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## **APPENDICES**

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## APPENDIX A

### **The average grafting yield**

The grafting yield of graft polymerization was calculated by following equation;

$$\text{Grafting yield (\%)} = \frac{(\text{Weight}_{\text{after grafting}} - \text{Weight}_{\text{before grafting}})}{\text{Weight}_{\text{before grafting}}} \times 100 \quad \text{A-1}$$

**Table A-1.** The average grafting yield of vulcanized and unvulcanized rubber as a function of time using 0.5 M PEGMA.

<b>Grafting time (min)</b>	<b>Average grafting yield (%)</b>	
	<b>Unvulcanized rubber</b>	<b>Vulcanized rubber</b>
30	0.348±0.167	0.146±0.128
60	0.522±0.077	0.237±0.024
90	0.730±0.166	0.528±0.044
120	1.369±0.380	0.791±0.032
150	1.496±0.335	0.846±0.103
180	2.592±0.518	1.096±0.197

**Table A-2.** The average grafting yield of vulcanized and unvulcanized rubber as a function of PEGMA concentration for 150 min.

<b>Monomer concentration (M)</b>	<b>Average grafting yield (%)</b>	
	<b>Unvulcanized rubber</b>	<b>Vulcanized rubber</b>
0.1	0.505±0.080	0.307±0.043
0.2	0.733±0.150	0.389±0.033
0.3	1.112±0.117	0.571±0.040
0.4	1.030±0.519	0.629±0.052
0.5	1.496±0.335	0.846±0.103

**Table A-3.** The average grafting yield of vulcanized and unvulcanized rubber as a function of time with 0.5 M VPy.

<b>Grafting time (min)</b>	<b>Average grafting yield (%)</b>	
	<b>Unvulcanized rubber</b>	<b>Vulcanized rubber</b>
30	0.113±0.167	0.000±0.000
60	0.228±0.080	0.101±0.076
90	0.284±0.056	0.153±0.042
120	0.318±0.070	0.155±0.024
150	0.353±0.045	0.189±0.085
180	0.411±0.008	0.242±0.027

**Table A-4.** The average grafting yield of vulcanized and unvulcanized rubber as a function of VPy concentration for 150 min.

<b>Monomer concentration (M)</b>	<b>Average grafting yield (%)</b>	
	<b>Unvulcanized rubber</b>	<b>Vulcanized rubber</b>
0.1	0.209±0.053	0.081±0.017
0.2	0.251±0.074	0.105±0.016
0.3	0.269±0.015	0.136±0.074
0.4	0.292±0.042	0.165±0.073
0.5	0.353±0.045	0.189±0.085

**Table A-5.** Water contact angle of vulcanized NR and unvulcanized NR after being graft copolymerized by 0.5 M PEGMA and VPy as a function of time.

<b>Grafting time (min)</b>	<b>Water contact angle (degree)</b>			
	<b>U-g-PEGMA</b>	<b>V-g-PEGMA</b>	<b>U-g-VPy</b>	<b>V-g-VPy</b>
0	86.18 ± 1.82	88.45 ± 3.72	86.18 ± 1.82	88.45 ± 3.72
30	79.38 ± 3.29	81.43 ± 1.71	81.08 ± 1.38	87.85 ± 1.13
60	74.23 ± 2.05	79.73 ± 2.80	79.75 ± 0.81	84.70 ± 1.54
90	73.80 ± 1.35	74.10 ± 1.90	78.1 ± 1.48	81.48 ± 3.67
120	72.50 ± 4.54	72.53 ± 1.58	77.70 ± 1.59	79.10 ± 1.42
150	71.28 ± 1.42	70.30 ± 4.60	75.15 ± 1.75	77.85 ± 1.47
180	59.23 ± 3.90	66.95 ± 1.77	66.95 ± 2.25	74.48 ± 3.32

**Table A-6.** Water contact angle of vulcanized NR and unvulcanized NR after being graft copolymerized by PEGMA and VPy for 150 min as a function of monomer concentration.

<b>Monomer Concentration (M)</b>	<b>Water contact angle (degree)</b>			
	<b>U-g-PEGMA</b>	<b>V-g-PEGMA</b>	<b>U-g-VPy</b>	<b>V-g-VPy</b>
0	86.18 ± 1.82	88.45 ± 3.72	86.18 ± 1.82	88.45 ± 3.72
0.1	76.50 ± 2.64	76.68 ± 1.13	80.00 ± 2.03	82.28 ± 2.87
0.2	76.33 ± 1.61	74.90 ± 2.22	79.03 ± 1.74	81.83 ± 2.95
0.3	74.63 ± 2.79	73.4 ± 0.94	78.25 ± 1.05	79.43 ± 1.77
0.4	73.78 ± 3.89	72.35 ± 2.09	76.90 ± 3.04	78.40 ± 0.88
0.5	71.28 ± 1.42	70.3 ± 4.60	75.15 ± 1.75	77.85 ± 1.47

#### Determination of Benzophenone residue

**Table A-7** Standard benzophenone solution, for the calibration curve, was prepared according to the following table.

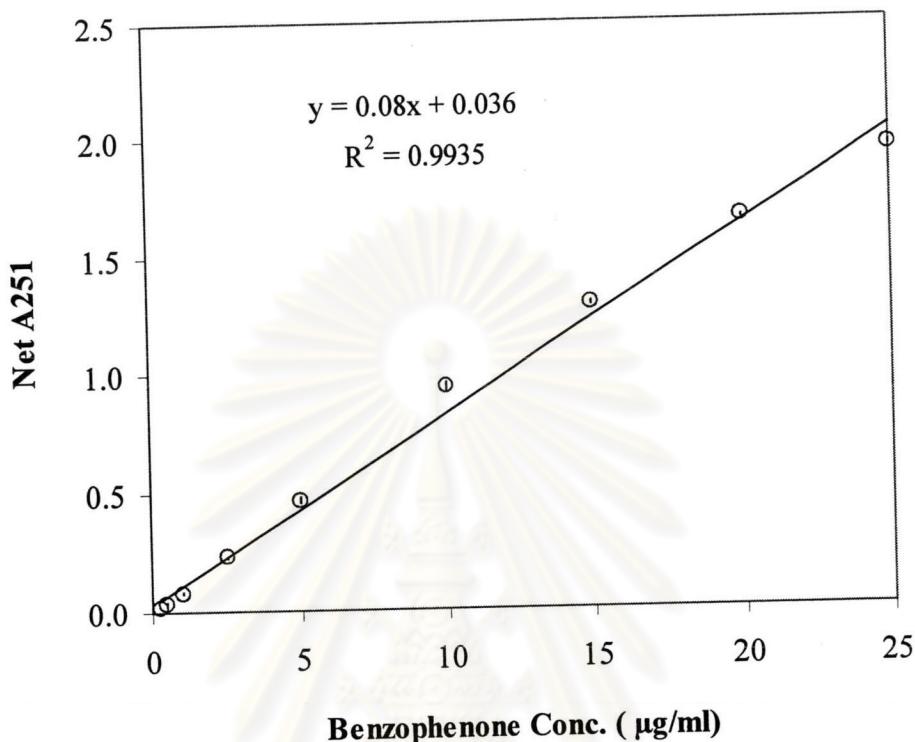
Standard	Solution (mL)	Ethanol (mL)	Benzophenone conc ( $\mu\text{g/mL}$ )
$S_1$	10.0 of Benzophenone solution (100 ( $\mu\text{g/mL}$ ) <sup>a</sup> )	30	25
$S_2$	12 of $S_1$	3.0	20
$S_3$	9.0 of $S_2$	3.0	15
$S_4$	6.0 of $S_3$	3.0	10
$S_5$	4.0 of $S_4$	4.0	5
$S_6$	4.0 of $S_5$	4.0	2.5
$S_7$	4.0 of $S_6$	6.0	1.0
$S_8$	4.0 of $S_7$	4.0	0.5
$S_9$	4.0 of $S_7$	4.0	0.25

After reading the UV absorbance of the samples and standard benzophenone solution at  $\lambda = 251 \text{ nm}$ . the calibration curve was plotted as shown in Figure A-1. The benzophenone concentration was determined from the calibration curve (C;  $\mu\text{g/mL}$ ). The amount of benzophenone (B) in the original solution (5 mL) from the sampling sample (1 mL)

$$\text{Total amount of benzophenone (B)} = C (\mu\text{g/mL}) \times 5 (\text{mL}) \quad \text{A-2}$$

After that the determined of the amount of adsorbed protein/surface area was calculated as follow;

$$\text{Adsorbed protein/surface area } B_{\text{ads}} = B / \text{surface area (2 sides)} (\mu\text{g/cm}^2) \quad \text{A-3}$$



**Figure A-1** A calibration curve of the amount of benzophenone as a function of UV absorbance at 251 nm.

**Table A-8** The amount of benzophenone per surface area ( $\mu\text{g}/\text{cm}^2$ ) dissolving out from vulcanized and unvulcanized NR before and after graft copolymerization.

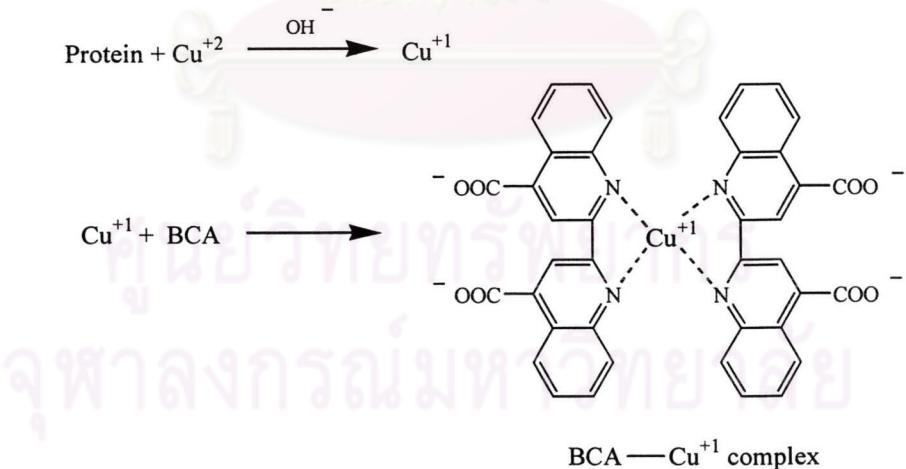
<b>Grafting condition</b>	<b>Amount of Benzophenone residue (μg/ml)</b>	
	<b>Unvulcanized rubber</b>	<b>Vulcanized rubber</b>
Before grafting	$32.14 \pm 1.77$	$42.91 \pm 1.78$
After grafting for 30 min	$5.09 \pm 0.45$	$3.58 \pm 0.84$
After grafting for 180 min	$4.80 \pm 1.20$	$4.30 \pm 0.73$

## APPENDIX B

### Bicinchoninic Acid Assay

Bicinchoninic acid assay is a method used for determination of the amount of proteins. The standard reagents used in this method are reagent A, reagent B and reagent C. Reagent A consists of an aqueous solution of Na<sub>2</sub>tartrate, Na<sub>2</sub>CO<sub>3</sub>, NaHCO<sub>3</sub> in 0.2 M NaOH, pH 11.25. Reagent B is 4% (W/V) bicinchoninic acid solution, pH 8.5. Reagent C is 4% CuSO<sub>4</sub>·5H<sub>2</sub>O in deionized water.

The principle of the bicinchoninic assay (BCA) relies on the formation of a Cu<sup>2+</sup>-protein complex under alkaline conditions, followed by reduction of the Cu<sup>2+</sup> to Cu<sup>1+</sup>. The amount of reduction is proportional to protein present. It has been shown that the peptide bond is able to reduce Cu<sup>2+</sup> to Cu<sup>1+</sup>. BCA forms a purple-blue complex with Cu<sup>1+</sup> in alkaline environments, thus providing a basis to monitor the reduction of alkaline Cu<sup>2+</sup> by proteins.<sup>30</sup> Figure B-1 shows complexation between bicinchoninic acid and Cu<sup>1+</sup>.



**Figure B-1.** Formation of purple complex between BCA and cuprous ion generated from the biuret reaction.

### Calculation of Protein Adsorption

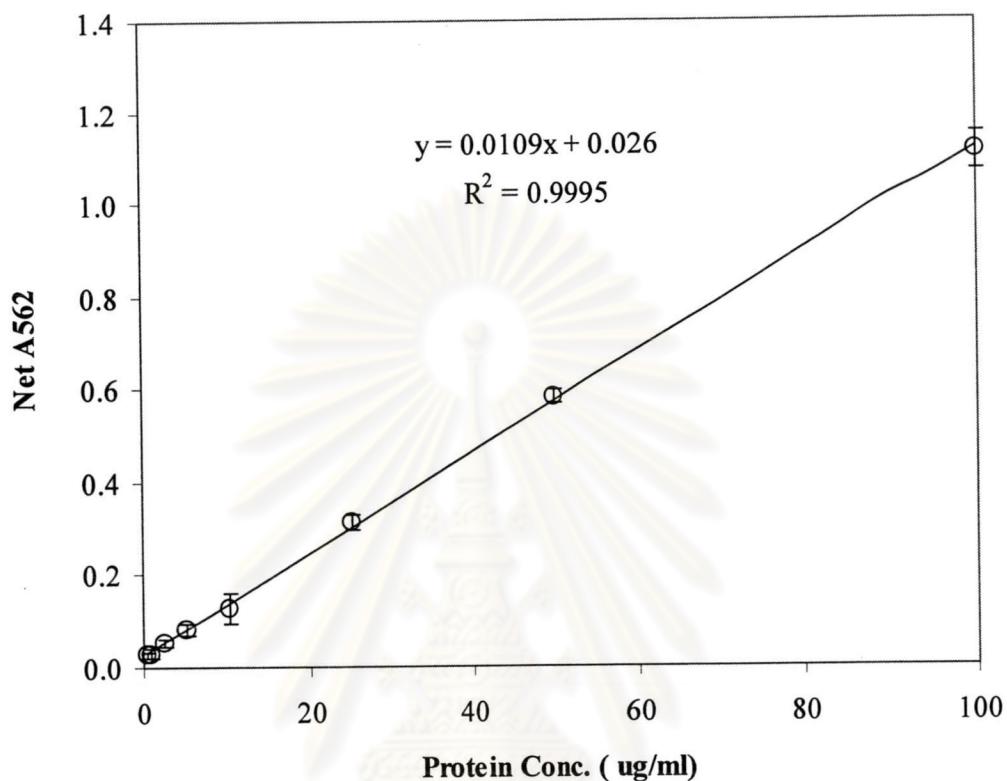
**Table B-1** Standard BSA solution, for the calibration curve.

Standard	Solution (mL)	SDS (mL)	BSA conc ( $\mu\text{g/mL}$ )
S <sub>1</sub>	0.5 of BSA (1000 ( $\mu\text{g/mL}$ ) <sup>a</sup> )	4.5	100
S <sub>2</sub>	4.0 of S <sub>1</sub>	4.0	5.0
S <sub>3</sub>	4.0 of S <sub>2</sub>	4.0	2.5
S <sub>4</sub>	4.0 of S <sub>3</sub>	6.0	1.0
S <sub>5</sub>	4.0 of S <sub>4</sub>	4.0	0.5
S <sub>6</sub>	4.0 of S <sub>5</sub>	4.0	0.25
S <sub>7</sub>	4.0 of S <sub>6</sub>	6.0	0.10
S <sub>8</sub>	4.0 of S <sub>7</sub>	4.0	0.05

a : standard BSA was pipette from 1 mg/mL ampule

After reading the UV absorbance of the samples and standard BSA solution at  $\lambda = 562 \text{ nm}$ , the result was then calculated for the net absorbance by subtracting the absorbance of the blank (SDS).

$$\text{Net } A_{562} = \text{recorded } A_{562} - A_{562} (\text{blank}) \quad \text{B-1}$$



**Figure B-2** A calibration curve of the amount of albumin adsorbed and the absorbance obtained from BCA microassay.

The protein concentration (C;  $\mu\text{g/mL}$ ) in each well was determined from the calibration curve. The total amount of protein (P) in the original solution (1 mL) was calculated from the sampling sample (100  $\mu\text{L}$ ) + BCA working solution (100  $\mu\text{L}$ )

$$\text{Total amount of protein (P)} = \frac{\text{C} (\mu\text{g/mL}) \times 200 (\mu\text{L})}{1000 (\mu\text{L/mL})} \times \frac{1000 (\mu\text{L})}{100 (\mu\text{L})} \quad \text{B-2}$$

$$\text{Adsorbed protein/surface area } P_{\text{ads}} = P / \text{surface area (2 sides)} (\mu\text{g/cm}^2) \quad \text{B-3}$$

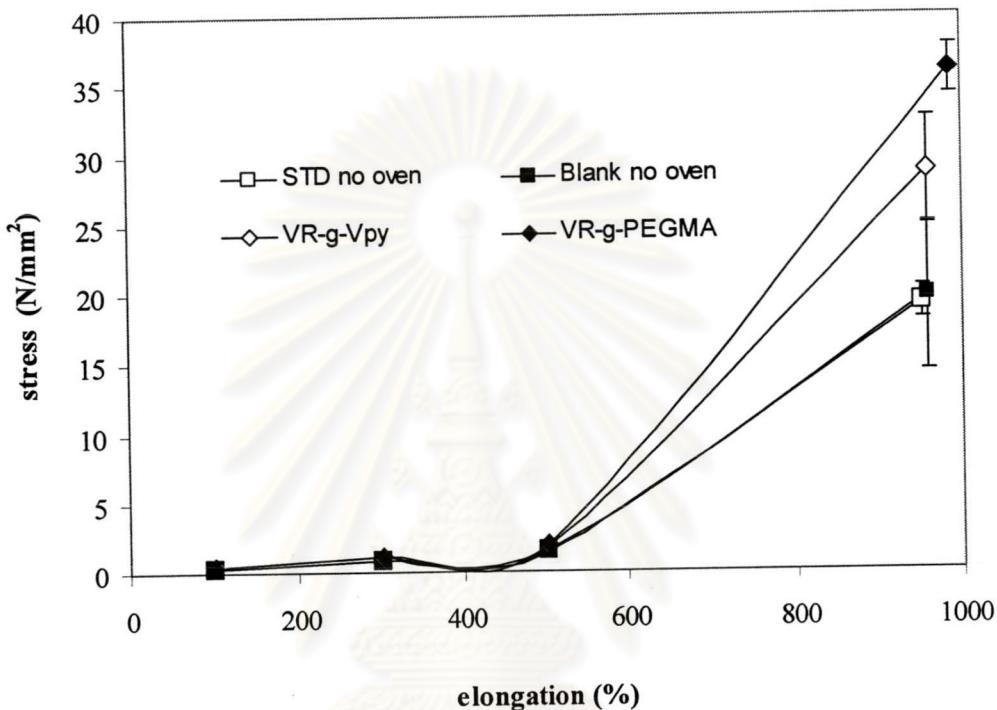
**Table B-2** The amount of plasma protein adsorption per surface area ( $\mu\text{g}/\text{cm}^2$ ) of Vulcanized and unvulcanized rubber before and after graft copolymerization with PEGMA and VPy as function of time.

<b>Grafting time (min)</b>	<b>The amount of plasma protein adsorption (<math>\mu\text{g}/\text{cm}^2</math>)</b>			
	<b>U-g-PEGMA</b>	<b>V-g-PEGMA</b>	<b>U-g-VPy</b>	<b>V-g-VPy</b>
0	26.396 $\pm$ 3.980	12.834 $\pm$ 1.570	26.396 $\pm$ 3.980	12.834 $\pm$ 1.570
30	8.380 $\pm$ 4.100	11.184 $\pm$ 4.100	19.419 $\pm$ 0.711	12.437 $\pm$ 3.115
60	3.286 $\pm$ 0.999	10.648 $\pm$ 0.999	13.673 $\pm$ 3.799	11.069 $\pm$ 1.375
90	0.596 $\pm$ 0.531	6.424 $\pm$ 0.531	7.807 $\pm$ 3.174	8.016 $\pm$ 3.497
120	-0.304 $\pm$ 1.153	2.275 $\pm$ 1.153	5.536 $\pm$ 0.887	7.671 $\pm$ 2.923
150	-1.027 $\pm$ 0.844	0.551 $\pm$ 0.844	3.046 $\pm$ 0.431	6.449 $\pm$ 1.053
180	-0.767 $\pm$ 2.839	-0.174 $\pm$ 2.839	2.612 $\pm$ 0.862	3.743 $\pm$ 0.526

**Table B-3** The amount of plasma protein adsorption per surface area ( $\mu\text{g}/\text{cm}^2$ ) of Vulcanized and unvulcanized rubber before and after graft copolymerization with PEGMA and VPy as function of monomer concentration.

<b>Monomer Concentration (M)</b>	<b>The amount of plasma protein adsorption (<math>\mu\text{g}/\text{cm}^2</math>)</b>			
	<b>U-g-PEGMA</b>	<b>V-g-PEGMA</b>	<b>U-g-VPy</b>	<b>V-g-VPy</b>
0	26.396 $\pm$ 3.980	12.834 $\pm$ 1.570	26.396 $\pm$ 3.980	12.834 $\pm$ 1.570
0.1	4.939 $\pm$ 0.612	10.596 $\pm$ 2.358	8.102 $\pm$ 4.347	11.220 $\pm$ 1.620
0.2	4.536 $\pm$ 1.669	7.064 $\pm$ 3.177	7.013 $\pm$ 3.085	10.447 $\pm$ 1.966
0.3	0.099 $\pm$ 0.642	3.261 $\pm$ 0.714	5.723 $\pm$ 1.156	9.134 $\pm$ 1.083
0.4	-0.347 $\pm$ 0.311	1.122 $\pm$ 1.059	3.269 $\pm$ 1.077	8.443 $\pm$ 0.547
0.5	-1.027 $\pm$ 1.794	0.551 $\pm$ 0.774	3.046 $\pm$ 0.431	6.449 $\pm$ 1.053

## APPENDIX C



**Figure C-1** Stress-strain curves of NR latex films before and after graft copolymerization using 0.5 M PEGMA and VPy for 150 min

## VITAE

Kamolmart Chombanpaew was born in Bangkok, Thailand, on July 10<sup>th</sup>, 1975. She received Bachelor Degree of Science (Industrial Chemistry) in 1997 from Department of Chemistry, Faculty of Science, Srinakarinwirote University. She started as a Master Degree student with a major in Polymer Science, Program of Petrochemistry and Polymer Science, Chulalongkorn University in 2001 and completed the program in May 2004.

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