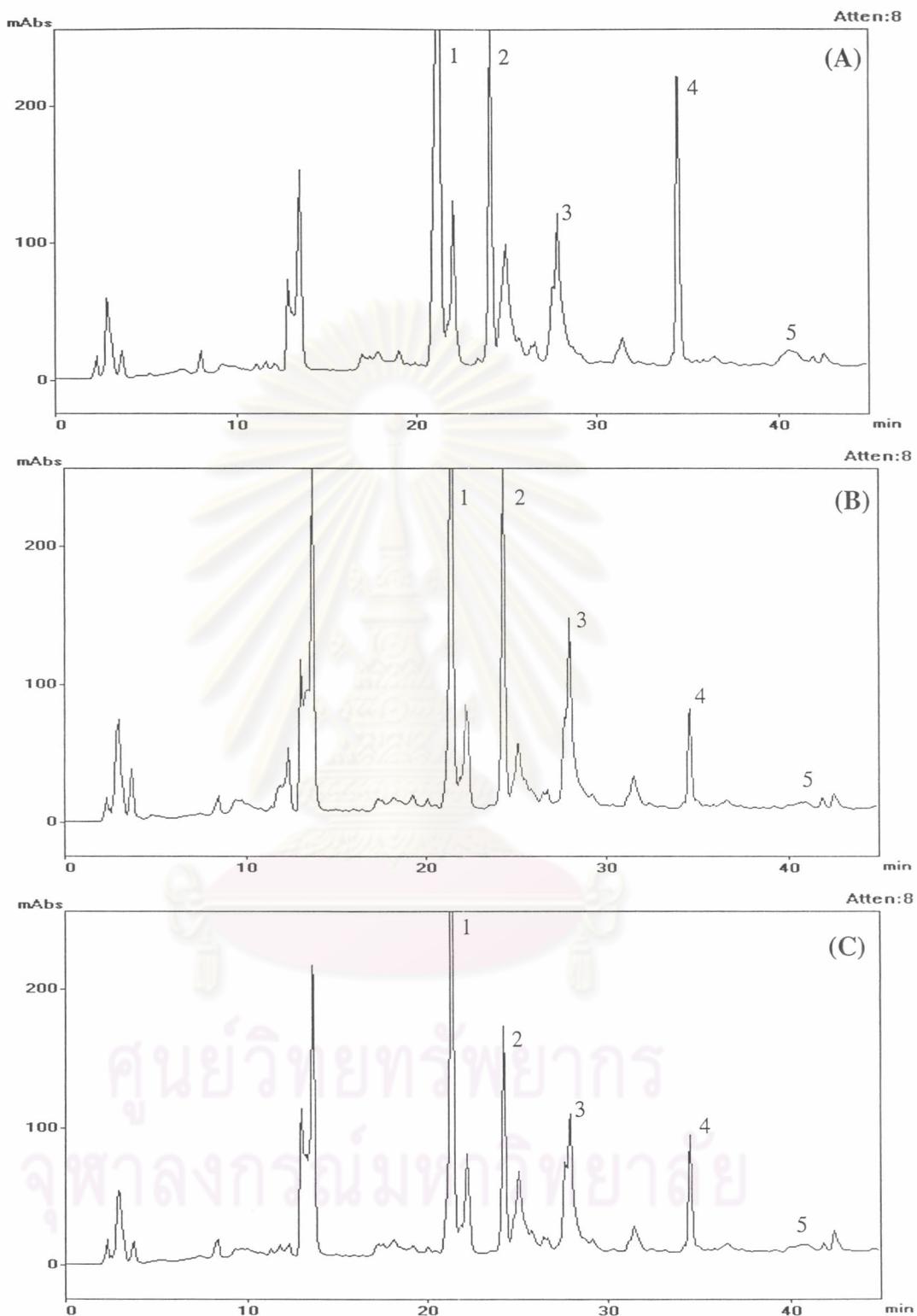


Figure 105 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao cultivated in winter

(A) sub-clone I (B) sub-clone II (C) sub-clone III

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

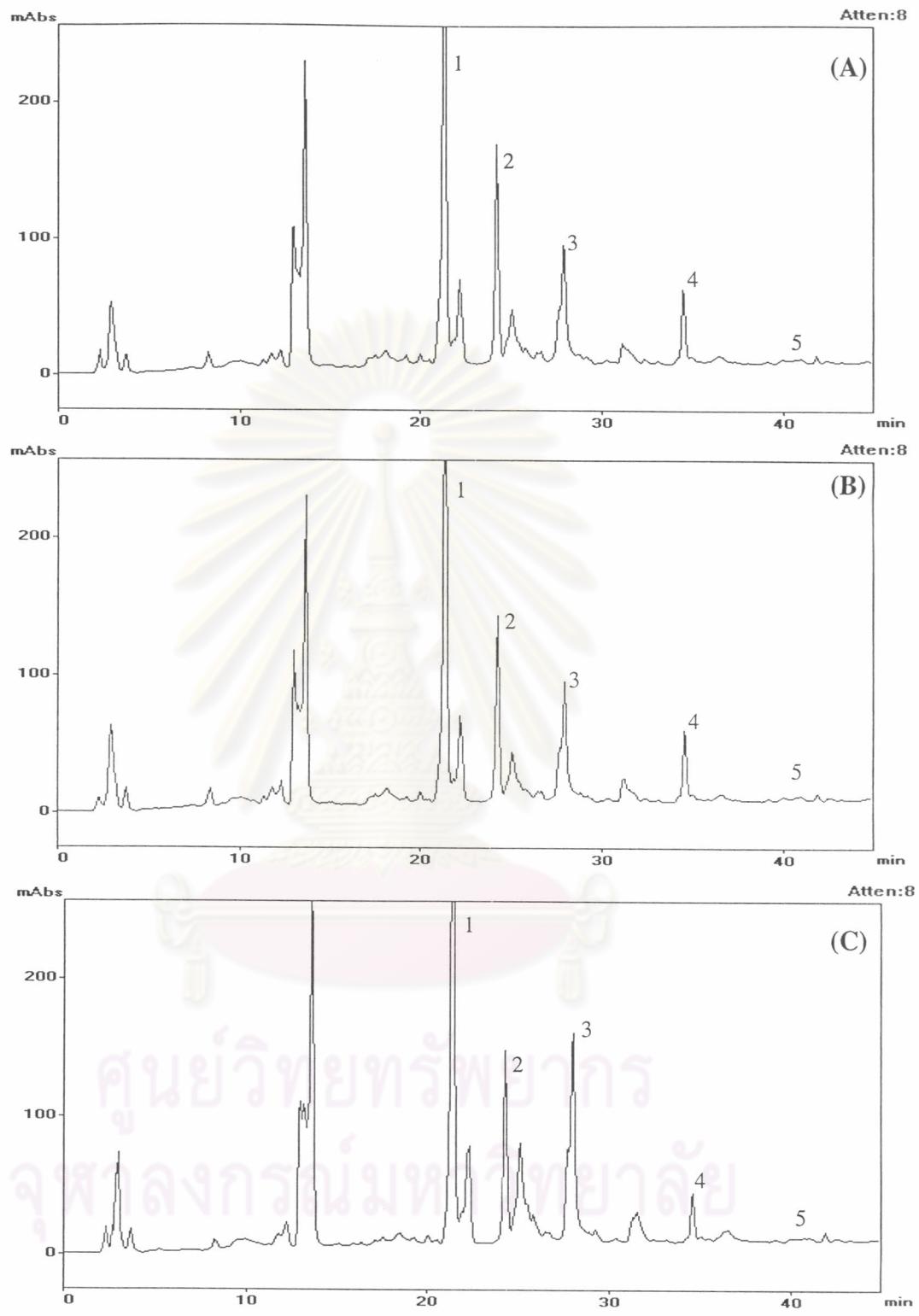


Figure 106 Isoflavone HPLC fingerprint of field grown *P. mirifica* clone Doi Tao cultivated in summer

(A) sub-clone I (B) sub-clone II (C) sub-clone III

1 = Puerarin 2 = Daidzin 3 = Genistin 4 = Daidzein 5 = Genistein

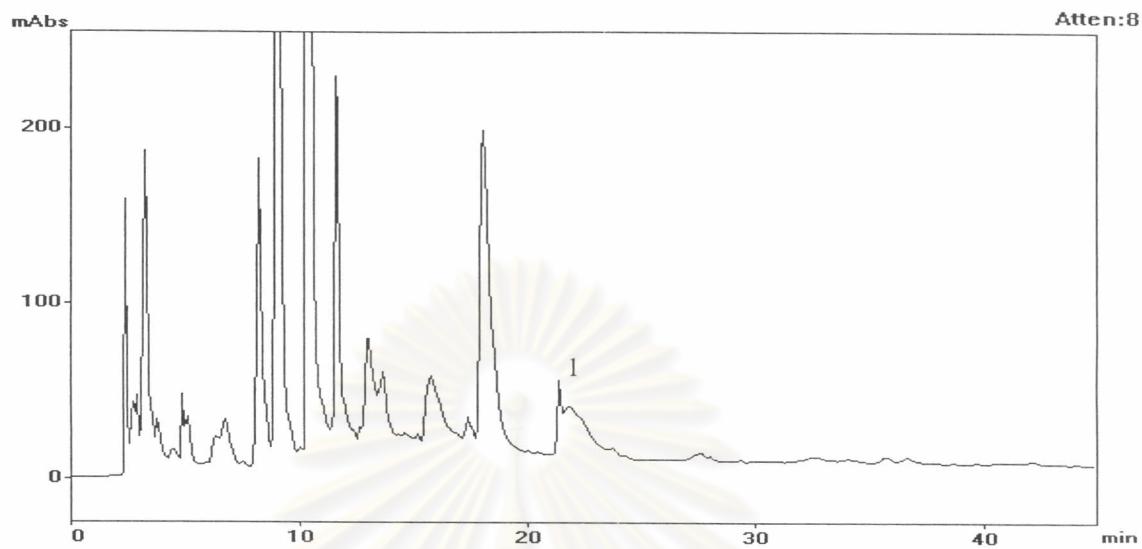


Figure 107 Isoflavone HPLC fingerprint of *P. mirifica* in chicken essence

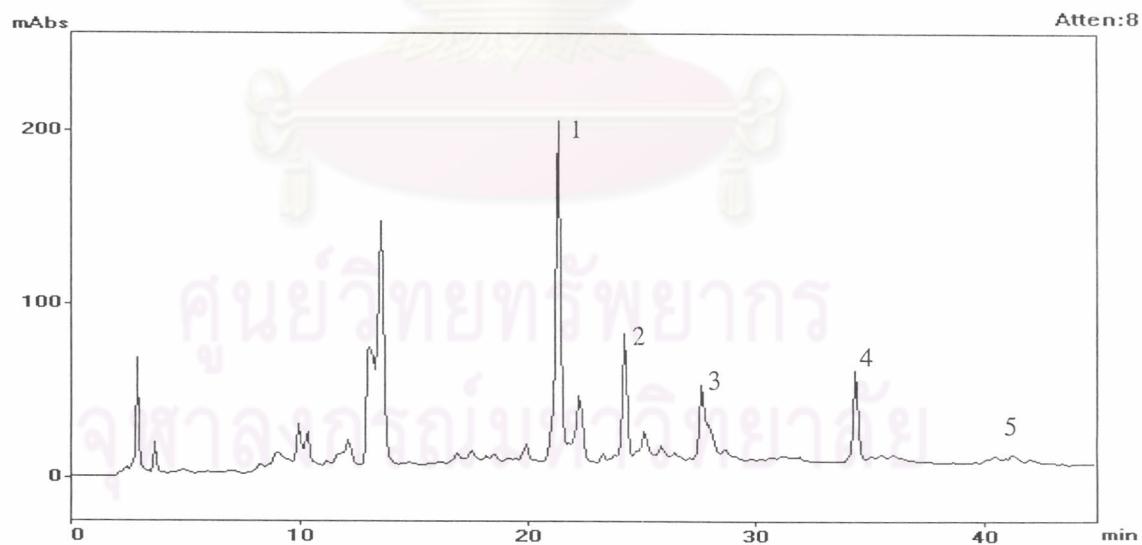


Figure 108 Isoflavone HPLC fingerprint of gamma-irradiated *P. mirifica*



APEENDIX III

HPLC FINGERPRINTS OF *B. superba*

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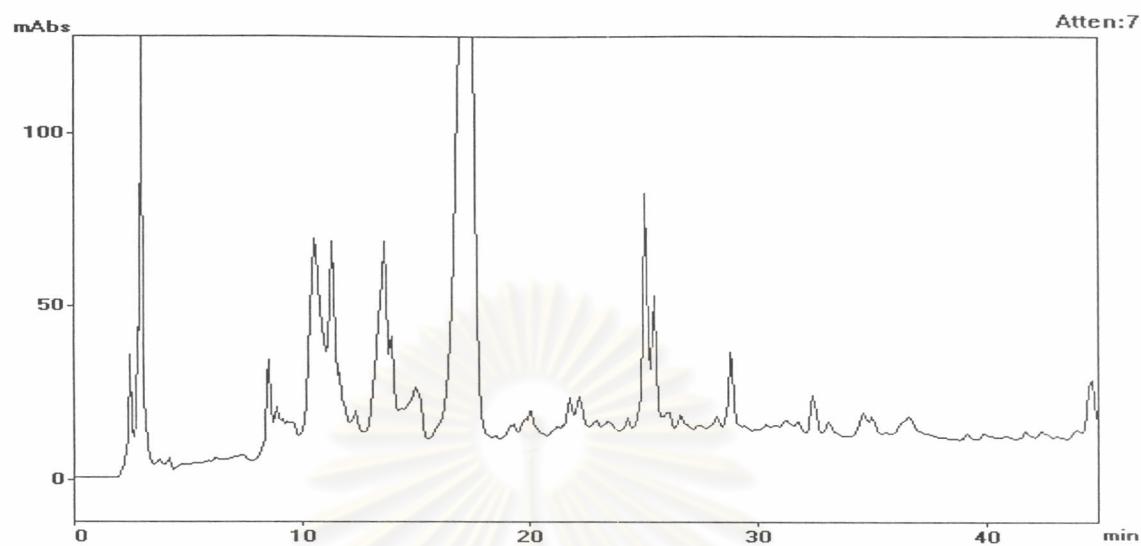


Figure 109 HPLC fingerprint of wild *B. superba* collected in Lampang province

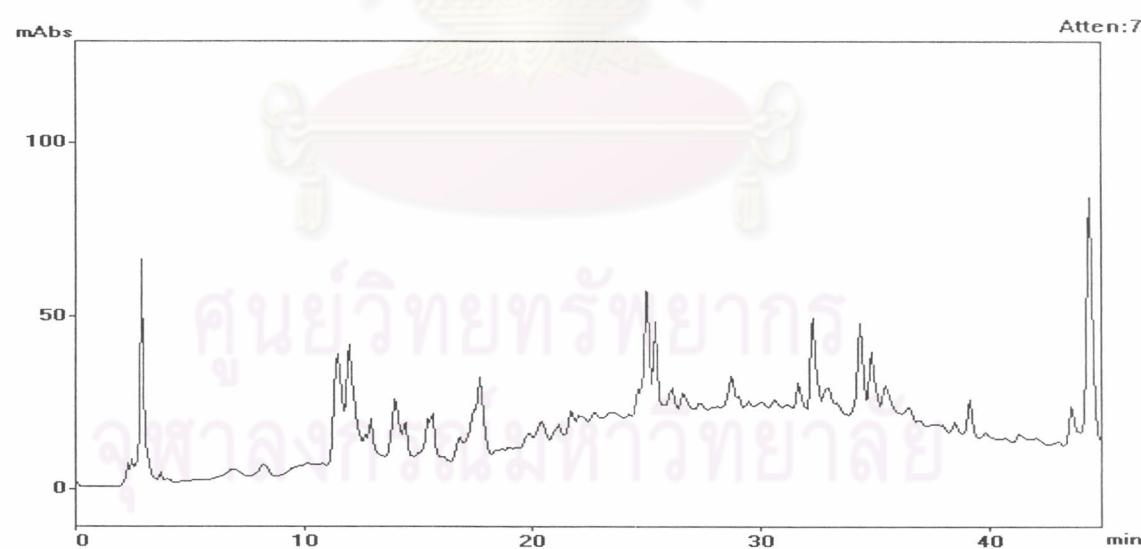


Figure 110 HPLC fingerprint of wild *B. superba* collected in Ratchaburi province

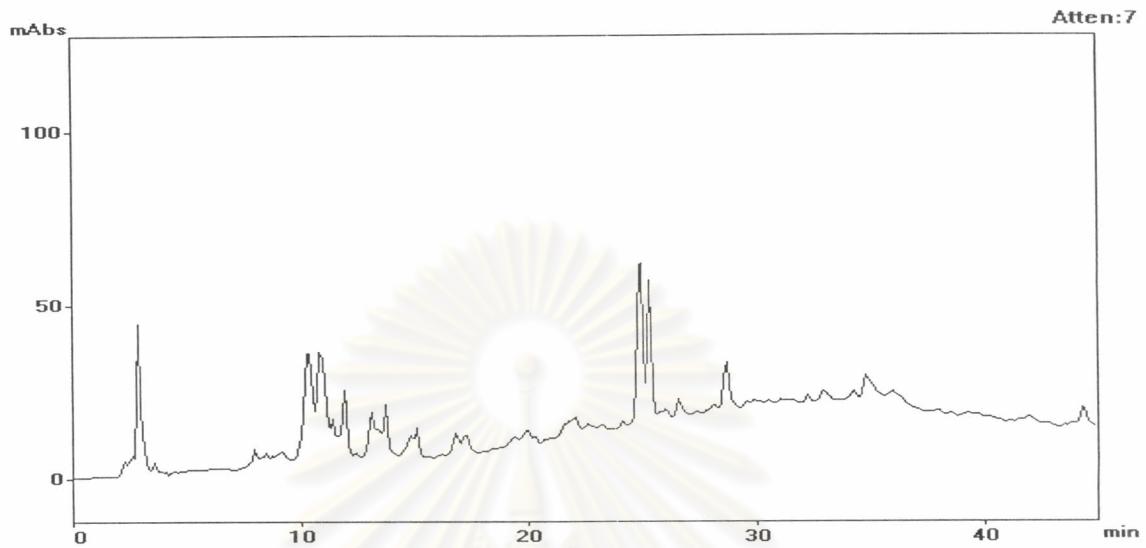


Figure 111 HPLC fingerprint of wild *B. superba* collected in Khon Kaen province

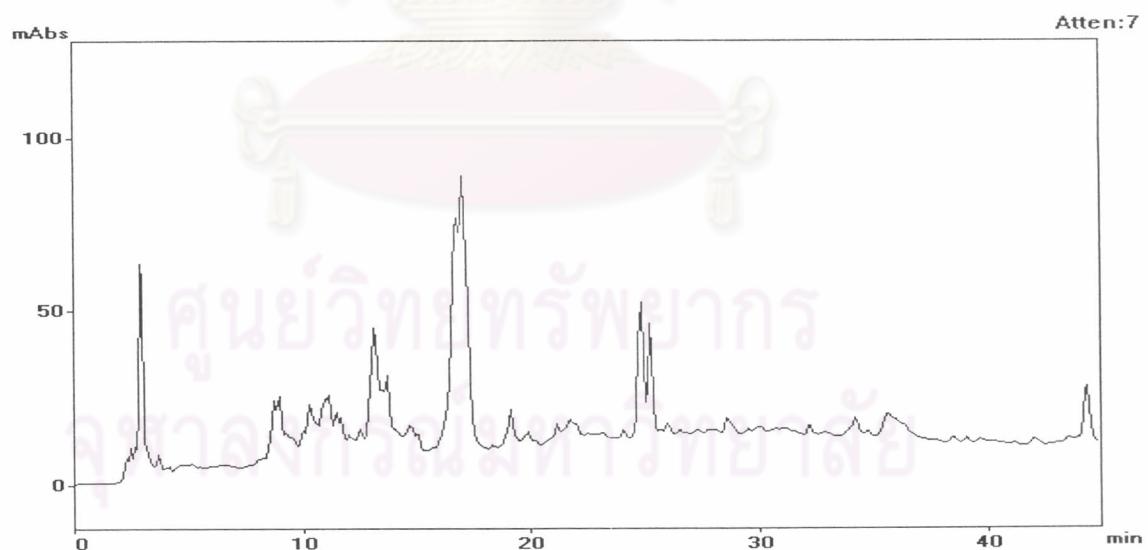


Figure 112 HPLC fingerprint of wild *B. superba* collected in Chantaburi province



APPENDIX IV

HPLC FINGERPRINTS OF *M. collettii*

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

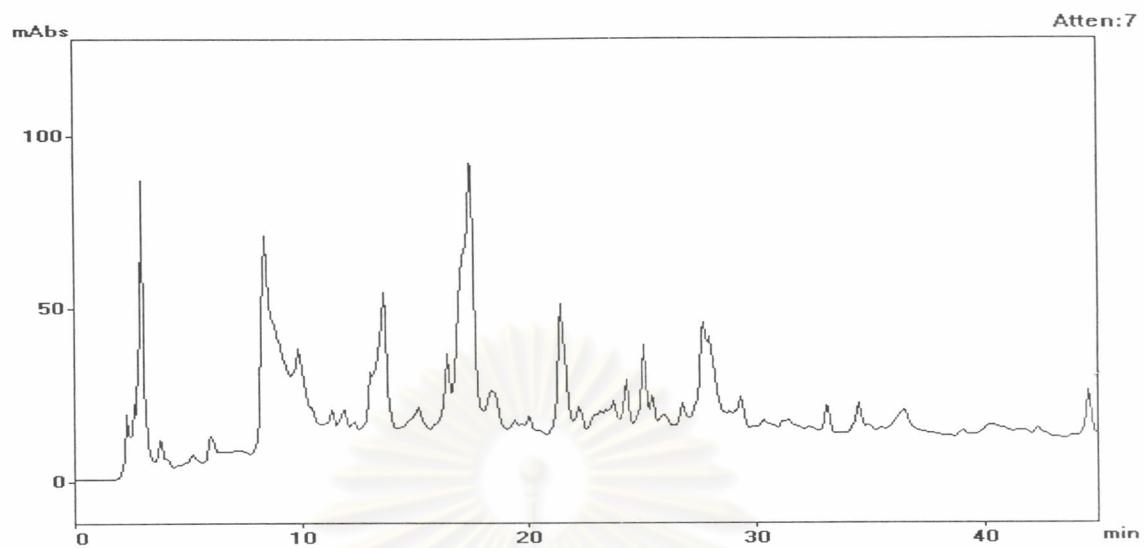


Figure 113 HPLC fingerprint of wild *M. collettii* collected in Chiang Rai province

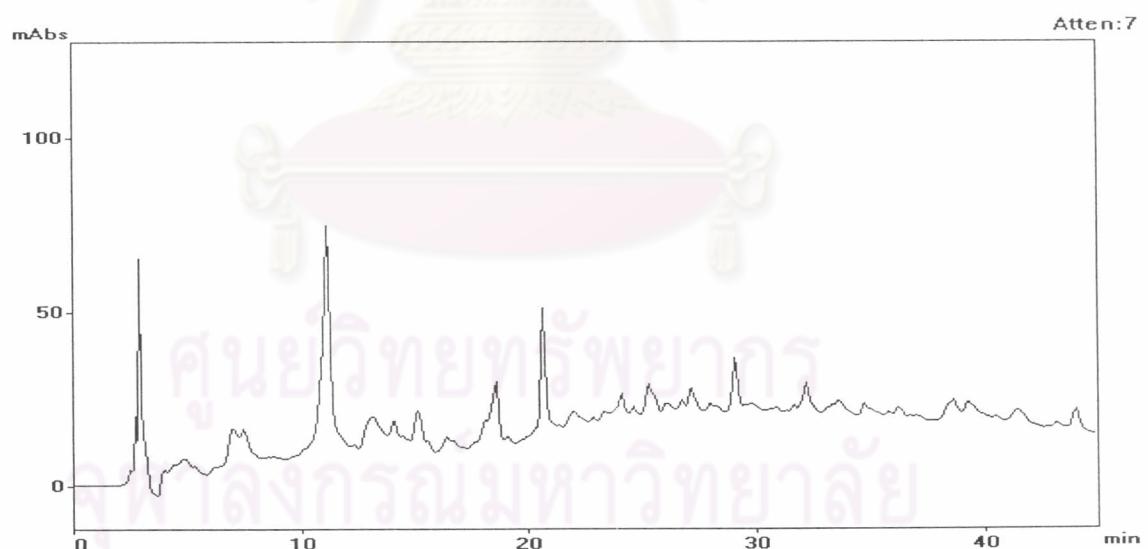


Figure 114 HPLC fingerprint of wild *M. collettii* collected in Lampang province

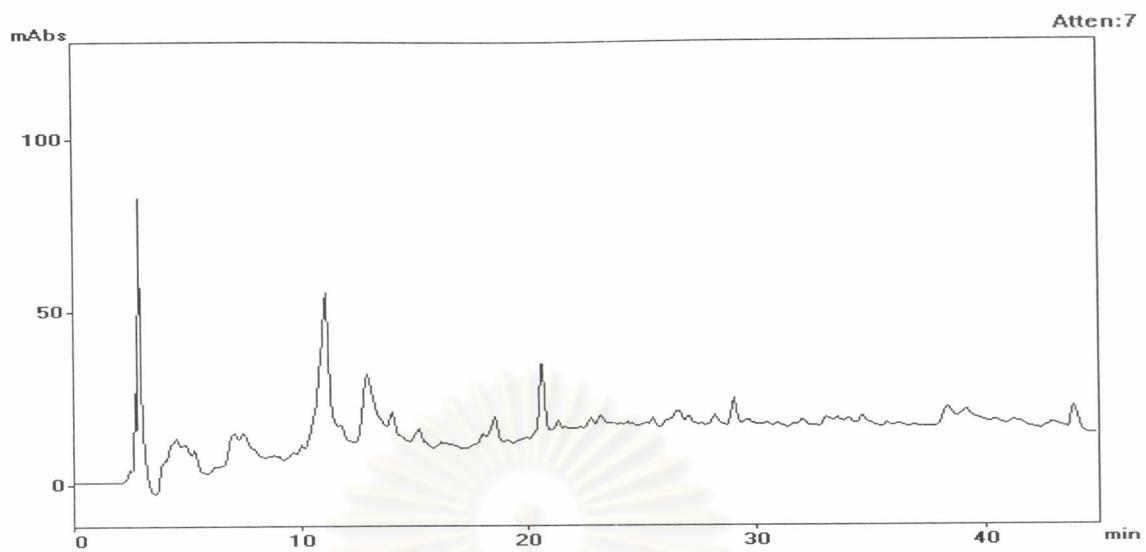


Figure 115 HPLC fingerprint of wild *M. collettii* collected in Kanchanaburi province

Biography

Ms. Subongkoch Subtang was born December 13, 1970 in Lamphang, Thailand. She was graduated with Bachelor degree of Science in Chemistry, Faculty of Science, Naraesuan University in 1992. She has enrolled in the Graduate School, Chulalongkorn University for Master Degree of Science in Biotechnology during 2000-2003.

