

ฤทธิ์ก่อกลายพันธุ์และฤทธิ์ต้านก่อกลายพันธุ์ของสารสกัดกวาวเครือขาว
Pueraria mirifica กวาวเครือแดง *Butea superba* และกวาวเครือดำ
Mucuna collettii ด้วยวิธีทดสอบแบบเอมส์



นายเกษตร พูลเจริญ

ศูนย์วิทยทรัพยากร

วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาวิทยาศาสตรมหาบัณฑิต
สาขาวิชา เทคโนโลยีชีวภาพ

คณะวิทยาศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย

ปีการศึกษา 2548

ISBN 974-14-2293-8

ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย

**MUTAGENIC AND ANTIMUTAGENIC ACTIVITIES OF
EXTRACTS FROM WHITE KWAO KRUA *Pueraria mirifica*,
RED KWAO KRUA *Butea superba* AND BLACK KWAO KRUA
Mucuna collettii BY AMES TEST**

Mr. Kade Pulcharoen

ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

**A Thesis Submitted in Partial Fulfillment of the Requirements
for the Degree of Master of Science Program in Biotechnology**

Faculty of Science

Chulalongkorn University

Academic Year 2005

ISBN 974-14-2293-8

Thesis Title MUTAGENIC AND ANTIMUTAGENIC ACTIVITIES OF
EXTRACTS FROM WHITE KWAO KRUA *Pueraria
mirifica*, RED KWAO KRUA *Butea superba* AND BLACK
KWAO KRUA *Mucuna collettii* BY AMES TEST

By Mr. Kade Pulcharoen

Program in Biotechnology

Thesis Advisor Associate Professor Wichai Cherdshewasart, D.Sc.

Thesis Co-advisor Associate Professor Sirirat Rengpipat, Ph.D.

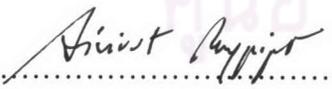
Accepted by the Faculty of Science, Chulalongkorn University in Partial
Fulfillment of the Requirements for the Master's Degree

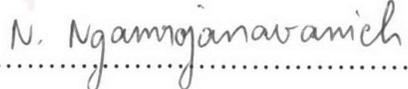

.....Dean of the Faculty of Science
(Professor Piamsak Menasveta, Ph.D.)

THESIS COMMITTEE


.....Chairman
(Associate Professor Kingkaew Wattanasermkit, Ph.D.)


.....Thesis Advisor
(Associate Professor Wichai Cherdshewasart, D.Sc.)


.....Thesis Co-advisor
(Associate Professor Sirirat Rengpipat, Ph.D.)


.....Member
(Associate Professor Nattaya Ngamrojanavanich, Ph.D.)


.....Member
(Assistant Professor Chanpen Chanchao , Ph.D.)

เกศ พูลเจริญ: ฤทธิ์ก่อกลายพันธุ์และฤทธิ์ต้านก่อกลายพันธุ์ของสารสกัดกวาวเครือขาว *Pueraria mirifica* กวาวเครือแดง *Butea superba* และกวาวเครือดำ *Mucuna collettii* ด้วยวิธีทดสอบแบบแอมส์ (MUTAGENIC AND ANTIMUTAGENIC ACTIVITIES OF EXTRACTS FROM WHITE KWAO KRUA *Pueraria mirifica*, RED KWAO KRUA *Butea superba* AND BLACK KWAO KRUA *Mucuna collettii* BY AMES TEST)

อ. ที่ปรึกษา: รศ.ดร.วิชัย เชิดชูวิทยาศาสตร์, อ. ที่ปรึกษาร่วม: รศ.ดร.ศิริรัตน์ เร่งพิพัฒน์
จำนวน 153 หน้า ISBN 974-14-2293-8

ทดสอบความเป็นพิษต่อ *Salmonella* Typhimurium TA98 และTA100 ของสารสกัดจากกวาวเครือทั้ง 3 ชนิด พบว่าสารสกัดจากกวาวเครือดำ มีความเป็นพิษมากที่สุด ต่อ *S. Typhimurium* TA98 ผลการทดสอบฤทธิ์ก่อกลายพันธุ์ และฤทธิ์ต้านก่อกลายพันธุ์ ด้วยวิธีทดสอบแบบแอมส์ ที่ ระดับความเข้มข้น 0.625, 1.25 และ 2.5 mg/plate (100 μ l) ด้วย *S. Typhimurium* TA98 และTA100 ในภาวะที่มีและไม่มีเอนไซม์ S9 พบว่าสารสกัดจากกวาวเครือขาว 28 จังหวัด กวาวเครือแดง 24 จังหวัด และกวาวเครือดำ 4 จังหวัด ไม่มีฤทธิ์ก่อกลายพันธุ์ ยกเว้นสารสกัดกวาวเครือแดงจากจังหวัดราชบุรีเมื่อนำมาทดสอบด้วย *S. Typhimurium* TA98 ที่ความเข้มข้น 2.5 mg/plate (100 μ l) ในภาวะที่ไม่มีเอนไซม์ S9 ซึ่งมีความแตกต่างจากกลุ่มควบคุมอย่างมีนัยสำคัญ ($P < 0.05$) การทดสอบฤทธิ์ต้านก่อกลายพันธุ์ที่ความเข้มข้น 2.5 mg/plate (100 μ l) ต่อสารก่อกลายพันธุ์ 2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide (AF-2) 0.01 μ g/plate (100 μ l) ด้วย *S. Typhimurium* TA 100 ในภาวะที่ไม่มีเอนไซม์ S9 พบว่าเปอร์เซ็นต์ยับยั้งการก่อกลายพันธุ์ด้วยสาร AF-2 ในกวาวเครือขาวจากจังหวัดอุทัยธานี กวาวเครือแดงจากจังหวัดกาญจนบุรี และกวาวเครือดำจากจังหวัดเชียงใหม่ มีค่าเท่ากับ 25.75, 53.36 และ 50.55 ตามลำดับ เมื่อทดสอบด้วยสารก่อกลายพันธุ์ Benzo(a)Pyrene (B(a)P) 5 mg/plate ในภาวะที่มีเอนไซม์ S9 พบว่า กวาวเครือขาวจากจังหวัดสุโขทัย กวาวเครือแดงจากจังหวัดชลบุรี และกวาวเครือดำจากจังหวัดกาญจนบุรี สามารถยับยั้งการก่อกลายพันธุ์ด้วยสาร B(a)P เท่ากับ 55.95, 88.61 และ 92.87 เปอร์เซ็นต์ตามลำดับ และจากการยืนยัน ฤทธิ์ก่อกลายพันธุ์ และฤทธิ์ต้านก่อกลายพันธุ์ ที่ ระดับความเข้มข้น 2.5, 5 และ 10 mg/well (10 μ l) เมื่อทดสอบด้วย *Bacillus subtilis* H17 and H45 ในภาวะที่ไม่มีเอนไซม์ S9 พบว่าสารสกัดกวาวเครือทั้งหมด ไม่มีฤทธิ์ก่อกลายพันธุ์ และตรวจพบสารสกัดกวาวเครือดำจากจังหวัดเชียงราย กวาวเครือแดงจากจังหวัดเลย ที่ระดับความเข้มข้น 2.5 mg/well สามารถยับยั้งการก่อกลายพันธุ์ของสาร AF-2 0.1 μ g/well (10 μ l) อย่างมีนัยสำคัญ ($P < 0.05$) และกวาวเครือขาวจากจังหวัดอุทัยธานี ที่ระดับความเข้มข้น 0.1 mg/well สามารถยับยั้ง การก่อกลายพันธุ์ของสาร AF-2 อย่างมีนัยสำคัญ ($P < 0.05$) โดยสรุปไม่พบฤทธิ์ก่อกลายพันธุ์ของสารสกัดจากกวาวเครือทั้งหมด แต่พบฤทธิ์ต้านก่อกลายพันธุ์ในกวาวเครือบางตัวอย่าง และพบฤทธิ์ต้านก่อกลายพันธุ์แรงที่สุดจากกวาวเครือดำ

สาขาวิชา.....เทคโนโลยีชีวภาพ.....ลายมือชื่อนิสิต.....

ปีการศึกษา.....2548.....ลายมือชื่ออาจารย์ที่ปรึกษา.....

ลายมือชื่ออาจารย์ที่ปรึกษาร่วม.....

4672212223: MAJOR BIOTECHNOLOGY

KEYWORD: Mutagenicity / Antimutagenicity / *Pueraria mirifica* / *Butea superba* / *Mucuna collettii* / Ames Test / *rec* assay

KADE PULCHAROEN: MUTAGENIC AND ANTIMUTAGENIC ACTIVITIES OF EXTRACTS FROM WHITE KWAO KRUA *Pueraria mirifica*, RED KWAO KRUA *Butea superba* AND BLACK KWAO KRUA *Mucuna collettii* BY AMES TEST. THESIS ADVISOR: ASSOC. PROF. WICHAI CHERDSHEWASART, D.Sc., THESIS CO-ADVISOR: ASSOC. PROF. SIRIRAT RENGPIPAT, Ph.D. 153 pp. ISBN 974-14-2293-8

Cytotoxicity evaluation of the 3 plant extracts on *Salmonella* Typhimurium TA98 and TA 100 revealed that *Mucuna collettii* extract exhibited the strongest cytotoxicity on *S. Typhimurium* TA98. Mutagenic and antimutagenic activities of *Pueraria mirifica*, *Butea superba* and *Mucuna collettii* collected from 28, 24 and 4 provinces, respectively were tested. Their extracts were evaluated by Ames test using *S. Typhimurium* strain TA98 and TA100 under the absence and presence of metabolic activity conditions at the concentration of 0.625, 1.25 and 2.5 mg/plate (100 μ l) in comparison with the control. No mutagenicity was induced by all plant extracts except *B. superba* collected from Ratchaburi province which exhibited mutagenicity ($P < 0.05$) in *S. Typhimurium* strain TA98 under the absence of metabolic activity condition at the concentration of 2.5 mg/plate (100 μ l). In the test with mutagenicity on *Salmonella* strains, all plant extracts exhibited significant inhibition ($P < 0.05$) against 2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide (AF-2) and Benzo(a)Pyrene (B(a)P). *P. mirifica* collected from Uthai Thani, *B. superba* collected from Kanchanaburi, *M. collettii* collected from Chiang Mai inhibited mutagenicity induced by AF-2 0.01 μ g/plate (100 μ l) on *S. Typhimurium* strain TA100 at the inhibition percentage of 25.72, 53.36 and 50.55, respectively. *P. mirifica* collected from Sukhothai, *B. superba* collected from Chonburi, *M. collettii* collected from Kanchanaburi inhibited mutagenicity induced by B(a)P 5 μ g/plate on *S. Typhimurium* strain TA100 at the inhibition percentage of 55.95, 88.61 and 92.87, respectively. The mutagenicity and antimutagenicity assay of the plant extracts were confirmed in *rec* assay with *Bacillus subtilis* strain H17 and H45 under the absence of metabolic activity conditions. All plant extracts at the concentration of 2.5, 5 and 10 mg/well (10 μ l) exhibited no mutagenicity. *Mucuna collettii* collected from Chiang Rai and *Butea superba* collected from Loei exhibited significant inhibition ($P < 0.05$) against AF-2 0.1 μ g/well (10 μ l) at the concentration of 2.50 mg/well (10 μ l), and *P. mirifica* collected from Uthai Thani exhibited significant inhibition ($P < 0.05$) against AF-2 at the concentration of 10 mg/well (10 μ l). In summary, all Kwao Krua in this study showed no mutagenicity but antimutagenicity was detected in certain samples. The strongest antimutagenicity was found in *M. collettii*.

Field of study...BiotechnologyStudent's signature.....*Kade Pulcharoen*.....

Academic.....2005.....Advisor's signature.....*Wichai Cherdshewasart*.....

Co-advisor's signature*Sirirat Rengpipat*.....

ACKNOWLEDGEMENTS

I would like to express my deeply appreciation and grateful thanks to my advisor, Associate Professor Dr. Wichai Cherdshewasart, Department of Biology, for his extremely helpful guidance, suggestions, continual encouragement and interest throughout this study as well as for his huge collected plant samples from his own research. Especially I would like to really thank Associate Professor Dr. Sirirat Rengpipat, Department of Microbiology, with her kindness guidance, Associate Professor Dr. Kingkaew Wattanasermkit, Department of Biology, Associate Professor Dr. Nattaya Ngamrojanavanich, Program in Biotechnology, and Assistant Professor Dr. Chanpen Chanchao, Department of Biology, for their valuable comments and all suggestions.

Special thanks for Department of Microbiology and Biology for laboratory facilities, Program in Biotechnology, Faculty of Science, Chulalongkorn University for access to use the necessary instruments for my thesis.

I am indebted to Mrs. Sunan Limthiamcharoen and her colleague to excellent assistance for my study. Sincere thanks for Ms. Rattana Panriansaen, Ms. Sutijit Sriwatcharakul, Ms. Yosaporn Kitsamai, Ms. Wanrawee Sungkapong, Ms. Wandee Sutijit and the members of Kwao Krua Research Laboratory, Department of Biology for their generous help. Thanks for Mr. Adipol Dilokpimol for his warm assistance. Special thanks for Dr. Wannee Kusamran, Ms. Anong Tepsuwan and Ms. Nantana Meesiriphun, National Cancer Institute, for their helpful guidance of Ames's technique.

I really express my whole-heartedly appreciation to my parents, my family members and all of my friends; Ache, Kaew, Mc, Orn, I.C and Bee who have never left me alone.

This project was granted by Central of Excellence in Biodiversity, Faculty of Science, Chulalongkorn University.

CONTENTS

	Page
THAI ABSTRACT.....	iv
ENGLISH ABSTRACT.....	v
ACKNOWLEDGEMENTS.....	vi
CONTENTS.....	viii
LIST OF TABLES.....	xii
LIST OF FIGURES.....	xv
LIST OF ABBREVIATION.....	xix
CHAPTER I INTRODUCTION.....	1
CHAPTER II LITERATURE REVIEW.....	3
2.1 Kwao Krua plants.....	3
2.1.1 White Kwao Krua <i>Pueraria mirifica</i>	3
2.1.1.1 Botanical characteristic of <i>P. mirifica</i>	3
2.1.1.2 Bioactivity and pharmacological effects of chemical constituents in <i>P. mirifica</i>	4
2.1.1.3 Pharmacological effects of <i>P. mirifica</i>	8
2.1.1.4 Safety test of <i>P. mirifica</i>	9
2.1.1.5 Clinical trial effects of <i>P. mirifica</i>	9
2.1.2 Red Kwao Krua <i>Butea superba</i>	10
2.1.2.1 Botanical characteristic of <i>B. superba</i>	10
2.1.2.2 Bioactivity and pharmacological effects of chemical constituents in <i>B. superba</i>	11
2.1.2.3 Pharmacological effects of <i>B. Superba</i>	12
2.1.2.4 Safety of <i>B. superba</i>	13
2.1.2.5 Clinical trial effects of <i>B. Superba</i>	13
2.1.3 Black Kwao Krua <i>Mucuna colletti</i>	14
2.1.3.1 Botanical characteristic of <i>M. collettii</i>	14
2.1.3.2 Bioactivity and pharmacological effects of chemical constituents in <i>M. collettii</i>	14
2.1.3.3 Pharmacological effects of <i>M. collettii</i>	16

	Page
2.1.3.4 Safety of <i>M. collettii</i>	16
2.2 Phytoestrogen.....	16
2.2.1 Flavonoids and Isoflavones.....	16
2.2.1.1 Source of Isoflavones.....	16
2.2.1.2 Isoflavone metabolism.....	17
2.2.1.3 Bioactivity and pharmacological effects.....	19
2.2.1.3.1 Hormonal effects.....	19
2.2.1.3.2 Anticarcinogenic effect	19
2.2.1.3.3 Other effects.....	20
2.3 Mutagenesis	20
2.3.1 Type of alterations in the genetic material.....	20
2.3.2 Mutagenicity and carcinogenicity testing.....	21
2.4 The Ames <i>Salmonella</i> /microsome mutagenicity assay	22
2.4.1 Metabolic activation systems.....	22
2.4.1.1 Oxidative metabolism.....	23
2.4.1.2 Reductive metabolism.....	23
2.4.2 The <i>Salmonella</i> tester strains.....	23
2.4.3 Construction of base-specific <i>Salmonella</i> tester.....	27
2.4.4 Assay procedures.....	27
2.4.4.1 The standard plate incorporation assay.....	27
2.4.4.2 The preincubation assay.....	28
2.4.4.4 Positive and negative control for diagnostic mutagens.....	28
2.4.4.5 Dose selection.....	28
2.5 Application of Ames test in plant medicine and phytoestrogens.....	29
CHAPTER III MATERIALS AND METHODS.....	33
3.1 The plant materials.....	33
3.1.1 Plant powder preparation.....	33
3.1.2 Plant crude extraction.....	33
3.2 Bacterial strains.....	34
3.3 Chemical and reagents.....	34

	Page
3.4 Mammalian liver enzyme preparation.....	35
3.5 Experimental protocol.....	35
3.5.1 Reverse mutation assay.....	35
3.5.1.1 Preparation of bacterial cultures.....	35
3.5.1.2 Preparation of nutrient agar	35
3.5.1.2.1 Minimal agar plate.....	35
3.5.1.2.2 Top agar.....	35
3.5.1.3 Preparation of S9 mix for mutagenicity and antimutagenicity assay.....	36
3.5.1.4 Experiment I - Survival test	36
3.5.1.5 Experiment II - Mutagenic activity of the plant extracts toward <i>S. Typhimurium</i> TA98 and TA 100 in the absence and presence of metabolic activation.....	37
3.5.1.6 Experiment III - Antimutagenic activity of the plant extracts toward <i>S. Typhimurium</i> TA98 and TA 100 in the absence and presence of metabolic activation.....	37
3.5.1.7 Interpretation.....	38
3.5.1.8 Statistical analysis.....	39
3.5.2 Forward mutation assay.....	40
3.5.2.1 Preparation of plant materials.....	40
3.5.2.2 Preparation of cultures.....	40
3.5.2.3 Experiment I - Mutagenic activity of the plant extracts toward <i>B. subtilis</i> H17 (<i>rec</i> ⁺) and M45 (<i>rec</i> ⁻) in the absence of metabolic activation.....	40
3.5.2.4 Experiment II - Antimutagenic activity of the plant extracts toward <i>B. subtilis</i> H17 (<i>rec</i> ⁺) and M45 (<i>rec</i> ⁻) in the absence of metabolic activation.....	41
3.5.2.5 Interpretation.....	41
3.5.2.6 Statistical analysis.....	41
CHAPTER IV RESULTS.....	42

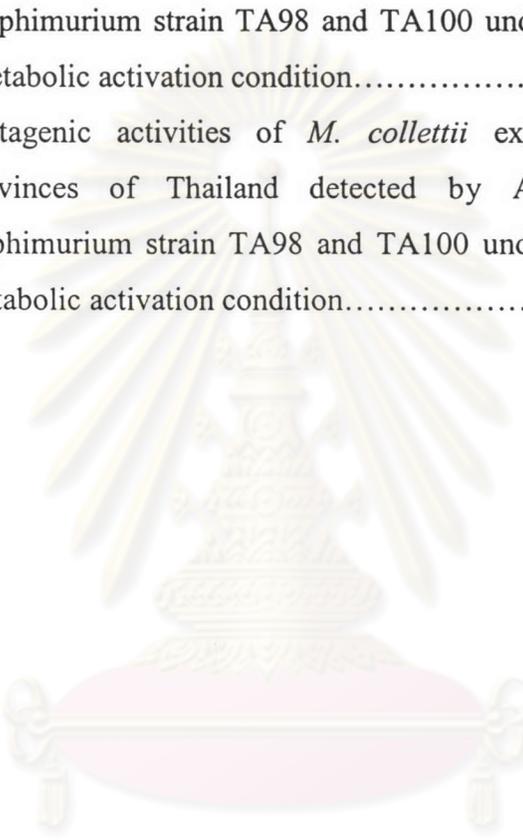
	Page
4.1 Reverse mutation assay	42
4.1.1 Survival Test.....	42
4.1.2 Mutagenic activity of the plant extracts toward <i>S. Typhimurium</i> TA98 and TA 100 in the absence and presence of metabolic activation.....	43
4.1.3 Antimutagenic activity of the plant extracts toward <i>S.</i> Typhimurium TA98 and TA 100 in the absence and presence of metabolic activation.....	44
4.1.3.1 Antimutagenic activity of the <i>P. mirifica</i> extracts.....	44
4.1.3.2 Antimutagenic activity of the <i>B. superba</i> extracts.....	57
4.1.3.3 Antimutagenic activity of the <i>M. collettii</i> extracts.....	69
4.1.3.4 Comparison of antimutagenic activity of plant extracts detected by Ames Test.....	77
4.2 Forward mutation assay.....	79
4.2.1 Mutagenic activity of the plant extracts toward <i>B. subtilis</i> H17 (<i>rec</i> ⁺) and M45 (<i>rec</i> ⁻) in the absence of metabolic activation.....	79
4.2.2 Antimutagenic activity of the plant extracts toward <i>B. subtilis</i> H17 (<i>rec</i> ⁺) and M45 (<i>rec</i> ⁻) in the absence of metabolic activation..	80
4.3 The antimutagenic activity correlation of <i>P. mirifica</i> , <i>B. superba</i> and <i>M.</i> <i>collettii</i> from this study and previous ones.....	80
CHAPTER V DISCUSSION AND CONCLUSION	85
REFERENCES.....	89
APPENDICES.....	107
APPENDIX A.....	108
APPENDIX B.....	152
BIOGRAPHY.....	153

LIST OF TABLES

		Page
Table 2.1	Summary of the chemical constituents of <i>P. mirifica</i>	5
Table 2.2	The structure and bioactivity effects of chemical constituents in <i>P. mirifica</i>	6
Table 2.3	Summary of the chemical constituents of <i>B. superba</i>	9
Table 2.4	The structure and bioactivity effects of chemical constituents in <i>B. superba</i>	11
Table 2.5	The structure and bioactivity effects of chemical constituents in <i>M. collettii</i>	15
Table 2.6	Classification and sources of phytoestrogens.....	17
Table 2.7	Selected shorted-term tests for detection of chemical carcinogens and promoting agents.....	21
Table 2.8	Genotype of the most commonly used <i>Salmonella</i> tester strains.....	23
Table 2.9	Reversion of tester strains for standard mutagens.....	29
Table 2.10	The screening for antimutagenic activities in plant medicine by Ames Test.....	30
Table 2.11	The screening for antimutagenic activities of flavonoids naturally occurring in medical herbs by Ames Test.....	32
Table 4.1	Survival test of the plant extracts on <i>S. Typhimurium</i> TA98 and TA100.....	43
Table 4.2	Antimutagenic activities of <i>P. mirifica</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition.....	47
Table 4.3	Antimutagenic activities of <i>B. superba</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition.....	59

	Page
Table 4.4	Antimutagenic activities of <i>M. collettii</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition..... 70
Table 4.5	Antimutagenic activity of the kwao krua plant extracts against direct-acting and indirect-acting mutagens detected by <i>S. Typhimurium</i> strain TA98 and TA100..... 78
Table 4.6	Mutagenicity effects of the plant extracts tested by <i>Rec</i> assay with <i>B. subtilis</i> var.H17 (<i>Rec</i> ⁺) and M45 (<i>Rec</i> ⁻) on non-metabolic activation condition..... 79
Table 4.7	Antimutagenicity effects of the plant extracts tested by <i>Rec</i> assay with <i>B. subtilis</i> var.H17 (<i>Rec</i> ⁺) and M45 (<i>Rec</i> ⁻) on non-metabolic activation condition..... 80
Table 4.8	The correlation between antimutagenic activity at the concentration 2.50 mg/plate of <i>P. mirifica</i> samples determined by Ames Test and Isoflavone contents (Subtang, 2002) 81
Table 4.9	The correlation between antimutagenic activity at the concentration 2.50 mg/plate of <i>P. mirifica</i> samples determined by Ames test, and antioxidant activity; and anti-proliferatin effect on MCF-7 cell (Sutjit, 2003; Trisap, 2003) 82
Table 4.10	The correlation of antimutagenic activity at the concentration 2.50 mg/plate of <i>B. superba</i> samples determined by Ames test, antioxidant activity and anti-proliferation effect on MCF-7 cell (Sutjit, 2003; Trisap, 2003) 83
Table 4.11	The correlation of antimutagenic activity at the concentration 2.50 mg/plate of <i>M. collettii</i> samples determined by Ames test, antioxidant activity and anti-proliferation effect on MCF-7 cell (Trisap, 2003; Sutjit, 2003) 84
Table 5.1	Spontaneous revertant control values.....

	Page
Table 5.2	Mutagenic activities of <i>P. mirifica</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition..... 124
Table 5.3	Mutagenic activities of <i>B. superba</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition..... 138
Table 5.4	Mutagenic activities of <i>M. collettii</i> extracts from different provinces of Thailand detected by Ames test using <i>S. Typhimurium</i> strain TA98 and TA100 under non metabolic and metabolic activation condition..... 150



ศูนย์วิทยทรัพยากร
จุฬาลงกรณ์มหาวิทยาลัย

LIST OF FIGURES

		Page
Figure 2.1	(a) Leaves, (b) flowers, (c) tuberous roots and (d) pods of <i>P. mirifica</i> from Chiang Mai Province, photos courtesy by W. Cherdshewasart.....	4
Figure 2.2	(a) Leaves, (b) tuberous root, (c) tuberous cross section and (d) flowers of <i>B. superba</i> from Lumpang Province, photos courtesy by W. Cherdshewasart.....	10
Figure 2.3	(a) Leaves and (b) stems of <i>M. collettii</i> , Photos of <i>M. collettii</i> from Chiang Rai Province, courtesy by W. Cherdshewasart.....	14
Figure 2.4	Metabolism of daidzein and genistein in mammals. The glycosidic forms present in plants (daidzein and genistein) are cleaved in the gastrointestinal tract, resorbed and metabolized by mammalian enzymes and gut bacteria.....	18
Figure 2.5	Schematic overview of <i>Salmonella</i> Typhimurium metabolism and the effect of non repaired point mutation in the synthesis of histidine.....	26
Figure 3.1	Diagram depicting the steps involved in the preincubation assay...	38
Figure 4.1	The <i>B. superba</i> extracted which collected from Ratchaburi province shown mutagenicity on <i>S. Typhimurium</i> strain TA100 at the concentration 2.50 mg/plate under non metabolic active condition.....	44
Figure 4.2	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 0.625 mg/plate.....	51
Figure 4.3	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 1.25 mg/plate.....	52

	Page	
Figure 4.4	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 2.50 mg/plate.....	53
Figure 4.5	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 0.625 mg/plate.....	54
Figure 4.6	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 1.25 mg/plate.....	55
Figure 4.7	Antimutagenic activities of <i>P. mirifica</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 2.50 mg/plate.....	56
Figure 4.8	Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 0.625 mg/plate.....	56
Figure 4.9	Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 1.25 mg/plate.....	63
Figure 4.10	Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 2.50 mg/plate.....	64

	Page
Figure 4.11 Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 0.625 mg/plate.....	65
Figure 4.12 Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 1.25 mg/plate.....	66
Figure 4.13 Antimutagenic activities of <i>B. superba</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 2.50 mg/plate.....	67
Figure 4.14 Antimutagenic activities of <i>M. collettii</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 0.625 mg/plate.....	71
Figure 4.15 Antimutagenic activities of <i>M. collettii</i> extracts against a.) direct-acting of AF-2 0.1 µg/plate and b.) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 1.25 mg/plate.....	72
Figure 4.16 Antimutagenic activities of <i>M. collettii</i> extracts against a) direct-acting of AF-2 0.1 µg/plate and b) indirect-acting of B(a)P 10 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA98 at concentration 2.50 mg/plate.....	73
Figure 4.17 Antimutagenic activities of <i>M. collettii</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 0.625 mg/plate.....	74

	Page
Figure 4.18	Antimutagenic activities of <i>M. collettii</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 1.25 mg/plate..... 75
Figure 4.19	Antimutagenic activities of <i>M. collettii</i> extracts against a) direct-acting of AF-2 0.01 µg/plate and b) indirect-acting of B(a)P 5 µg/plate mutagens analyzed by <i>S. Typhimurium</i> strain TA100 at concentration 2.50 mg/plate..... 76
Figure 4.20	Antimutagenic activities of kwao krua plant extracts against direct-acting and indirect-acting mutagens detected by Ames test at the concentration (a) 0.625, (b) 1.25 and (c) 2.50 mg/plate..... 78
Figure 5.1	The each single revertant colonies from ampicillin plate for confirm genotype; (a,c) Histidine dependence (<i>his</i>), (b,d) Biotin dependence (<i>bio</i>), Histidine and biotin dependence (<i>his, bio</i>) of <i>S. Typhimurium</i> strains TA98 and TA100..... 117
Figure 5.2	The each of single revertant colonies from ampicillin plate for confirm genotype; (a,c) <i>rfa</i> mutation (non clear zone) and R-factor (clear zone), (b,d) <i>uvrB</i> mutation of <i>S. Typhimurium</i> strains TA98 and TA100..... 118
Figure 5.3	The spontaneous revertant colonies (a,c) and revertants colonies which induced by standard mutagen (b) AF-2 0.1 µg/plate, (d) AF-2 0.01 0.1 µg/plate, (c) B(a)P 10 µg/plate, (d) B(a)P 5 µg/plate on of <i>S. Typhimurium</i> strains TA98 and TA100..... 120
Figure 5.4	Protein content of S-9 fraction sample calculated from standard protein c curve, w hich t he S -9 fraction s ample h ave p rotein 4 1.86 mg/ml..... 123

LIST OF ABBREVIATION

°C	Degree Celsius
AF-2	2-(2-furyl)-3-(5-nitro-2-furyl)acrylamide
B(a)P	Benzo(a)Pyrene
BW	Body weight
°C	Degree Celsius
cm	Centrimetre
DMSO	Dimethyl sulphoxide
DNA	Deoxy ribonucleic acid
hr	hour
IC ₅₀	Median Inhibition Concentration
Kg	Kilogram
L	Liter
LD ₅₀	Median Lethal Dose
LC ₅₀	Median Lethal Concentration
MGA	Minimal Glucose Agar
MW.	Molecular Weight
m	Metre
mm	Millimetre
μM	Micromolor
μg	Microgram
μl	Microliter
ml	Milliliter
min	Minute
ng	Nanogram
PI	Percentage Inhibition
S.E.M.	Standard Mean Error