

REFERENCES

- Abdel-Rahman, S. H., Kanazawa, A., and Teshima, S. I. 1979. Effects of dietary carbohydrate on the growth and the levels of the hepatopancreatic glycogen and serum glucose of prawn. Bulletin of the Japanese Society of Scientific Fisheries. 45(12): 1491-1494.
- Adron, J. W., and Mackie, A. M. 1978. Studies on the chemical nature of feeding stimulants for rainbow trout *Salmo gairdneri*. Journal of Fish Biology. 12: 303.
- Akiyama, D. M., and Dominy, W. G. 1989. Penaeid shrimp nutrition for the commercial feed industry. Texas Shrimp Farming Manual. Vol. 1. Corpus Christi, Texas, USA: Extension Service Texas A&M University.
- Alava, V. R., and Lim, C. 1983. The quantitative dietary protein requirements of *Penaeus monodon* juveniles in a controlled environment. Aquaculture. 30: 53-61.
- Alava, V. R., and Pascual, F. P. 1987. Carbohydrate requirements of *Penaeus monodon* (Fabricius) juveniles. Aquaculture. 61: 211-217.
- Alfaro, J. 2001. Controlled reproduction of penaeid shrimp: a contribution to its improvement. PhD thesis, Wageningen University.
- Ali, S. A. 1882. Effect of carbohydrate (starch) level in purified diets on growth of *P. indicus*. Indian Journal of Fish. 29: 201-208.
- Amend, D. F. 1981. Potency testing of fish vaccines. In D. P. Anderson, and W. Hennessen (eds.), Fish biologics: serodiagnosis and vaccines. Developments in biological standardization, pp. 447-454. Basel, Switzerland: S. Karger.
- Anderson, D. P., and Siwicki, A. K. 1994. Duration of protection against *Aeromonas salmonicida* in brook trout immunostimulated with glucan or chitosan by injection or immersion. Progressive Fish-Culturist. 56: 258-261.
- Anderson, D. P., Siwicki, A. K., and Rumsey, G. L. 1995. Injection or immersion delivery of selected immunostimulants to trout demonstrate enhancement of nonspecific defense mechanisms and protective immunity. In M. Shariff, R. P. Subasighe, and J. R. Arthur (eds.), Diseases in Asian Aquaculture II, pp. 413-426. Philippines: Fish Health Section, Asia Fisheries Society.

- Anderson, I. G., Shamsudin, M. N., Shariff, M. et al. 1988. Bacterial septicemia in juvenile tiger shrimp, *Penaeus monodon*, cultured in Malaysian brackish water ponds. *Asian Fish. Science.* 2: 93-108.
- Andrews, J. W., and Sick, L. V. 1972. Studies on the nutritional requirements of penaeid shrimp. *Proceedings of the World Mariculture Society.* 3: 403-414.
- Andrews, J. W., Sick, L. V., and Baptist, G. J. 1972. The influence of dietary protein and energy level on growth and survival of penaeid shrimp. *Aquaculture.* 1: 341-347.
- Anon. 1991 Ecuador's shrimp culture industry. *Marine Fisheries Review.* 52(3): 38-39.
- Arkarajamon, A. 1991. *Histology of tiger shrimp (Penaeus monodon Fabricius)*. Master thesis. Department of Marine Science, Faculty of Fisheries, Kasetsart University.
- Armstrong, P. B., and Quigley, J. P. 1999. α_2 -macroglobulin: an evolutionarily conserved arm of the innate immune system. *Developmental and Comparative Immunology.* 23: 375-390.
- Baba, M., Nakajima, M., Schols, D. et al. 1988. Pentosan polysulfate, a sulfated oligosaccharide, is a potent and selective anti-HIV agent *in vitro*. *Antiviral Research.* 9(6): 335-343.
- Bachère, E. 2000. Shrimp immunity and disease control. *Aquaculture.* 191: 3-11.
- Bachère, E., Mialhe, E., and Rodriguez, J. 1995. Identification of defense effctors in the hemolymph of crustaceans with particular reference to the shrimp *Penaeus japonicus* (Bate.): prospects and applications. *Fish and shellfish immunology.* 5: 597-612.
- Bagasra, O., and Lischner, H. W. 1988. Activity of dextran sulfate and other polyanionic polysaccharides against human immunodeficiency virus. *Journal of Infectious Diseases.* 158: 1084-1087.
- Baily-Brock, J. H., and Moss, S. M. 1992. Penaeid taxonomy, biology and zoogeography. In A. W. Fast, and L. J. Lester (eds.), *Marine shrimp culture: principle and practices*, pp. 9-28. Amsterdam: Elsevier Science Publishers.
- Balazs, G. H., Ross, E., and Brooks, C. C. 1973. Preliminary studies on the preparation and feeding of crustacean diets. *Aquaculture.* 2: 369-377.

- Balazs, G. H., Ross, E., Brooks, C. C. et al. 1974. Effect of protein source and level on growth of the captive fresh water prawn, *Macrobrachium rosenbergii*. Proceedings of the World Mariculture Society. 3: 145-157.
- Baliao, D. D. 2000. Environment-friendly schemes in intensive shrimp farming, pp. 1-25. Philippines: Southeast Asian Fisheries Development Center Association of Southeast Asian Nations.
- Bardach, J. E., Ryther, J. H., and McLarney, W. O. 1972. Aquaculture: The farming and husbandry of freshwater and marine organisms. New York, USA: John Wiley&Sons.
- Barnes, H., Morris, R. J., and Achituv, Y. 1978. Changes in the biochemical composition of *Balanus balanoides* (L.) under experimental conditions: The effect of a starch diet. Journal of Experimental Marine Biology and Ecology. 31(3): 267-281.
- Bauchau, A. G. 1981. Crustaceans. In N. A. Ratcliffe, and A. F. Rowley (eds.), Invertebrate blood cells, pp. 385-420. London and New York: Academic Press.
- Bell, T. A., and Lightner, D. V. 1988. A handbook of normal penaeid shrimp histology. p.114. Baton Rouge: The World Aquaculture Society.
- Bilan, M. I., Grachev, A. A., Ustuzhanina, N. E. et al. 2002. Structure of a fucoidan from the brown seaweed *Fucus evanescens* C. Ag. Carbohydrate Research. 337(8): 719-730.
- Blanshard, J. M. V., and Mitchell, J. R. 1979. Polysaccharide in food. Norfolk: Fakenham Press.
- Boonyaratpalin, M. 1998. Nutrition of *Penaeus merguiensis* and *Panaeus idicus*. Review in Fisheries Science. 6(1-2): 69-78.
- Boonyaratpalin, S., Boonyaratpalin, M., Supamattaya, K. et al. 1995. Effects of peptidoglycan on growth, survival, immune response, and tolerance to stress in black tiger shrimp, *Penaeus monodon*. In M. Shariff, J. R. Arthur, and R. P. Subasinghe (eds.), Disease in Asia Aquaculture II, pp. 467-477. Philipines: Fish Health Section, Asian Fisheries Society.
- Bradford, M. M. 1975. A rapid and sensitive method for quatitation of microgram quantities of protein utilizing the principle of protein-dye binding. Analytical Biochemistry. 72: 248-254.

- Brock, T. D., Madigan, M. T., Martinko, J.M. et al. 1994. Biology of Microorganisms. 7 th ed., pp.118-124. Prentice Hall: Englewood Cloffs, N. J.
- Brusca, R. C., and Brusca, G. J. 1990. Invertebrates, p. 922. Massachusetts, USA: Sinauer Associates.
- Callahan, L. N., Phelan, M., Mallinson, M. et al. 1991. Dextran sulfate blocks antibody binding to the principal neutralizing domain of human immunodeficiency virus type 1 without interfering with gp 120-CD4 interaction. Journal of Virology. 65: 1543-1550.
- Cameron, J. N., and Magnum, C. P. 1983. Environmental adaptations of the respiratory system: Ventilation, circulation, and oxygen transport. In F. J. Vernberg, and W. B. Vernberg (eds.), The Biology of Crustacea, pp. 43-63. New York, USA: Academic Press.
- Capuzzo, J. M. 1982. Crustacean bioenergetics: The role of environmental variables and dietary levels of macronutrients on energetic efficiencies. Proceedings 2nd Int'l Conference on Aquaculture Nutrition, Biochemistry and Physiological Approaches to Shellfish Nutrition, pp.71-86. 27-29 October 1982. Rehoboth Beach, Delaware, USA.
- Carlucci, M. J., Ciancia, M., and Matulewicz, M. C. 1999. Antiherpetic activity and mode of action of natural carrageenans of diverse structural types. Antiviral Research. 43(2): 93-102.
- Castell, J. D. 1979. Review of lipid requirements of finfish. In J. E. Halver, and K. Tiews (eds.), Finfish nutrition and fish feed technology, pp. 59-84. Hamburg, Germany: Berlin, Federal Republic.
- Cerenius, L., and Söderhäll, K. 2004. The prophenoloxidase-activating system in invertebrates. Immunological Reviews. 198: 116-126.
- Chang, C. F., Chen, H. Y., Su, M. S. et al. 2000. Immuno modulation by dietary β -1,3-glucan in the brooders of the black tiger shrimp *Penaeus monodon*. Fish and Shellfish Immunology. 10: 505-514.
- Chang, C. F., Su, M. S., Chen, H. Y. et al. 1999. Effect of dietary β -1,3-glucan on resistance to white spot syndrome virus (WSSV) in postlarval and juvenile *Penaeus monodon*. Diseases of Aquatic Organisms. 36: 163-168.

- Chang, C. F., Su, M. S., Chen, H. Y. et al. 2003. Dietary β -1,3-glucan effectively improves immunity and survival of *Penaeus monodon* challenged with white spot syndrome virus. *Fish and Shellfish Immunology*. 15: 297-310.
- Chang, P. S., Lo, F. C., and Kou, G. H. 1996. Identification of white spot syndrome associated baculovirus (WSBV) target organs in the shrimp *Penaeus monodon* by in situ hybridization. *Diseases of Aquatic Organisms*. 27: 131–139
- Chansiripornchai, P., Chansiripornchai, N., and Pongsamart, S. 2004 b. Evaluation of polysaccharide extracted from the fruit-hull of durian (*Durio zibethinus* L.) on surgical wound healing in dog and cats. *The 13th Federation of Asian Veterinary Associations Congress*, p. 285. 25-27 October 2004. Millennium Seoul Hilton, Seoul Korea.
- Chansiripornchai, P., Rangsipipat, A., and Pongsamart, S. 2004 a. The treatment of surgical wounds in dogs and cats using a film dressing of polysaccharide, extractes from the fruit-hull of durian (*Durio zibethinus* L.). *The Thai Journal of Pharmaceutical Sciences*. 26(1): 65.
- Chen, K. J., and Co, W. G. 1988. *Prawn culture*. The Philippines: Chuson Printing.
- Cheng, W., Liu, C. H., Yen, S. T. et al. 2004. The immune stimulatory effect of sodium alginate on the white shrimp *Litopenaeus vannamei* and its resistance against *Vibrio alginolyticus*. *Fish and Shellfish Immunology*. 17: 41-51.
- Chihara, G. 1988. Characteristics and evaluation of lentinan as host defense potentiator. *International Journal of Immunopharmacology*. 10(supple.1): 87.
- Chizhov, A. O., Dell, A., Morris, H. R. et al. 1999. A study of fucoidan from the brown seaweed *Chorda filum*. *Carbohydrate Research*. 320(1-2): 108-119.
- Chong, K. C. 1990. Structure, conduct and performance of the Asian shrimp aquaculture industry. In M. B. New, H. de Saram, and T. Singh. (eds.), *Technical and Economic Aspects of Shrimp Farming, Proceedings of the Aquatec '90 Conference*, pp. 191-206. 11-14 June 1990. Kuala Lumpur, Malaysia.
- Chotigeat, W., Tongsupa, S., Supamataya, K. et al. 2004. Effect of fucoidan on disease resistance of black tiger shrimp. *Aquaculture*. 233: 23-30.

- Chou, H. Y., Haung, C. Y., and Kou, G. H. 1998. Studies on transmission of white spot syndrome associated baculovirus (WSBV) in *Penaeus monodon* and *Penaeus japonicus* via waterborne contact and oral ingestion. *Aquaculture*. 164: 263–276.
- Chou, H. Y., Huang, C.Y., Wang, C. H. et al. 1995. Pathogenecity of a baculovirus infection causing white spot syndrome in cultured penaeid shrimp in Taiwan. *Diseases of Aquatic Organisms*. 23: 165–173.
- Chow, K. W., and Halver, J. M. 1980. Carbohydrates. In ADCP (eds.), *Fish feed technology*. FAO, ADCP/REP/80/11. Rome, Italy.
- Chua, T. E., Paw, T. N., and Guarin, F. Y. 1989. The environmental impact of aquaculture and the effects of pollution on coastal aquaculture development. *Asia Marine Pollution Bull*. 20(7): 335–343.
- Chuang, J. L. 1990. Nutrient requirements, feeding and culturing practices of *Penaeus monodon*: A review. Basel, Switzerland: F. Hoffmann-La Roche.
- Clark, D. J., Lawrence, A. L., and Swakon, D. H. D. 1993. Apparent chitin digestibility in penaeid shrimp. *Aquaculture*. 109: 51-57.
- Cody, M. 1984. Substances without vitamin status. Machlin, In I. J. (ed.), Handbook of vitamins. New York and Basel: Marcel Dekker.
- Coelho, M. B. 1991. Effects of processing and storage on vitamin stability. *Feed International*. 12(12): 39-45.
- Combs, G. F. jr. 1988. Vitamin tolerance of animals. Corrnell Nutrition Conference. Washington, D. C.: National Academy Press.
- Corsin, F., Turnbull, J. F., Hao, N. V. et al., 2001. Risk factors associated with white spot syndrome virus infection in a Vietnamese rice-shrimp farming system. *Diseases of Aquatic Organisms*. 47: 1-12.
- Cowey, C. B., and Forster, J. R. M. 1971. The essential amino acid requirements of the prawn *Palaemon serratus*. The growth of prawns on diets containing proteins of different amino acid compositions. *Marine Biology*. 10: 77-81.
- Curatella, B., Bartolini, B., Di Caro, A. et al. 2005. Sepharose-bound, highly sulfated glycosaminoglycans can capture HIV-1 from culture medium. *Carbohydrate Research*. 340(4): 759-764.

- Davis, D. A. 1990. Dietary mineral requirements of *Penaeus vannamei*: Evaluation of the essentiality for thirteen minerals and the requirements for calcium, phosphorus, copper, iron, zinc and selenium. Ph.D. Dissertation. Texas A&M University, USA.
- De Clercq, E. 1993. Antiviral agents: characteristic activity spectrum depending on the molecular target with which they interact. Advances in Virus Research. 42: 1-55.
- Department of Internal Trade, Ministry of Commerce, Thailand. 2003. Shrimp. Thai Feed Mill Association. 20(92): 5-22.
- Deshimaru, O., and Yone, Y. 1978. Effect of dietary carbohydrate sources on the growth and feed efficiency of prawn. Bulletin of the Japanese Society of Scientific Fisheries. 44(10): 1161-1163.
- Destoumieux, D., Munoz, M., Cosseau, C. et al. 2000. Penaeidins, antimicrobial peptides with chitin-binding activity, are produced and stored in shrimp granulocytes and released after microbial challenge. Journal of Biological Chemistry. 272: 28398-28406.
- DeWalt, B. R., Vergne, P., and Hardin, M. 1996. Shrimp culture development and the environment: people, mangroves and fisheries on the Gulf of Fonseca. World Development. 24(7): 1193-1208.
- Doner, L. W., and Whistler, R. L. 1973. Industrial gums polysaccharide and their derivatives, pp. 115-120. New York: Academic Press.
- Dumitriu, S. 1998. Structural diversity and functional versatility: Polysaccharide, p. 1147. New York: Marcel Dekker.
- Dupree, H. K., and Huner, J. V. 1984. Third report to the fish farmers. Washington, D. C.: U. S. Fish and Wildlife Service.
- Durand, S., Lightner, D. V., Redman, R. M. et al. 1997. Ultrastructure and morphogenesis of white spot syndrome baculovirus (WSBV). Diseases of Aquatic Organisms. 29: 205-211.
- Ewart, J. W. 1982. The growth of juvenile oysters *Crassostrea virginica* (Gmelin) fed algae supplemented with silt or kaolin. Proceedings 2nd Conference on Aquaculture Nutrition: Biochem. and Physiol. Approaches to Shellfish Nutrition, p. 427. October 1982. Rohoboth Beach, Delaware, USA.

- Fabregas, J., Garcia, D., Fernaudez, A. M. et al. 1999. *In vitro* inhibition of the replication of haemorrhagic septicaemia virus (VHSV) and African swine fever virus (ASFV) by extracts from marine microalgae. Antiviral Research. 44: 67-73.
- Fair, P. H., Fortner, A. R., Millikin, M. R., and Sick, L. V. 1980. Effects of dietary fiber on growth, assimilation and cellulose activity of the prawn (*Macrobrachium rosenbergii*). Proceeding of the World Mariculture Society. 11: .359-381.
- Fallu, R. 1991. Abalone Farming. Fishing News Books. Oxford, England: Blackwell Science.
- Feltwell, R., and Fox, S. 1978. Practical poultry feeding. London, UK: Faber and Faber.
- Fillar, L. T., and Wirick, M. G. 1978. Bulk and solution properties of chitosan. In R. A. A. Muzzarelli, and E. R. Pariser (eds.), Proceeding of the 1st international conference on chitin and chitosan at MIT Sea Grant, pp. 169-181. USA.
- Flaherty, M., and Karnjanakesorn, C. 1995. Marine shrimp culture and natural resource degradation in Thailand. Environmental Management. 19(1): 27-37.
- Flaherty, M., and Vandergeest, P. 1997. 'Low salt' shrimp aquaculture in Thailand: Goodbye coastline hello Khon Kaen. Environmemtal management. 22(6): 817-830.
- Flegal, T. W., Fegan, D. F., and Sriurairatana, S. 1995. Environmental control of infectious shrimp diseases in Thailand. In M. Shariff, J. R. Arthur, and R. P. Subasinghe (eds.), Diseases in Asian Aquaculture II, pp. 65-79. Manila, Philippines: Fish Health Section, Asian Fisheries Society.
- Flegal, T. W., Fegan, D. F., Kongsom, S. et al. 1992. Occurrence, diagnosis and treatment of shrimp diseases in Thailand. In W. Fulks, and K. L. Main (eds.), Diseases of cultured penaeid shrimp in Asia and the United States, pp. 57-112. Hawaii, USA: The Oceanic Institute.
- Flegel, T. W. 1997. Special topic review: Major viral diseases of the black tiger prawn (*Penaeus monodon*) in Thailand. World Journal of Microbiology and Biotechnology. 13: 433-442.
- Flegel, T. W. 2001. The shrimp response to viral pathogens. In C. L. Browdy, and E. J. Darryl (eds.), The New Wave, Proceedings of the special session on sustainable

- shrimp culture, Aquaculture 2001, pp. 254-278. Baton Rouge, USA: The World Aquaculture Society.
- Flegel, T. W., Fegan, D. F., and Sriurairatana, S. 1995. Environmental control of infectious shrimp diseases in Thailand. In M. Shariff, J. R. Arthur, and R. P. Subasinghe (eds.), Diseases in Asian Aquaculture II, pp. 65-79. Philipines: Fish Health Section, Asian Fisheries Society.
- Folke, C., and Kautsky, K. 1992. Aquaculture with its environment: prospects for sustainability. Ocean and Coastal Management. 17(1): 5-24.
- Forster, J. R. M., and Gabbott, P. A. 1971. The assimilation of nutrients from compounded diets by prawns, *Palaemon serratus* and *Pandalus platyceros*. Journal of the Marine Biological Association of the United Kingdom. 51: 943-961.
- Fox, C. J. 1993. The effect of dietary chitin on the growth, survival and chitinase levels in the digestive gland of juvenile *Penaeus monodon* (Fab.). Aquaculture. 109: 39-49.
- Fujiki, K., and Yano, T. 1997. Effects of sodium alginate on the non-specific defense system of the common carp (*Cyprinus carpio* L.). Fish and Shellfish Immunology. 7: 417-427.
- Fujiki, K., Matsuyama, H., and Yano, T. 1994. Protective effect of sodium alginates against bacterial infection in common carp, *Cyprinus carpio* L. Journal of fish disesses. 17: 349-355.
- Funge-Smith, S. 1997. Thailand. World shrimp farming 1997. In B. Rosenberry (ed.), Shrimp News International, pp. 142-154. San Diego, USA: Aquaculture Digest.
- Furda, I. 1983. Aminopolysaccharide-their potential as dietary fiber. In I. Furda (ed.), Unconventional Sources of dietary fibers, pp. 105-122. Washington: American Chemical Society.
- Furuichi, M., and Yone, Y. 1982. Availability of carbohydrate in nutrition of carp and red sea bream. Bulletin of the Japanese Society of Scientific Fisheries. 48: 945-948.
- Gargioni, R., and Barracco, M. A. 1998. Hemocytes of the palaemonids *Macrobrachium rosenbergii* and *M. acanthurus*, and of the penaeid *Penaeus paulensis*. Journal of morphology. 236: 209-221.

- Gaxiola, G, Cuzon, G, García, T. et al. 2005. Factorial effects of salinity, dietary carbohydrate and moult cycle on digestive carbohydrases and hexokinases in *Litopenaeus vannamei* (Boone, 1931). Comparative Biochemistry and Physiology. 140(1): 29-39.
- Gerddit, W. 2002. Polysaccharide gel from dried fruit-hulls of durian as dressing-patch. Master's thesis. Department of Biochemistry, Faculty of Pharmaceutical sciences, Chulalongkorn University.
- Girddit, W., Tipayakul, C., Lertchaiporn, J. et al. 2001. Characterization and properties of polysaccharide gel from durian fruit-hulls. The Thai Journal of Pharmaceutical Sciences. 25(suppl.): 6.
- Goebbels, J. H. G. 1991. Residue problem with regard to cultivated fish and shellfish. In C. Michel, and D. J. Aldermann (eds.), Problems of chemotherapy in aquaculture from theory to reality, pp. 333-335. Paris: Office International DES EPI ZOO TIES.
- González, M. E., Alarcón, B., and Carrasco, L. 1987. Polysaccharides as antiviral agents: antiviral activity of carrageenan. Antimicrobial Agents and Chemotherapy. 31: 1388-1393.
- Gräslund, S., and Bengtsson, B. E. 2001. Chemical and biological products used in South-East Asian shrimp farming, and their potential impact on the environment. The Science of The Total Environment. 280(1-3): 93-131.
- Harada, K. 1987. Relationship between structure and feeding attraction activity of certain L-amino acids and lecithin in aquatic animals. Bulletin of the Japanese Society of Scientific Fisheries. 53: 2243-2247.
- Harris, L. J., and Owens, L. 1999. Production of exotoxins by two luminous *Vibrio harveyi* strains known to be primary pathogens of *Penaeus monodon* larvae. Diseases of Aquatic Organisms. 38: 11-22.
- Haywood, M., and Wells, S. 1989. The manual of marine invertebrates. London: Salamander Books.
- Hertrampf, J. W. 1985. Feeding phosphates and their biological value. 6th Australian. Australia.

- Hertrampf, J. W. 1991. Feeding aquatic animals with phospholipids. Crustaceans. Lucas Meyerr Publication.
- Hoffman-La Roche, F. 1987. Recommended vitamin supplementation levels for domestic animals. 2 nd revised ed. Basel, Switzerland.
- Hokputsa, S., Gerddit, W., Pongsamart, S. et al. 2004. Water-soluble polysaccharides with pharmaceutical importance from the rinds of durian (*Durio zibethinus* L.): isolation, fraction, characterization and bioactivity. Carbohydrate polymers. 56: 471-481.
- Holthuis, L.B. 1980. FAO species catalogue. Vol. 1. Shrimps and prawns of the world. An annotated catalogue of species of interest to fisheries. FAO Fisheries Synopsis. 125: 261.
- Hood, M. A., and Meyers, S. P. 1973. Microbial aspects of penaeid shrimp digestion. In J. B. Higman (ed.), Proceeding Gulf Caribbean Fish. Inst., 26 Annual Sess., October 1973, pp. 81-91. USA: University of Miami.
- Hopkins, J. S., Hamilton, R. D., and Sandifer, P. A. 1993. Effect of water exchange rate on production, water quality, effluent characteristics and nitrogen budget of intensive shrimp ponds. Journal of the World Aquaculture Society. 24: 304–320.
- Hopkins, J. S., Sandifer, P. A., and Browdy, C. L. 1995. A review of water management regimes which abate the environmental impact of shrimp farming. In C. L. Browdy, and J. S. Hopkins (eds.), Swimming through troubled water, pp. 13-22. Baton Rouge, LA: World Aquaculture Society.
- Hose, J. E. and Martin, G. G. 1989. Defense functions of granulocytes in the ridgeback prawn, *Sicyonia ingentis*. Journal of Invertebrate Pathology. 53: 335-346.
- Hose, J. E., Martin, G. G., and Gerard, A. S. 1990. A decapod hemocyte classification scheme integrating morphology, cytochemistry and function. The Biological bulletin. 178: 33-45.
- Huang, C. C., and Song, Y. L. 1999. Maternal transmission of immunity to white spot syndrome associated virus (WSSV) in shrimp (*Penaeus monodon*). Developmental and Comparative Immunology. 23: 545-552.

- Inouye, K., Miwa, S., Oseko, N. et al. 1994. Mass mortalities of cultured kuruma shrimp, *Penaeus japonicus* in Japan in 1993: electron microscopic evidence of the causative virus. *Fish pathology*. 29: 149–158.
- Ishihara, C., Yoshimatsu, K., Tsuji, M. et al. 1993. Anti-viral activity of sulfated chitin derivatives against friend murine leukaemia and herpes simplex type-1 viruses. *Vaccine*. 11(6): 670-674.
- Israngkura, A., and Sae-Hae, S. 2002. A review of the economic impacts of aquatic animal disease. In J. R. Arthur, M. J. Phillips, R. P. Subasinghe et al. (eds.), *Primary Aquatic Animal Health Care in Rural, Small-scale, Aquaculture Development*, pp. 253-286. FAO Fisheries Technical Paper.
- Itami, T., Asano, M., Tokushige, K. et al. 1998. Enhancement of disease resistance of kumura shrimp, *Penaeus japonicus*, after oral administration of peptidoglycan derived from *Bifidobacterium thermophilum*. *Aquaculture*. 164: 277-288.
- Itami, T., Takahashi, Y., Yoneoka, K. et al. 1991. Survival of larval giant tiger prawns *Penaeus monodon* after addition of killed Vibrio cells to a microencapsulated diet. *Journal of Aquatic Animal Health*. 3: 151-152.
- Jagodzinski, P. P., Wiaderkiewicz, R., Kurzawski, G. et al., 1994. Mechanism of the inhibitory effect of curdlan sulfate on HIV-1 infection *in Vitro*. *Virology*. 202(2): 735-745.
- Jarp, J., and Tverdal, A. 1997. Statistical aspect of fish vaccination trials. In R. Gudding, A. Lillehaug, P. J. Midtlyng, and F. Brown (eds.), *Fish vaccinology. Developments in biological standardization*, pp. 311-320. Basel, Switzerland: S. Karger.
- Jenkins, S., Smith, P. T., Tookwinas, S. 1999. An assesment of the status of the shrimp farming industry in Thailand. In S. T. Paul (ed.), *In Coastal shrimp aquaculture in Thailand: key issues for research*, pp. 14-68. Canberra, Australian: Arawang Communication Group.
- Jiravanichpaisal, P., Miyazaki, T., and Limsuwan, C. 1994. Histopathology, biochemistry and pathogenicity of *Vibrio harveyi* infecting black tiger prawn *Penaeus monodon*. *Journal of aquatic animal health*. 6:27-35.

- Johansson, M. W., and Söderhäll, K. 1988. Isolation and purification of a cell adhesion fraction from crayfish blood cells. Journal of Cell Biology. 106: 1795-1803.
- Johansson, M. W. 1999. Cell adhesion molecules in invertebrate immunity. Developmental and Comparative Immunology. 23: 303-315.
- Johansson, M. W., and Söderhäll, K. 1985. Exocytosis of the prophenoloxidase activating system from crayfish hemocytes. Journal of comparative physiology. 156: 175-181.
- Johansson, M. W., and Söderhäll, K. 1989. Cellular immunity in crustaceans and the proPO system. Parasitol Today. 5: 171-176.
- Johansson, M. W., and Söderhäll, K. 1992. Cellular defense and cell adhesion in crustaceans. Animal Biology. 1: 97-107.
- Johansson, M. W., Keyser, P., Sritunyalucksana, K. et al. 2000. Crustacean hemocytes and hematopoiesis. Aquaculture. 191: 45-52.
- Johnson, P. T. 1980. Histology of blue crab, *Callinectes sapidus*. A model for the Decapoda, p. 440. New York: Praeger.
- Johnson, S. K. 1989. Handbook of Shrimp Diseases, p. 27. USA: Texas A&M University.
- Jomori, T., and Natori, S. 1992. Function of the lipopolysaccharide-binding protein of *Periplaneta Americana* as an opsonin. FEBS letters. 296: 283-286.
- JWH. 1995 Using drugs in aquatic animals. Lecithin Trends 23/95. May (Lucas Meyer Newsletter).
- Kajiwara, K., and Miyamoto, T. 1998. Progress in structural characterization of functional polysaccharides. In S. Dumitriu, and M. Dekker (eds.), Polysaccharides: structural diversity and functional versatility, p.1. The United States of America.
- Kamazawa, A. 1995. Recent developments in shrimp nutrition and industry. Technplogy Session of INDAQUA 1995. 27-30 January 1995. Madras, India.
- Kanazawa, A. 1982. Penaid nutrition. Proceedings. 2nd Int' 1 Conference on Aquaculture Nutrition, Biochemistry and Physiological Approaches to Shellfish Nutrition, pp.87-105. 27-29 October 1982. Rehoboth Beach, Delaware, USA.

- Kanazawa, A. 1984. Nutrition of penaeid prawns and shrimps In Y. Taki, J. H. Primavera, and J. A. Llobrera (eds.), Proceeding 1st Int. Conference Culture penaeid prawns/shrimp, pp. 123-130. Philippines: Aquaculture Deparment, Southeas Asian Fisheries Development.
- Kanazawa, A. Teshima, S., and ssaki, M. 1984. Requirement of the juvenile prawn for calcium, phosphorus, magnesium, potassium, copper, manganese and iron. Memoirs of the Faculty of Fisheries Kagoshima University. 33:63-71.
- Kanazawa, A., Shimaya, M., Kawashai, M. et al. 1970. Nutritional requirements of prawn. I. Feeding on artificial diet. Bulletin of the Japanese Socirty of Scientific Fisheries. 36: 949-954.
- Kanchanaphum, P., Wongteerasupaya, C., Sitidilokratana, N. et al. 1998. Experimental transmission of white spot syndrome virus (WSSV) from crabs to shrimp *Penaeus monodon*. Diseases of Aquatic Organisms. 34: 1-7.
- Kanost, M. R. 1999. Serine proteinase inhibitors in arthropod immunity. Developmental and Comparative Immunology. 23: 291-301.
- Kasornchandra, J., Boonyaratpalin, S., Aekpatithanpong, U. et al. 1995. Mass mortality caused by systemic bacilliform virus in cultured penaeid shrimp, *Penaeus monodon*, in Thailand. Asian Shrimp News. 5(2): 2-3.
- Katesombun, B. 1992. Aquaculture promotion: endangering the mangrove forests. The future of pepple and forests after the logging ban, pp. 103-122. Bangkok: Project for ecological recovery.
- Kiriratnikom, S., Ruangsri, J., Wanadet, M. et al. 2000. The abiotic factors influencing the growth of luminescent bacteria, in seawater. Journal of science and technology. 22(Suppl.): 697-705.
- Kitabayashi, K., Kurata, H., Shudo, K. et al. 1971. Studies on formula feed for Kuruma prawn. I. On the relationship among glucosamine, phosphorus and calcium. Bulletin of Tokai Regional Fisheries Research Laboratory. 65: 91-107.
- Kitabayashi, K., Shudo, K., Nakamura, K. et al. 1971. Studies on formula feed for Kuruma prawn. II. On the utilization values of glucose. Bulletin of Tokai Regional Fisheries Research Laboratory. 65: 109-118.

- Kobayashi, M., Johansson, M. W, and Söderhäll, K. 1990. The 76kD cell adhesion factor from crayfish hemocytes promotes encapsulation *in vitro*. Cell and Tissue Research. 260: 13-18.
- Kooiman, P. 1964. Occurrences of carbohydراse in digestive juice in hepatopancreas of *Astacus fluviatilis* and *Homarus vulgaris*. Journal of Cellular Comparative Physiology. 63: 197-201.
- Kopácek, P., Grubhoffer, L., and Söderhäll. K. 1993. Isolation and characterization of a hemagglutinin with affinity for lipopolysaccharides from plasma of the crayfish *Pacifastacus leniusculus*. Developmental and Comparative Immunology. 23: 291-301.
- Kurmaly, K. 1994. Balancing out nutrition. Intern.Milling Flour and Feed. 188(2): 75-80.
- Lackie, A. M. 1980. Invertebrate immunity. Parasitology. 80: 393-412.
- Lall, S. P. 1989. The minerals. In J. E. Halver (ed.), Fish nutrition. 2 nd ed. London: Academic Press.
- Lasson, J., Folke, C., and Kautsky, N. 1994. Ecological limitations and appropriation of ecosystem support by shrimp farming in Columbia. Environmental Management. 18(5): 663-676.
- Lavilla-Pitogo, C. R. 1995. Bacterial diseases of penaeid shrimp: an Asia view. In M. Shariff, J. R. Arthur, and R. P. Subasinghe (eds.), Diseases in Asia Aquaculture II, pp. 107-121. Manila: Fish Health Section, Asian Fisheries Society.
- Lavilla-Pitogo, C. R., Baticados, M. C. L., Cruz-Lacierda et al. 1990. Occurrence of luminous bacterial disease of *Penaeus monodon* larvae in the Philippines. Aquaculture. 91: 1-13.
- Le Moullac, G., Le Groumellec M., Ansquer D. et al. 1997. Haematological and phenoloxidase activity changes in the shrimp *Penaeus stylirostris* in relation with the moult cycle: protection against vibriosis. Fish and Shellfish Immunology. 7: 227-234.
- Le, T. X., and Munekage, Y. 2004. Residues of selected antibiotics in water and mud from shrimp ponds in mangrove areas in Viet Nam. Marine pollution Bulletin. 49(11-12): 922-929.

- Le, T. X., Munekage, Y., and Kato, S. I. 2005. Antibiotic resistance in bacteria from shrimp farming in mangrove areas. In J. O. Nriagu (ed.), Science of the Total Environment, USA: ELSEVIER Commenced publication.
- Lee, D. O. C., and Wickins, J. F. 1992. Crustacean farming, p. 392. Cambridge: The University Press.
- Lee, M. H., and Shiau, S. Y. 2002. Dietary vitamin C and its derivatives effect immune responses in grass shrimp *Penaeus monodon*. Fish and Shellfish Immunology. 12: 119-129.
- Lee, M. H., and Shiau, S. Y. 2004. Vitamin E requirements of juvenile grass shrimp *Penaeus monodon* and effects on non-specific immune responses *Penaeus monodon*. Fish and Shellfish Immunology. 16: 475-485.
- Lerchaiporn, J., Vayumhasawan, P., and Pongsamart, S. 2003. Polysaccharide gel from fruit-hulls of durian as a mouth refreshing film. The 29th Congress on Science and Technology of Thailand, p. 43. 20-22 October 2003. Golden Jubilee Convention, Khon Kean University, Khon Kean, Thailand.
- Li, H. X., Meng, X. L., and Xu, J. P. 2005. Protection of crayfish, *Cambarus clarkii*, from white spot syndrome virus by polyclonal antibodies against a viral envelope fusion protein. Journal of fish diseases. 28(5):285-91.
- Liao, I. C. 1990. The world's marine prawn culture industries: today and tomorrow, pp. 11-17. Manila, Philippines: Asian Fisheries Society.
- Liao, I. C., Lee, W. C., and Hsu, Y. K. 1995. Aquaculture in Taiwan: Towards a sustainable industry. International cooperation for fisheries and aquaculture development. Proceedings of the 7th Biennial Conference of the international institute of fisheries economics and trade, pp. 1-13, 18-21 July 1994. Taipei, Taiwan. National Taiwan Ocean University.
- Liao, I. C., Su, M. S., Chang, C. F. et al. 1996. Enhancement of the resistance of grass prawn *Penaeus monodon* against *Vibrio damsela* infection by beta-1,3 glucan. Journal of the Fisheries Society of Taiwan. 23: 109-116.
- Lightner, D. V. 1983. Disease of cultured penaeid shrimp. In J. P. McVey. (ed.), Handbook of mariculture. Volume I. Crustacean Aquaculture, pp. 289-377. Baton Rouge: CRC Press.

- Lightner, D. V. 1996. A handbook of shrimp pathology and diagnostic procedures for diseases of cultured penaeid shrimp. Section 3: Viruses. Baton Rouge, Louisiana, USA: The World Aquaculture Society.
- Lightner, D. V., and Redman, R. M. 1998. Shrimp diseases and current diagnostic methods. Aquaculture. 164: 201-220.
- Lightner, D. V., Beell, T. A., Redman, R. M. et al. 1992. A review of some major diseases of economic significance in penaeid prawns/shrimp of the Americas and Indopacific. In M. Shariff, R. P. Subasinghe, and J. R. (eds.), pp. 57-80, Diseases in Asian Aquaculture I. Manila: Fish Health Section, Asian Fisheries Society.
- Lim, C., and Persey, A. 1988. Practical feeding-penaeid shrimps. Lovell, R.T. (ed.), Nutrition and feeding of fish. New York, USA: Van Nostrand Reinhold.
- Lin, C. K. 1995. Progression of Intensive marine shrimp culture in Thailand. In C. Browdy, and J. S. Hopkins (eds.), Swimming through troubled water, pp. 13-22. Baton Rouge, LA: World Aquaculture Society.
- Lo, C. F, Ho, C. H., Peng, S. E. et al. 1996. White spot syndrome baculovirus (WSBV) detected in cultured and captured shrimp, crabs and other arthropods. Diseases of Aquatic Organisms. 27: 215-225.
- Lorian, V. 1991. Antibiotics in laboratory medicine. 3 rd ed., pp.739-786. London: Williams & Wilkins.
- Maeda, M., Itami, T., Furumoto, A. et al. 1998. Detection of rod-shaped DNA virus (PRDV) in wild-caught shrimp and other crustaceans. Fish pathology. 33: 373-380.
- Maeda, M., Kasornchandra, J., Itami, T. et al. 1998. Effect of various treatments on white spot syndrome virus (WSSV) from *Penaeus japonicus* (Japan) and *Penaeus monodon* (Thailand). Fish Pathology. 33: 381-387.
- Maktrirat, R., Ajariyakhajorn, K., and Pongsamart, S. 2004. *In vitro* antibacterial activity of polysaccharide gel from durian fruit hulls against bacteria isolated from dairy cow mastitis. The Thai Journal of Pharmaceutical Sciences. 28 (Suppl.): 22.
- Marques, M. R. F., and Barracco, M. A. 2000. Lectins, as non-self recognition factors, in crustaceans. Aquaculture. 191: 23-44.

- Martin, G. G., and Graves, B. L. 1985. Fine structure and classification of shrimp hemocytes. Journal of Morphology. 185: 339-348.
- Martin, G. G., and Hose, J. E. 1992. Vascular elements and blood (hemolymph). In F. W. Harrison, and A. G. Humes (eds.), Microscopic Anatomy of Invertebrates, pp. 117-146. New York, USA: Wiley-Liss.
- Masae, A., and Rakkheaw, S. 1992. Social aspects of artisanal fisheries and shrimp farming in Pak Phanang Bay. p. 74. Asian Fisheries Social Science Research Network: Coastal Resources Institute, Prince of Songkhla University, Hat Yai, Thailand.
- Mastromario, A., Petruzzello, R., Macchia, S. et al. 1997. Antiviral activity of natural and semisynthetic polysaccharides on the early steps of rubella virus infection. Journal of Antimicrobial Chemotherapy. 39: 339-345.
- Matsuyama, H., Remy, E. P., Mangindaan, T. et al. 1992. Protective effect of schizophyllan and scleroglucan against *Streptococcus sp.* infection in yellowtail (*Seriola quinqueradiata*). Aquaculture. 101(3-4): 197-203.
- Mc Clure, M. O., Moore, P. J., Blanc, D. F. et al. 1992. Investigations into the mechanism by which sulfated polysaccharides inhibit HIV infection in *vitro*. AIDS Research and Human Retroviruses. 8: 19-26.
- McKay, D., and Jenkin, C. R. 1970. Immunity in the invertebrates. The role of serum factors in phagocytosis of erythrocytes by hemocytes of freshwater crayfish (*Parachaeraps bicarinatus*). The Australian journal of experimental biology and medical science. 48: 139-150.
- McKee, T., and McKee, J. R. 1999. Biochemistry: an introduction. 2 nd ed., p.151. The United States of America: The McGraw-Hill.
- Mearns, K. J. 1985. Response of Atlantic salmon (*Salmo gairdneri* L.) yearlings to individual L-amino acids. Aquaculture. 48:253.
- Menasveta, P. 2002. Improved Shrimp Grow out Systems for Disease Prevention and Environmental Sustainability in Asia. Reviews in Fisheries Science. 10(3-4): 391-402.
- Meyer, F. 1991. Aquaculture disease and health management. Journal of Animal Science. 69: 4201-4208.

- Meyers, S. P. 1986. Attractants, aquatic diet development. Feedstuffs. 58(29): 12.
- Mialhe, E., Bachè, E., and Boulo, V. 1995. Strategy for research and international cooperation in marine invertebrate pathology, immunology and genetics. Aquaculture. 132: 33-41.
- Miao, B., Geng, M., Li, J. et al., 2004. Sulfated polymannuroguluronate, a novel anti-acquired immune deficiency syndrome (AIDS) drug candidate, targeting CD4 in lymphocytes. Biochemical Pharmacology. 68(4): 641-649.
- Miettinen, O. S. 1947. Proportion of disease caused or prevented by a given exposure, trait or prevention. American journal of epidemiology. 99: 325-332.
- Millamena, O. M., Pudadera, R. A., and Catacutan, M. R. 1984. Effects of diet on reproductive performance of ablated *Penaeus monodon* broodstock. Proceedings 1st International Conference on Culture of Penaeid prawns/shrimps, p.178. Iloilo City, Philippines.
- Millar, D. A., and Ratcliffe, N. A. 1994. Invertebrates. In R. J. Turner (ed.), pp. 29-68, Immunology, a comparative approach. England: John Wiley and Sons.
- Mohamed, K. H. 1970. Synopsis of the biological data on the jumbo tiger prawn *Penaeus monodon* Fabricius, 1798. FAO Fisheries Report. 57(4): 1251–1266.
- Motoh, H. 1981. Studies on the fisheries biology of the giant tiger prawn, *Penaeus monodon* in the Philippines. Technical Report, No. 7, p. 128. Tigbauan, Iloilo: Aquaculture Department, Southeas Asian Fisheries Development.
- Murai, T., Akiyama, T., and Nose, T. 1983. Effects of glucose chain length of various carbohydrates and frequency of feeding on their utilization by fingerling carp. Bulletin of the Japanese Society of Scientific Fisheries. 49(10): 1607-1611.
- Mushiake, K., Shimizu, K., Satoh, J. et al., 1999. Control of penaeid acute viremia (PAV) in *Penaeus japonicus*: selection of eggs based on the PCR detection of the causative virus (PRDV) from receptaculum seminis of spawned brood stock. Fish pathology. 34: 203-207.
- NACA (Network of Aquaculture Centres in Asia-Pacific). 1994. Impact of shrimp farming on the environment: study I. Sixth Meeting of the Governing Council (GCM-6) of NACA, p. 195. Beijing, China

- Nakchat, O., Nantawan, N., Lipipan, V. et al. 2003. Antibacterial polysaccharide gel of durian as a water soluble dressing for healing wounds in the skin of pigs (*in vivo*). Asian symposium on medicinal plants, spices and other natural products XI (ASOMPS XI), p. 319. 26-30 October 2003. Kunming, China.
- Namikoshi, A., Wu, J. L., Yamashita, T. et al. 2004. Vaccination trials with *Penaeus japonicus* to induce resistance to white spot syndrome virus. Aquaculture. 229: 25-35.
- Nantawanit, N. 2001. Antimicrobial property of polysaccharide gel from durian fruit-hulls. Master's thesis. Department of Biochemistry, Faculty of Pharmaceutical sciences, Chulalongkorn University.
- Nash, G. C., Nithimathachoke, C., Tungmandi, A. et al. 1992. Vibriosis an dits control in pond-reared *Penaeus monodon* in Thailand. In I. M. Shariff, R. P. Subasinghe, and J. R. Arthur (eds.), Diseases in Asian Aquaculture, pp. 143-155. Manila: Fish Health Section, Asian Fisheries Society.
- Neiland, A., Soley, N., and Baron, J. 1997. A review of the Literature on Shrimp culture. Paper presented at the Bangkok FAO technical consultation on policies for sustainable shrimp culture. 8-11 December 1997. Bangkok, Thailand. 572(suppl./supl.): 209-229.
- Neu, T. R., Dengler, T., Jann, B. et al. 1992. Structural studies of an emulsion-stabilizing exopolysaccharides produced by an adhesive, hydrophobic *Rhodococcus* strain. Journal of General Microbiology. 138: 2531-2537.
- New, M. B. 1976. A review of dietary studies with shrimps and prawns. Aquaculture. 9: 101-144.
- New, M. B. 1987. Feed and feeding of fish and shrimps. A manual on the preparation and presentation of compound feeds for shrimps and fish in aquaculture. UNDP/FAO/ADCP/REP/87/26. Rome, Italy.
- New, M. W. 1980. A bibliography of shrimp and prawn nutrition. Aquaculture. 21: 101-128.
- Niemeier, P. E. 1990. Thailand's shrimp culture growing. Marine Fisheries Review. 52(2): 21-25.
- Nørhede, J. S. 1991. Feed quality and aroma of feed. Feed Magazine. 1: 14-18.

- NRC. 1981. Nutrient requirements of coldwater fishes. Washington, D. C., USA: National Academy Press.
- Nuruzzaman, A. K. M. 1996. Socio-environmental impact of expansion of shrimp culture: Lessons from Bangladesh. Proceedings of the 1996 Annual Meeting of the World Aquaculture Society, p. 287. Bangkok, Thailand.
- Omori, S. A., Martin, G. G., and Hose, J.E. 1989. Morphology of hemocyte lysis and clotting in the ridgeback prawn *Sicyonia ingentis*. Cell and tissue research. 255: 117-123.
- Osawa, Z., Morota, T. Hatanaka, K. et al. 1993. Synthesis of sulfate derivatives of curdlan and their anti-HIV activity. Carbohydrate Polymer. 21: 283-288.
- Pandian, T. J. 1989. Protein requirements of fish and prawns cultured in Asia. Proceedings Third Asian Fish Nutrition Network Meeting, pp. 11-22. Manila, Philipines: Asian Fish. Soc. Spec
- Paphattarapong, N., Lipipun, V., and Pongsamart, S. 2004 a. Polysaccharide gel from durian with betel oil in antimicrobial lotion. The Thai Journal of Pharmaceutical Sciences. 28 (Suppl.): 22.
- Paphattarapong, N., Pongwiwatana, U., Lipipun, V. 2004 b. Antimicrobial activity (*in vivo*) of essential oil useable in gel preparation of antibacterial polysaccharide from durian fruit-hull. The 30th Congress on Science and Technology of Thailand, pp. 61-62. 19-21 October 2004. Impact Exhibition and Convention Center, Muang Thong Thani, Bangkok.
- Parish, C. R., Low, L., Warren, H. S. et al. 1990. A polyanion binding site on the CD4 molecule. Proximity to the HIV-gp 120 binding region. Journal of Immunology. 145: 1188-1198.
- Pascual, F. P., Coloso, R. M., and Tamse, C. T. 1983. Survival and some histological change in *P. monodon* Fabricius juveniles fed various carbohydrates. Aquaculture. 31: 169-180.
- Paulraj, R. 1995. Aquaculture. Marine Production Export Development. 2 nd ed. Kochi, India: Publisher.

- Peñaflorida, V. D., and Golez, N. V. 1996. Use of seaweed meals from *Kappaphycus alvarezii* and *Gracilaria heteroclada* as binders in diets for juvenile shrimp *Penaeus monodon*. Aquaculture. 143(3-4): 393-401.
- Petriella, A. M., Muller, M. I. Fenucci, J. C. et al. 1984. Influence of dietary fatty acids and cholesterol on the growth and survival of the Argentine prawns, *Artemia longinares* Bate. Aquaculture. 37: 11-20.
- Piedad, P. F. 1989. Mineral requirement of penaeids. Aquacop Actes de Colloque. 9: 309-318.
- Piefer, A., and Pfeffer, E. 1980. Studies on the comparative efficiency of utilization of gross energy from som carbohydrates, proteins and fats by rainbow trout (*Salmo gairdneri*). Aquaculture. 20: 323-332.
- Pongsamart, S. 1989 b. The studies of carbohydrate extracts from durian rinds to use as suspending agent. Research report. Department of Biochemistry, Faculty of Pharmaceutical Sciences, Chulalongkorn University.
- Pongsamart, S., and Panmaung, T. 1998. Isolation of polysaccharides from fruit-hulls of durian (*Durio zibethinus* L.). Songklanakarin Journal Science and Technology. 20(3): 323-332.
- Pongsamart, S., Dhumma-Upakorn, R., and Panmaung, T. 1989 a. The studies of carbohydrate from durian rind for pharmaceutical and food preparations. Research Report. Rachadapiseksompoach Research Funds, Chulalongkorn University.
- Pongsamart, S., Jesadanont, S.N., and Markman, N. 1989. The studies on safety a toxicity of the consumption of pectin-like substance isolated from durian rinds. Reserch Report. Faculty of Pharmaceutical Sciences, Chulalongkorn University.
- Pongsamart, S., Lipipun, V., Nantawanit., N. et al. 2003. Novel water soluble antibacterial dressing of durian polysaccharide gel. The 3rd World Congress on Medicinal and Aromatic Plants for Human Welfare (WOCMAP III), p. 104. 3-7 February 2003. Chiang Mai University, Chiang Mai. Thailand.
- Pongsamart, S., Sukrong, S., and Tawatsin, A. 2001 a. The determination of toxic effects at a high oral does of polysaccharide gel extracts from fruit-hulls of durian (*Durio zibethinus* L.) in mice and rats. Songklanakarin Journal Science and Technology. 23(1): 56-62.

- Pongsamart, S., Tawatsin, A., and Sukrong, S. 2001 b. Long-term consumption of polysaccharide gel from durian fruit-hull in mice. Faculty of Pharmaceutical Sciences, Chulalongkorn University (in press).
- Pongwiwatana, U., Lipipun, V. Jesadanont, S. et al. 2004. Polysaccharide gel from durian with tea tree oil in antiseptic gel preparation. The Thai Journal of Pharmaceutical Sciences. 28 (Suppl.): 22.
- Pongwiwatana, U., Paphattarapong, N., Keddit, W. et al. 2003. *In vitro* suscepthipility test of durian polysaccharide gel (PG) against isolates of *Staphylococcus aureus* from dairy cow mastitis. The 29th Congress on Science and Technology of Thailand, p. 76. 20-22 October 2003. Golden Jubilee Convention, Khon Kean University, Khon Kean, Thailand.
- Primavera, J. H. 1992. Prawn/shrimp culture in the Philippines. In A. W. Fast, and L. J. Lester (eds.), Marine Shrimp Culture: Principles and Practices, pp. 701-728. Amsterdam: Elsevier.
- Primavera, J. H. 1998. Tropical shrimp farming and its sustainability. In S. S. De Silva (ed.), Tropical Mariculture, pp. 257-289, San Diego: Academic Press.
- Primavera, J. H., 1993. A critical review of shrimp pond culture in the Philippines. Fisheries Science. 1(2): 151–201.
- Rao, P. S., and Parekh, K. S. 1981. Antibacterial activity of Indian seaweed extracts. Botanica Marina. 25: 577-582.
- Rasheed, A. A. 1963. Nutritive value of marine oil. I. Menhaden oil at varying oxidation levels with and without antioxidants in rat diets. The Journal of Nutrition. 79: 323-332.
- Ratcliffe, N. A., Rowley, A. F., Fitzgerald, S. W. et al. 1985. Invertebrate immunity: basic concepts and recent advances. International review of cytology. 97: 183-350.
- Regelson, W. 1968. Physiological effects of polyanionic macromolecules. Advances in Chemotherapy. 3: 303.
- Rosenberry, B. 1997. World Shrimp Farming 1997. Shrimp News International, p. 284. San Diego.

- Rosenberry, B. 2001. World Shrimp Farming 2001. Shrimp News International, p. 324. San Diego.
- Ruangpan, L., and Kitao, T. 1991. *Vibrio* bacteria isolated from black tiger shrimp, *Penaeus monodon* Fabricious. Journal of fish diseases. 14: 383-388.
- Ruby, E. G., and Nealson, K. H. 1978. Seasonal changes in the species composition of luminous bacteria in near shore seawater. Limnology and Oceanography. 23: 530-533.
- Rukprataporn, S. 1999. Immunoenhancement in black tiger shrimp Penaeus monodon by Bacillus strain S11. Master thesis. Department of Microbiology, Faculty of Pharmaceutical sciences, Chulalongkorn University.
- Schols, D., De Clercq, E., Balzarini, J. et al. 1990. Sulfated polymers are potent and selective inhibitors of various enveloped viruses, including herpes simplex virus, cytomegalovirus, vesicular stomatitis virus, respiratory syncytial virus, and togavirus- and retroviruses. Antiviral Chemistry and Chemotherapy. 1: 233-240.
- Sermwatanakul, A. 1994. Antibiotic Drugs Residue in Shrimp. Thai Fisheries Gazette. 47(6): 517-522.
- Shang, Y. C., Leung, P., and Ling, B. H. 1998. Comparative economics of shrimp farming in Asia. Aquaculture. 164: 183-200.
- Shiau, S. Y., and Chou B.S. 1991. Effect of dietary protein and energy on growth performance of tiger shrimp *Penaeus monodon* reared in seawater. Nippon Suisan Gakkaishi. 57(12): 2271-2276.
- Shiau, S. Y., and Peng, C. Y. 1992. Utilization of different carbohydrates at different dietary protein levels in grass prawn, *Penaeus monodon*, reared in seawater. Aquaculture. 101: 241-250.
- Shiau, S. Y., and Yu, Y. P. 1998. Chitin but not chitosan supplementation enhances growth of grass shrimp, *Penaeus monodon*. The Journal of Nutrition. 128: 908-912.
- Shiau, S. Y., Kwok, C. C., and Chou, B. S. 1991. Optimal dietary protein level of *Penaeus monodon* reared in seawater and brackish water. Nippon Suisan Gakkaishi. 57: 711-716.

- Shilo, M., and Yetinson, T. 1979. Physiological characteristics underlying the distribution patterns of luminous bacteria in the Mediterranean Sea and the Gulf of Elat. Applied and Environmental Microbiology. 38: 577-584.
- Shinbut, S. 2002. Antibiotic residues in shrimp. Thai Fisheries Gazette. 55(3).
- Sick, L. V., and Andrews, J. W. 1973. The effect of dietary lipids, carbohydrates and proteins on the growth, survival and body composition of *Penaeus duorarum*. Proceedings of the World Mariculture Society. 4: 263-276.
- Siegmund, O. H., Mclean, J. W., Armistead, W. W. et al. 1961. The Merck veterinary manual. 2 nd ed. USA: Merck and Co.
- Siripokasupkul, R., Chansiripornchai, P., Pramatwinai, C. et al. 2004 a. The wound healing efficacy of durian polysaccharide gel (PG) dressing film in dog skin. The 30th Congress on Science and Technology of Thailand, p 61. 19-21 October 2004. Impact Exhibition and Convention Center, Muang Thong Thani, Bangkok, Thailand.
- Siripokasupkul, R., Chansiripornchai, P., Pramatwinai, C. et al. 2004 b. Evaluation the effect of durian polysaccharide gel (PG) dressing preparations on wound healing in dog skin (*in vivo*). The Thai Journal of Pharmaceutical Sciences. 28 (Suppl.): 29.
- Smith, V. J., and Chisholm, J. R. S. 1992. Non-cellular immunity in crustaceans. Fish and Shellfish Immunology. 2: 1-31.
- Smith, V. J., and Söderhäll, K. 1983. β -1,3-glucan activation of crustacean hemocytes in vitro and in vivo. The Biological bulletin. 164: 299-314.
- Smith, V., and Chisholm, J. R. S. 1992. Non-cellular immunity in crustaceans. Fish and Shellfish Immunology. 2: 1-31.
- Söderhäll, K. 1999. Invertebrate immunity. Developmental and Comparative Immunology. 23: 263-266.
- Söderhäll, K., and Cerenius, L. 1998. Role of the prophenoloxidase-activating system in invertebrate immunity. Current Opinion in Immunology. 10: 23-28.
- Söderhäll, K., and Cerenius, L. 1992. Crustacean immunity. Annual Review of Fish Diseases. 2: 3-23.

- Söderhäll, K., and Cerenius, L. 1998. Role of the prophenoloxidase-activating system in invertebrate. Immunology Reviews. 198: 116-126.
- Söderhäll, K., and Cerenius, L. 2004. The prophenoloxidase-activating system in invertebrate immunity. Current Opinion in Immunology. 10: 23-28.
- Söderhäll, K., and Häll, L. 1984. Lipopolysaccharide induced activation of prophenoloxidase activating system in crayfish hemocyte lysate. Biochimica et biophysica acta. 797: 99-104.
- Söderhäll, K., and Smith, V. J. 1983. Separation of hemocyte populations of *Carcinus maenus* and other marine decapods and phenoloxidase distribution. Developmental and Comparative Immunology. 7: 229-239.
- Söderhäll, K., and Thornqvist, P. O. 1997. Crustacean immunity-A short review. In R. Gudding, A. Lillehaug, P. J. Midtlyng, and F. Brown (eds.), Fish vaccinology. Developments in biological standardization, pp. 45-51. Basel, Switzerland: S. Karger.
- Söderhäll, K., and Unestam, T. 1979. Activation of serum prophenoloxidase in arthropod immunity. The specificity of cell wall glucan activation and activation by purified fungal glycoproteins of crayfish phenoloxidase. Canadian Journal of Microbiology. 25: 406-414.
- Söderhäll, K., Cerenius, L., and Johansson, M. W. 1996. The prophenoloxidase activating system in invertebrates. Söderhäll, K., In Iwanaga, S. and Vasta, G. (eds.), New Directions in Invertebrate Immunology, pp. 229-254. Fairhaven, USA: SOS.
- Solis, N. B. 1988. Biology and ecology. Biology and culture of *Penaeus monodon*. Brackish water aquaculture information system, pp. 3-15. Tigbauan, Iloilo, Philippines: Aquaculture Department, Southeas Asian Fisheries Development.
- Somsiri, T. 1995. Antibiotic resistance in Thailand. Aquatic Animal Health Research Institute Newsletter Article. 4(1).
- Song, Y. L., and Hsieh, Y. T. 1994. Immunostimulation of tiger shrimp (*Penaeus monodon*) hemocytes for generation of microcidal substances: analysis of reactive oxygen species. Developmental and Comparative Immunology. 18: 201-209.

- Song, Y. L., and Sung, H. H. 1990. Enhancement of growth in tiger shrimp (*Penaeus monodon*) by bacterin prepared from *Vibrio vulnificus*. Bulletin of the European Association of Fish Pathologists. 10(4): 98.
- Song, Y. L., Liu, J. J., Chan, L. C. et al. 1997. Glucan-induced disease resistance in tiger shrimp (*Penaeus monodon*). Developments in Biological Standardization. 90: 423-42.
- Song, Y. L., Liu, J. J., Chan, L. C. et al. 1997. Glucan-induced disease resistance in tiger shrimp (*Penaeus monodon*). In R. Gudding, A. Lillehaug, P. J. Midtlyng (eds.), Fish vaccinology developments in biological standardization, pp. 413-421. Norway: Karger.
- Sritunyalucksana, K., and Söderhäll, K. 2000. The proPO and clotting system in crustaceans. Aquaaculture. 191: 53-69.
- Sritunyalucksana, K., Lee, S. Y., and Söderhäll, K. 2002. A β -1,3-glucan binding protein from the black tiger shrimp, *Penaeus monodon*, Developmental and Comparative Immunology. 26: 237-245.
- Sritunyalucksana, K., Sithisarn, P., Withayachummarkul, B. et al. 1999. Activation of prophenoloxidase, agglutinin and antibacterial activity in hemolymph of the black tiger prawn, *Penaeus monodon*, by immunostimulants. Fish and Shellfish Immunology. 9: 21-30.
- Sritunyalucksana, K., Wongsuebsantati, K., Johansson, M. W. et al. 2001. Peroxinectin, a cell adhesion protein associated with the proPO system from the black tiger shrimp shrimp, *Penaeus monodon*. Developmental and Comparative Immunology. 23: 179-186.
- Stewart, J. E., and Zwicker, B. M. 1972. Natural and induced bactericidal activities in the hemolymph of the lobster, *Homarus americanus*: products of hemocyte plasma interaction. Canadian journal of microbiology. 18: 1499-1508.
- Sugawara, I., Itoh, W., Kimura, S. et al. 1989. Further characterization of sulfated homopolysaccharide as anti-HIV agent. Experientia. 45: 996-998.
- Sung, H. H., Kou, G. H., and Song, Y. L. 1994. Vibriosis resistance induced by glucan treatment in tiger shrimp (*Penaeus monodon*). Fish and Shellfish Immunology. 29: 11-7.

- Sung, H. H., Yang, Y. L., and Song, Y. L. 1996. Enhancement of microbiocidal activity in the tiger shrimp *Penaeus monodon* via immunostimulation. *Journal of Crustacean Biology*. 16(2): 278-284.
- Supamattaya, K., Kiattapew, S., and Hoffmann, R.W. 2000. The immune system in black tiger shrimp *Penaeus monodon* (Fabricius) III: Electron microscopic studies on hemocytes of black tiger shrimp *Penaeus monodon* (Fabricius). *Journal of science and technology*. 22(suppl.):589-596.
- Tacon, A. G. J. 1987. The nutrition and feeding of farmed fish and shrimp. *Nutrient sources and composition*. FAO Field Document 5/E. Brasilia, Brazil.
- Takahashi, Y. Kondo, M., Itami, T. et al. 2000. Enhancement of disease resistance against penaeid acute viraemia and induction of virus-inactivating activity in hemolymph of kuruma shrimp, *Penaeus japonicus*, by oral administratioo of *Pantoea agglomerans* lipopolysaccharide (LPS). *Fish and Shellfish Immunology*. 10: 555-558.
- Teshima, S., and Kannazawa, A. 1986. Nutritive value of sterols for the juvenile prawn. *Bulletin of the Japanese Society of Scientific Fisheries*. 52(3): 519-524.
- Tookwinas, S. 1993. Intensive marine shrimp farming techniques in Thailand. *Proceedings of the first international symposium on aquaculture technology and investment opportunities*, pp. 230-240. 11-14 April 1993. Ministry of Agriculture and Water, King Abdulaziz City for Science and Technology and Riyadh Chamber and Industry. Riyadh, Saudi Arabia.
- Tsai, G. J., and Su, W. H. 1999. Antibacterial activity of shrimp chitosan against *Escherichia coli*. *Journal of Food Protection*. 62(3): 239-243.
- Tsai, M. F., Yu, H. T., Tzeng, H. F. et al. 2000 Identification and characterization of a shrimp White spot syndrome virus (WSSV) gene that encodes a novel chimeric polypeptide of cellular-type thymidine kinase and thymidylate kinase, *Virology* 277: 100–110.
- Tsing, A., Arcier, J. M., and Brehèlin, M. 1989. Hemocytes of penaeids and palaemonid shrimp: morphology, cytochemistry and hemograms. *Journal of Invertebrate Pathology*. 53: 64-77.

- Tyagi, A. P., and Prakash, A. 1967. A study on the physiology of digestion in freshwater prawn *Macrobrachium dayanum*. Journal of Zoology Society India. 19: 77-83.
- Uglea, C. V., and Ottenbrite, R. M. 1996. Polysaccharides as supports for antiviral and antitumoral drugs. In S. Dumitriu (ed.), Polysaccharides in medicinal application, pp. 765-779. New York: Marcel Dekker.
- Umprayn, K., Chanpaparp, K., and Pongsamart, S. 1990 b. The studies of durian rind extract as an aqueous binder I: Evaluation of granule properties. The Thai Journal of Pharmaceutical Sciences. 15(2): 95-115.
- Umprayn, K., Chanpaparp, K., and Pongsamart, S. 1990 c. The studies of durian rind extracts as an aqueous binder II: Evaluation of tablets properties. The Thai Journal of Pharmaceutical Sciences. 15(3): 173-186.
- Umprayn, K., Kaimonkong, R., and Pongsamart, S. 1990 a. Evaluation of tablet disintegrating properties of durian rind extracts. NUS-JSPS Seminar. 23-26 October 1990. Chiba, Japan.
- Van de Braak, C. B. T. 2002. Haemocytic defense in black tiger shrimp (*Peneaus monodon*). PhD. Thesis. Wageningen University.
- Van de Braak, C. B. T., Botterblom, M. H. A., Taverne, N. et al. 2002. The roles of hemocytes and the lymphoid organ in the clearance of injected *Vibrio* bacteria in *Penaeus monodon* shrimp. Fish and Shellfish Immunology. 13: 293-309.
- Van de Braak, C. B. T., Faber, R., and Boon, J. H. 1996. Cellular and humoral characteristics of *Penaeus monodon* (Fabricius, 1798) hemolymph. Comparative Haematology International. 6: 194-203.
- Van Hulten, M. C. W., Tsai, M. F., Schipper, C. A. et al. 2000. Analysis of a genomic segment of white spot syndrome virus of shrimp containing ribonucleotide reductase genes and repeat regions, Journal of Virological Methods. 81: 307-316.
- Vandergeest, P., Platong, C., Flaherty, M. et al. 1999. Shrimp aquaculture and the politics of zoning in Thailand. The 7th International conference on Thai studies, pp. 1-17. 4-8 July 1999. University of Amsterdam, Netherlands.
- Vargas-Albores, F. 1997. Shrimp immunology. Developmental & Comparative Immunology. 21(2): 206.

- Vargas-Albores, F., and Yepiz-Plascencia, G. 2000. Beta glucan binding protein and its role in shrimp immune response. Aquaculture. 191: 13-21.
- Vargas-Albores, F., Guzman, M. A., and Ochoa, J. L. 1993. A lipopolysaccharide bindind agglutinin isolated from brown shrimp (*Penaeus californiensis* Holmes) hemolymph. Comparative biochemistry and physiology. 104B: 407-413.
- Vargas-Albores, F., Jimenez-Vega, F., and Söderhäll. 1996. A plasma protein isolated from brown shrimp (*Penaeus californiensis*) which enhances the activation of prophenoloxidase system by β -1,3-glucans. Developmental and Comparative Immunology. 20: 299-306.
- Vargas-Albores, F., Jimenez-Vega, F., and Yepiz-Plascencia. 1997. Purification and comparison of β -1,3-glucans binding protein from white shrimp (*Penaeus vannami*). Comparative Biochemistry and Physiology. 116B:453-458.
- Wang, J. K. 1990. Managing shrimp pond water to reduce discharge problems. Aquacultural Engineering. 9(1): 61–73.
- Wang, Y. C., Lo, C. F., Chang, P. S. et al., 1998. Experimental infection of white spot baculovirus in som cultured and wild decapods in Taiwan. Aquaculture. 164: 221-232.
- Ward, N. E. 1991. Chemoattractants for trout and salmon. Feed Management. 42(3): 6-10.
- Weinreich, O., Koch, V., and Knippel, J. 1994. Futtermittelrechtliche Vorschriften. Buchedition Agrimedia. Hamburg, Germany.
- Wigglesworth, J. M., and Griffith, D. R. W. 1994. Carbohydrate digestion in *Penaeus monodon*. Marine Biology. 120: 571-578.
- Witteveldt, J., Vlak, J. M., Mariëlle, C. W. et al. 2004. Protection of *Penaeus monodon* against white spot syndrome virus using a WSSV subunit vaccine. Fish and Shellfish Immunology. 16: 571-579.
- Witvrouw, M., and De Clercq, E. 1997. Sulfated polysaccharides extracted from sea algae as potential antiviral drugs. General Pharmacology. 29: 497-511.
- Wiwattanachisat Y. 2002. Strategy plan of fishery commodity. Thai Fisheries Gazette. 55(4).

- Wongteerasupaya, C., Vickers, J. E., Sriurairatana, S. et al. 1995. A non-occluded, systemic baculovirus that occurs in cells of ectodermal and mesodermal origin and causes high mortality in the black tiger prawn *Penaeus monodon*. Diseases of Aquatic Organisms. 21: 69–77.
- Wu, J. L., Nishioka, T., Mori, K. et al. 2002. Preparation of an inoculum of white spot syndrome virus for challenge tests in *Penaeus japonicus*. Fish Pathology. 37: 65-69.
- Yamamoto, I., Takayama, K., Honma, K. et al. 1990. Synthesis, structure and antiviral activity of sulfates of cellulose and its branched derivatives. Carbohydrate Polymers. 14(1): 53-63.
- Yang, F., He, J., Lin, X. et al. 2001. Complete genome sequence of the shrimp white spot bacilliform virus, Journal of virology. 75: 11811–11820.
- Yeh, M. S., Chen, Y. L., and Tsai, I. H. 1998. The hemolymph clottable proteins of tiger shrimp, *Penaeus monodon*, and related species. Comparative Biochemistry and Physiology. 121B: 169-176.
- Yoshida, O., Nakashima, H., Yoshida, T. et al. 1988. Sulfation of the immunomodulating polysaccharide lentinan: A novel strategy for antivirals to human immunodeficiency virus (HIV). Biochemical Pharmacology. 37(15): 2887-2891.
- Zafran Des Rosa, B., Sugama, K., Wada, S. et al. 1992. Histological study of luminescent vibriosis deseases in hatchery-reared larvae of *Penaeus monodon*. Abstrats of the Third Asian Fisheries Forum held in Singapore from October 26-30, 1992., Manila: Asian Fisheries Society.



APPENDICES

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APPENDIX

A: Chemical reagents

1. Cacodylate (CAC buffer); pH 7.0 and kept at 4 °C.

Sodium cacodylate	2.14 g.
CaCl ₂ .2H ₂ O	0.74 g.
MgCl ₂ .6H ₂ O	10.16 g.

The solid chemicals were dissolved in distilled water, adjusted to pH 7.0 and adjusted to volume 1,000 ml in volume metric flak. This solution was sterilized by autoclave for 15 min. at 121 °C, 15 pounds pressure and kept at 4 °C in a dark bottle.

2. Anticoagulant solution (AC-1); pH 7.0, kept at 4 °C.

NaCl	26.3 g.
Trisodium citrate	8.82 g.
Citric acid	5.46 g.
Na EDTA	3.72 g.

The chemical materials were dissolved in distilled water, adjusted to pH 7.0 and adjusted to volume 1,000 ml in volume metric flak. This solution was sterilized by autoclave for 15 min. at 121 °C, 15 pounds pressure and kept at 4 °C. Mix the solution with sterilized 0.1 M glucose, previously used.

3. 0.1% Trypsin; pH 7.0±0.2, kept at 4 °C.

Trypsin	0.1 g.
CAC buffer	100 ml.

The trypsin was dissolved in CAC buffer, adjusted to pH 7.0 and adjusted to volume 100 ml. in volume metric flak. This solution was sterilized by filtration through a 0.22 µm. pore diameter of membrane filter. This fresh solution was freshly prepared, before used, and kept in a dark bottle.

4. 0.3% L-DOPA; pH 7.0±0.2, kept at 4 °C.

L-3,4-dihydroxyphenylalanine	0.3	g.
CAC buffer	100	ml.

The L-3,4-dihydroxyphenylalanine was dissolved in CAC buffer, adjusted to pH 7.0 and adjusted to volume 100 ml in volume metric flak. This solution was sterilized by filtration through a 0.22 µm. pore diameter of membrane filter. This fresh solution was freshly prepared, before used, and kept in a dark bottle.

5. 5.0% L-cysteine; pH 7.0±0.2, kept at 4 °C.

L-cystein	5.0	g.
Lobster haemolymph medium	100	ml.

The L-cystein was dissolved in lobster haemolymph medium (LHM), adjusted to pH 7.0 and adjusted to volume 100 ml in volume metric flak. This solution was sterilized by filtration through a 0.22 µm. pore diameter of membrane filter. This fresh solution was freshly prepared, before used, and kept in a dark bottle.

6. 0.3% Eosin

Eosin Y	0.3	g
95% Ethyl alcohol	100	ml.
Glacial acetic acid	0.5	ml.

The chemical materials were dissolved in 95% ethyl alcohol and adjusted to volume 100 ml in volume metric flak.

7. Divison fixation (ml/L.)

Glacial acetic acid	115 ml
95% Ethyl alcohol	330 ml
39% Formaldehyde	220 ml
Distilled water	335 ml

The chemical reagents were mixed in distilled water and adjusted to volume 1,000 ml in volume metric flak.

8. Hematoxylin reagent compound of:

Hematoxylin crystals	2.0	g.
Sodium iodate	0.4	g.
Potassium aluminium sulfate	100	g.
Citric acid	2.0	g.
Chloral hydrate	100	g.
Distilled water	1,000	ml.

The chemical materials were dissolved in distilled water and adjusted to volume 1,000 ml in volume metric flak.

APPENDIX

B: Media

1. Agar media

1.1 Tryptic soy agar (TSA); pH 7.3±0.2

Peptone from casein	17.0	g.
Peptone from soymeal	3.0	g.
D (+) glucose	2.5	g.
Sodium chloride	5.0	g.
Di-potassium hydrogen phosphate	2.5	g.
Agar powder	15	g.

Dispensed solid media in distilled water until dissolved and sterilized by autoclave for 21 min. at 121 °C, 15 pounds pressure.

1.2 Thiosulfate citrate bile salt sucrose agar (TCBSA); pH 8.6±0.2

Yeast extract	5.0	g.
Proteose peptone No.3	10.0	g.
Sodium citrate	10.0	g.
Sodiumthiosulfate	10.0	g.
Oxgall	8.0	g.
Saccharose	20.0	g.
Sodium chloride	10.0	g.
Ferric citrate	1.0	g.
Bromthymol blue	0.04	g.
Agar powder	15.0	g

Dispensed solid media in distilled water until dissolved and sterilized by autoclave for 21 min. at 121 °C, 15 pounds pressure.

1.3 Mueller Hinton agar (MHA); pH 7.3±0.2

Beef extract powder	2.0	g.
Casein hydrolysate	17.5	g.
Soluble starch	1.5	g.
Agar powder	13.0	g.

Dispensed solid media in distilled water until dissolved and sterilized by autoclave for 21 min. at 121 °C, 15 pounds pressure.

2. Both media

2.1 Mueller Hinton broth (MHB); pH 7.3±0.2

Beef extract powder	2.0	g.
Casein hydrolysate	17.5	g.
Soluble starch	1.5	g.

Dispensed solid media in distilled water until dissolved and sterilized by autoclave for 21 min. at 121 °C, 15 pounds pressure.

2.2 Lobster haemolymph medium (LHM); pH 7.6 ±0.2 and kept at 4 °C.

MEM essential amino acid with L-glutamine (2X)	500	ml.
11% NaCl (w/v)	100	ml.
11% CaCl ₂ .2H ₂ O (w/v)	100	ml.
Hepes buffer	2.38	g/L.
Steriled water	200	ml.
Salt mixture reagent compound of:	100	ml.
- KCl	0.4	g.
- NaH ₂ PO ₄ .2H ₂ O	0.05	g.

All solid materials were dissolved in distilled water and mixed with all chemical solutions adjusted to pH 7.6 with 7.5% NaHCO₃ and adjusted to volume 1,000 ml in volume metric flak. This medium was sterilized by filtration through a 0.45 μm. pore diameter of membrane filter.

APPENDIX

C: Table

Table 13. Feeding of *Penaeus monodon* shrimp number 100,000 pieces have density 25 pieces per a square meter.

Week	Weight gain (g/pcs.)	Growth rate (g/pcs/day)	Survival rate (%)	Feeding rate per shrimp weight (%)	Frequency of feedind rate per day (a time)
1	PL15-0.10	0.03	95	100	3
2	0.50	0.06	92	20	3
3	1.00	0.07	90	10	3
4	1.80	0.11	88	10	3
5	2.80	0.14	86	9.0	3
6	3.90	0.16	84	8.0	4
7	5.50	0.23	83	7.0	4
8	7.60	0.30	82	7.0	4
9	9.80	0.31	81	6.0	4
10	12.00	0.31	80	5.0	4
11	14.30	0.33	79	5.0	4
12	17.10	0.40	78	4.0	4
13	20.00	0.41	77	4.0	5
14	23.30	0.47	76	4.0	5
15	26.70	0.49	75	4.0	5
16	30.60	0.56	74	3.5	5

Table 14. Effect of polysaccharide gel (PG) from durian fruit-hulls on growth performances of *Penaeus monodon* juvenile in the trial rearing were evaluated after every 4 weeks for 8 weeks and 12 weeks feeding period, using four groups of shrimp (100 shrimps/group) in each treatments. Values are mean \pm SD. Control = 0.0% PG. ns = not significant. a, b, c = significantly different between groups, ($P<0.05$).

Subject	Shrimp number (n)	Growth performance after feeding with shrimp diets with different concentration of PG. (mean \pm SD)			
		Control	0.5% PG	1.0% PG	2.0% PG
1. body weight (gram)	4 group	0.36 \pm 0.00 ^{ns}	0.36 \pm 0.00 ^{ns}	0.36 \pm 0.00 ^{ns}	0.36 \pm 0.00 ^{ns}
		1.77 \pm 0.21 ^{ns}	1.89 \pm 0.18 ^{ns}	2.04 \pm 0.10 ^{ns}	2.03 \pm 0.21 ^{ns}
		5.90 \pm 0.37 ^b	5.65 \pm 0.19 ^b	6.06 \pm 0.16 ^{ab}	6.55 \pm 0.48 ^a
2. survival rate (%)	4 group	92.00 \pm 6.53 ^{ns}	92.00 \pm 7.30 ^{ns}	94.00 \pm 2.31 ^{ns}	89.00 \pm 6.00 ^{ns}
		57.00 \pm 5.03 ^c	75.00 \pm 8.87 ^a	69.00 \pm 6.00 ^{ab}	63.00 \pm 6.83 ^{cb}
		3.84 \pm 0.10 ^{ns}	3.84 \pm 0.10 ^{ns}	3.84 \pm 0.10 ^{ns}	3.84 \pm 0.10 ^{ns}
3. body length (mm.)	30 pcs.	65.88 \pm 4.74 ^{ns}	64.73 \pm 5.27 ^{ns}	67.93 \pm 4.82 ^{ns}	67.94 \pm 4.68 ^{ns}
		93.60 \pm 6.96 ^{ns}	93.44 \pm 7.74 ^{ns}	94.39 \pm 7.24 ^{ns}	97.27 \pm 8.48 ^{ns}
		1.88 \pm 0.51 ^{ns}	1.70 \pm 0.29 ^{ns}	1.47 \pm 0.08 ^{ns}	1.60 \pm 0.21 ^{ns}
4. FCR	4 group	2.56 \pm 0.17 ^b	1.98 \pm 0.30 ^a	2.02 \pm 0.24 ^a	2.09 \pm 0.36 ^a
		40.82 \pm 6.85 ^{ns}	43.48 \pm 6.01 ^{ns}	47.93 \pm 2.20 ^{ns}	45.15 \pm 4.86 ^{ns}
		83.79 \pm 5.03 ^b	105.91 \pm 12.45 ^a	104.74 \pm 11.60 ^a	103.44 \pm 15.93 ^a

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Table 15. Water quality during the feeding period in the trial rearing was evaluated at every week.

Environmental category	Value of water quality (minimum-maximum)
pH	7.6-8.2
Salinity (ppt)	16-21
Hardness (ppm)	2,100-3,250
Temperature (°C)	29.5-33.0
Nitrite, NO ₂ (ppm)	0.0-0.2
Total alkalinity (ppm)	100-160
Dissolved oxygen, DO (ppm)	3.5-6.0
Total ammonia nitrogen, TAN (ppm)	0.0-0.2

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Table 16. The survival rate of black tiger shrimp (*Penaeus monodon*) on challenge test with WSSV, 10^6 (1:100 dilution) by cohabitation method after 4 weeks feeding period, using three groups of the rearing shrimp (30 shrimp/group) in each treatments. Values are mean \pm SD. Control = 0.0% PG.

Date	The survival rate of black tiger shrimp with different concentration of PG. (%)			
	Control	0.5% PG	1.0% PG	2.0% PG
0	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
1	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
2	97 \pm 0.58	100 \pm 0.00	100 \pm 0.00	97 \pm 0.58
3	97 \pm 0.58	100 \pm 0.00	100 \pm 0.00	97 \pm 0.58
4	93 \pm 0.58	100 \pm 0.00	100 \pm 0.00	97 \pm 0.58
5	83 \pm 1.53	93 \pm 1.15	97 \pm 0.58	97 \pm 0.58
6	70 \pm 1.00	83 \pm 0.58	97 \pm 0.58	87 \pm 0.58
7	53 \pm 1.53	57 \pm 0.58	97 \pm 0.58	77 \pm 1.15
8	20 \pm 1.73	37 \pm 0.58	93 \pm 1.15	60 \pm 1.73
9	13 \pm 1.53	17 \pm 1.53	90 \pm 1.00	47 \pm 3.06
10	3 \pm 0.58	10 \pm 1.73	80 \pm 1.00	20 \pm 3.46
11	0 \pm 0.00	0 \pm 0.00	57 \pm 1.53	17 \pm 2.89
12	0 \pm 0.00	0 \pm 0.00	27 \pm 0.58	10 \pm 1.73
13	0 \pm 0.00	0 \pm 0.00	7 \pm 0.58	3 \pm 0.58
14	0 \pm 0.00	0 \pm 0.00	3 \pm 0.58	0 \pm 0.00

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Table 17. The survival rate of black tiger shrimp (*Penaeus monodon*) on challenge test with WSSV, 10^6 (1:100 dilution) by cohabitation method after 8 weeks feeding period, using three groups of the rearing shrimp (24 shrimp/group) in each treatments. Values are mean \pm SD. Control = 0.0% PG.

Date	The survival rate of black tiger shrimp with different concentration of PG. (%)			
	Control	0.5% PG	1.0% PG	2.0% PG
0	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
1	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
2	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
3	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
4	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
5	100 \pm 0.00	100 \pm 0.00	83 \pm 1.15	96 \pm 0.58
6	100 \pm 0.00	96 \pm 0.58	58 \pm 3.06	83 \pm 2.31
7	100 \pm 0.00	96 \pm 0.58	54 \pm 3.21	75 \pm 3.46
8	88 \pm 0.00	92 \pm 1.15	42 \pm 4.04	71 \pm 4.04
9	79 \pm 0.58	67 \pm 2.08	33 \pm 4.62	71 \pm 4.04
10	54 \pm 0.58	38 \pm 2.65	25 \pm 3.46	58 \pm 4.16
11	42 \pm 1.15	21 \pm 2.89	13 \pm 1.73	50 \pm 3.61
12	21 \pm 1.15	21 \pm 2.89	8 \pm 1.15	46 \pm 3.51
13	4 \pm 0.58	17 \pm 2.31	4 \pm 0.58	38 \pm 3.61
14	0 \pm 0.00	4 \pm 0.58	4 \pm 0.58	33 \pm 3.79

Table 18. The survival rate of black tiger shrimp (*Penaeus monodon*) on challenge test with *Vibrio harveyi* 1526, 2.15×10^7 CFU/ml by immersion method after 8 weeks feeding period, using three groups of the rearing shrimp (24 shrimp/group) in each treatments. Values are mean \pm SD. Control = 0.0% PG.

Date	The survival rate of black tiger shrimp In different concentration of PG. (%)			
	Control	0.5% PG	1.0% PG	2.0% PG
0	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00	100 \pm 0.00
1	100 \pm 1.00	67 \pm 0.58	71 \pm 0.58	88 \pm 1.73
2	50 \pm 1.00	67 \pm 0.58	67 \pm 0.58	75 \pm 3.46
3	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
4	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
5	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
6	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
7	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
8	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
9	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
10	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
11	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
12	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
13	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46
14	50 \pm 1.00	58 \pm 0.58	67 \pm 0.58	75 \pm 3.46

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Table 19. Water quality of black tiger shrimp (*Penaeus monodon*) on challenge test with WSSV, 10^6 (1:100 dilution) by cohabitation method after 4 weeks feeding period was evaluated at everyday.

Environmental category	Value of water quality (minimum-maximum)
pH	7.9-8.2
Salinity (ppt)	15
Nitrite, NO_2 (ppm)	0.0-0.5
Temperature ($^{\circ}\text{C}$)	27-28
Total ammonia nitrogen, TAN (ppm)	0.0-0.5

Table 20. Water quality of black tiger shrimp (*Penaeus monodon*) on challenge test with *Vibrio harveyi* 1526, 2.15×10^7 CFU/ml by immersion method after 8 weeks feeding period was evaluated at everyday.

Environmental category	Value of water quality (minimum-maximum)
pH	7.9-8.2
Salinity (ppt)	15
Nitrite, NO_2 (ppm)	0.0-0.5
Temperature ($^{\circ}\text{C}$)	29-30
Total ammonia nitrogen, TAN (ppm)	0.0-0.5

Table 21. Water quality of black tiger shrimp (*Penaeus monodon*) on challenge test with WSSV, 10^6 (1:100 dilution) by cohabitation method after 8 weeks feeding period was evaluated at everyday.

Environmental category	Value of water quality (minimum-maximum)
pH	7.9-8.2
Salinity (ppt)	15
Nitrite, NO_2 (ppm)	0.0-0.5
Temperature ($^{\circ}\text{C}$)	27-28
Total ammonia nitrogen, TAN (ppm)	0.0-0.5

Table 22. Antimicrobial activity of PG inhibits growth of bacteria *Vibrio harveyi* 1526 by agar diffusion metod. nz = no inhibition zone.

Concentration of PG (%)	Diameter of inhibition zone (mm.)					
	5.0	2.5	1.25	0.63	0.32	NSS
5.0	19.0	18.8	20.1	22.9	19.6	22.2
2.5	15.1	15.6	15.9	17.5	18.6	16.1
1.25	11.4	11.7	11.6	12.7	12.5	13.0
0.63	10.8	11.4	10.8	10.6	10.9	9.7
0.32	9.2	9.6	9.1	8.2	8.5	8.7
NSS	nz	Nz	nz	Nz	nz	nz

Table 23. Total haemocyte count (THC) of *Penaeus monodon* juvenile shrimp were counted using haemacytometer after 8 weeks feeding period, using 12 shrimps/group in each treatments. Values are mean \pm SD. Control = 0.0% PG.

No. sample (piece)	Total haemocyte count (THC) (10^7 cells/ml)			
	Control	0.5% PG	1.0% PG	2.0% PG
1	1.75 \pm 0.25	1.29 \pm 0.19	2.13 \pm 0.13	2.33 \pm 0.19
2	1.21 \pm 0.26	1.38 \pm 0.13	1.04 \pm 0.19	1.58 \pm 0.26
3	1.54 \pm 0.19	1.58 \pm 0.38	1.08 \pm 0.19	0.83 \pm 0.07
4	0.58 \pm 0.07	1.00 \pm 0.22	1.00 \pm 0.25	2.33 \pm 0.19
5	1.58 \pm 0.07	1.25 \pm 0.13	1.79 \pm 0.19	1.54 \pm 0.19
6	1.33 \pm 0.19	0.79 \pm 0.07	1.54 \pm 0.07	1.46 \pm 0.07
7	1.17 \pm 0.07	2.04 \pm 0.19	1.21 \pm 0.26	2.46 \pm 0.19
8	0.75 \pm 0.13	0.88 \pm 0.22	2.04 \pm 0.07	1.96 \pm 0.07
9	0.54 \pm 0.07	1.17 \pm 0.19	2.08 \pm 0.19	1.04 \pm 0.07
10	0.54 \pm 0.07	1.79 \pm 0.26	1.92 \pm 0.07	1.00 \pm 0.22
11	1.63 \pm 0.13	0.58 \pm 0.07	1.63 \pm 0.22	2.21 \pm 0.19
12	1.29 \pm 0.14	0.67 \pm 0.07	1.25 \pm 0.25	1.46 \pm 0.07
Average	1.16 \pm 0.45	1.20 \pm 0.45	1.56 \pm 0.43	1.68 \pm 0.57

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Table 24. Total phenoloxidase activity of *Penaeus monodon* juvenile shrimp were measured in the haemolymph after 8 weeks feeding period, using 12 shrimps/group in each treatments. Control = 0.0% PG.

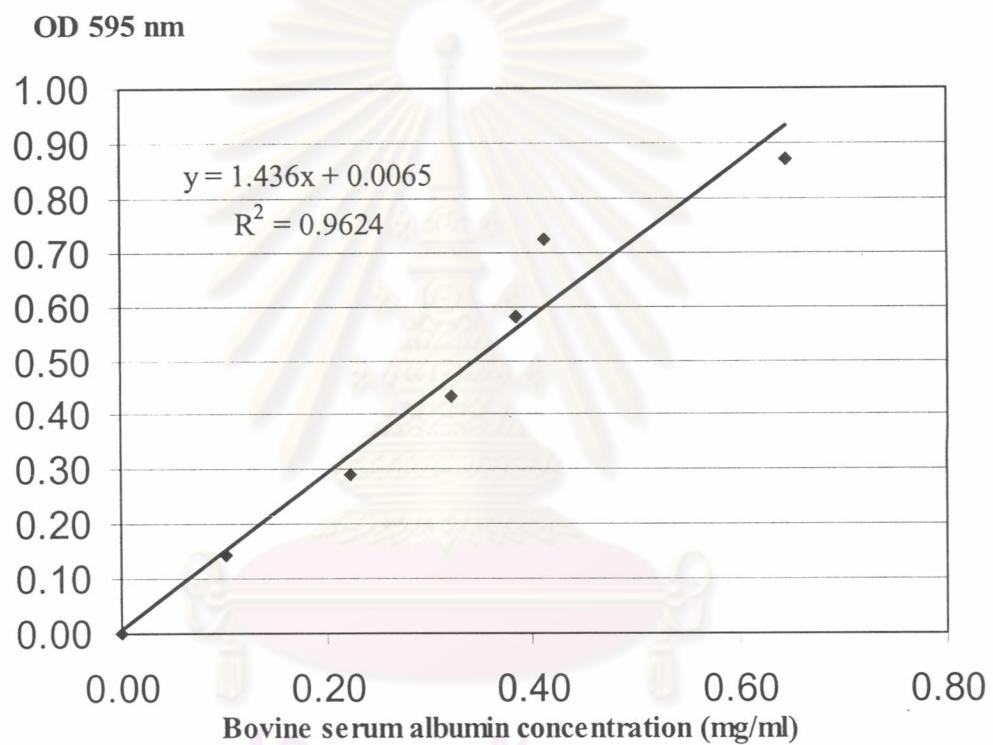
No. sample (piece)	The phenoloxidase activity (PO) (units/min./mg protein)			
	Control	0.5% PG	1.0% PG	2.0% PG
1	620.00	812.50	659.57	777.78
2	547.17	207.79	898.55	1125.00
3	538.46	928.57	740.74	571.43
4	571.43	702.13	1232.56	956.52
5	584.62	857.14	1290.32	1235.29
6	595.74	844.83	1078.43	1083.33
7	596.49	980.77	1319.15	1081.08
8	509.43	613.64	1035.71	1127.66
9	756.10	637.93	698.11	511.11
10	839.29	651.16	615.38	743.59
11	741.94	513.16	714.29	948.72
12	411.76	722.22	518.52	558.14
Average	609.37±117.71	705.99±208.97	900.11±281.85	893.30±252.58

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APPENDIX

D: Figure

Figure 31. Standard curve of bovine serum albumin protein by Bradford protein assay for phenoloxidase activity test.



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APPENDIX

E: Statistics

The biomass of shrimp feeding period for 4 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	40.8225	6.85032	3.42516	29.9221	51.7229	30.66	45.00
2	4	43.4750	6.00947	3.00473	33.9126	53.0374	36.54	51.12
3	4	47.9250	2.20158	1.10079	44.4218	51.4282	45.77	50.16
4	4	45.1525	4.85873	2.42936	37.4212	52.8838	39.79	51.36
Total	16	44.3438	5.42228	1.35557	41.4544	47.2331	30.66	51.36

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	106.533	3	35.511	1.274	.328
Within Groups	334.484	12	27.874		
Total	441.018	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	-2.6525	3.73321	.491	-10.7865	5.4815
	3	-7.1025	3.73321	.081	-15.2365	1.0315
	4	-4.3300	3.73321	.269	-12.4640	3.8040
2	1	2.6525	3.73321	.491	-5.4815	10.7865
	3	-4.4500	3.73321	.256	-12.5840	3.6840
	4	-1.6775	3.73321	.661	-9.8115	6.4565
3	1	7.1025	3.73321	.081	-1.0315	15.2365
	2	4.4500	3.73321	.256	-3.6840	12.5840
	4	2.7725	3.73321	.472	-5.3615	10.9065
4	1	4.3300	3.73321	.269	-3.8040	12.4640
	2	1.6775	3.73321	.661	-6.4565	9.8115
	3	-2.7725	3.73321	.472	-10.9065	5.3615

The biomass of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	83.7925	5.0275	2.5138	75.7926	91.7924	78.82	90.72
2	4	105.9075	12.4472	6.2236	86.1013	125.7137	88.32	116.97
3	4	104.7350	11.6018	5.8009	86.2739	123.1961	93.44	117.80
4	4	103.4350	15.9292	7.9646	78.0881	128.7819	82.18	120.78
Total	16	99.4675	14.2087	3.5522	91.8962	107.0388	78.82	120.78

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	1322.667	3	440.889	3.102	.067
Within Groups	1705.651	12	142.138		
Total	3028.319	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	-22.1150(*)	8.4302	.022	-40.4829	-3.7471
	3	-20.9425(*)	8.4302	.029	-39.3104	-2.5746
	4	-19.6425(*)	8.4302	.038	-38.0104	-1.2746
2	1	22.1150(*)	8.4302	.022	3.7471	40.4829
	3	1.1725	8.4302	.892	-17.1954	19.5404
	4	2.4725	8.4302	.774	-15.8954	20.8404
3	1	20.9425(*)	8.4302	.029	2.5746	39.3104
	2	-1.1725	8.4302	.892	-19.5404	17.1954
	4	1.3000	8.4302	.880	-17.0679	19.6679
4	1	19.6425(*)	8.4302	.038	1.2746	38.0104
	2	-2.4725	8.4302	.774	-20.8404	15.8954
	3	-1.3000	8.4302	.880	-19.6679	17.0679

* The mean difference is significant at the 0.05 level.

The feed conversion ratio (FCR) of shrimp feeding period for 4 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	1.8750	.5055	.2528	1.0706	2.6794	1.59	2.63
2	4	1.6950	.2931	.1465	1.2286	2.1614	1.36	2.07
3	4	1.4700	8.165E-02	4.082E-02	1.3401	1.5999	1.39	1.55
4	4	1.6000	.2099	.1050	1.2660	1.9340	1.35	1.85
Total	16	1.6600	.3189	7.972E-02	1.4901	1.8299	1.35	2.63

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	.349	3	.116	1.185	.357
Within Groups	1.177	12	9.805E-02		
Total	1.525	15			

Post Hoc tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	.1800	.2214	.432	-.3024	.6624
	3	.4050	.2214	.092	-7.7424E-02	.8874
	4	.2750	.2214	.238	-.2074	.7574
2	1	-.1800	.2214	.432	-.6624	.3024
	3	.2250	.2214	.330	-.2574	.7074
	4	9.5000E-02	.2214	.675	-.3874	.5774
3	1	-.4050	.2214	.092	-.8874	7.742E-02
	2	-.2250	.2214	.330	-.7074	.2574
	4	-.1300	.2214	.568	-.6124	.3524
4	1	-.2750	.2214	.238	-.7574	.2074
	2	-9.5000E-02	.2214	.675	-.5774	.3874
	3	.1300	.2214	.568	-.3524	.6124

The feed conversion ratio (FCR) of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

Group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	2.5625	.1666	8.330E-02	2.2974	2.8276	2.34	2.74
2	4	1.9825	.3017	.1509	1.5024	2.4626	1.71	2.41
3	4	2.0175	.2403	.1202	1.6351	2.3999	1.76	2.26
4	4	2.0875	.3630	.1815	1.5099	2.6651	1.77	2.61
Total	16	2.1625	.3465	8.663E-02	1.9779	2.3471	1.71	2.74

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	.876	3	.292	3.789	.040
Within Groups	.925	12	7.708E-02		
Total	1.801	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	.5800(*)	.1963	.012	.1523	1.0077
	3	.5450(*)	.1963	.017	.1173	.9727
	4	.4750(*)	.1963	.032	4.728E-02	.9027
2	1	-.5800(*)	.1963	.012	-1.0077	-.1523
	3	-3.5000E-02	.1963	.861	-.4627	.3927
	4	-.1050	.1963	.603	-.5327	.3227
3	1	-.5450(*)	.1963	.017	-.9727	-.1173
	2	3.5000E-02	.1963	.861	-.3927	.4627
	4	-7.0000E-02	.1963	.728	-.4977	.3577
4	1	-.4750(*)	.1963	.032	-.9027	-4.7278E-02
	2	.1050	.1963	.603	-.3227	.5327
	3	7.0000E-02	.1963	.728	-.3577	.4977

* The mean difference is significant at the 0.05 level.

The total length of shrimp feeding period for 4 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean (mm)	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1.00	12	65.8750	4.73654	1.36732	62.8655	68.8845	55.60	75.20
2.00	12	64.7250	5.27087	1.52157	61.3761	68.0739	55.70	71.00
3.00	12	67.9250	4.82138	1.39181	64.8616	70.9884	62.00	79.70
4.00	12	67.9417	4.67711	1.35017	64.9700	70.9134	58.90	72.60
Total	48	66.6167	4.92468	.71082	65.1867	68.0466	55.60	79.70

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	91.150	3	30.383	1.275	.295
Within Groups	1048.717	44	23.834		
Total	1139.867	47			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	1.1500	1.99309	.567	-2.8668	5.1668
	3	-2.0500	1.99309	.309	-6.0668	1.9668
	4	-2.0667	1.99309	.305	-6.0835	1.9501
2	1	-1.1500	1.99309	.567	-5.1668	2.8668
	3	-3.2000	1.99309	.116	-7.2168	.8168
	4	-3.2167	1.99309	.114	-7.2335	.8001
3	1	2.0500	1.99309	.309	-1.9668	6.0668
	2	3.2000	1.99309	.116	-.8168	7.2168
	4	-.0167	1.99309	.993	-4.0335	4.0001
4	1	2.0667	1.99309	.305	-1.9501	6.0835
	2	3.2167	1.99309	.114	-.8001	7.2335
	3	.0167	1.99309	.993	-4.0001	4.0335

The total length of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean (mm)	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	30	93.6033	6.96437	1.27151	91.0028	96.2039	76.60	104.20
2	30	93.4400	7.74203	1.41349	90.5491	96.3309	80.00	113.70
3	30	94.3900	7.24247	1.32229	91.6856	97.0944	81.60	108.70
4	30	97.2667	8.47944	1.54813	94.1004	100.4329	79.40	114.00
Total	120	94.6750	7.68892	.70190	93.2852	96.0648	76.60	114.00

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	284.150	3	94.717	1.627	.187
Within Groups	6751.075	116	58.199		
Total	7035.225	119			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) Group	(J) Group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	.1633	1.96975	.934	-3.7380	4.0647
	3	-.7867	1.96975	.690	-4.6880	3.1147
	4	-3.6633	1.96975	.065	-7.5647	.2380
2	1	-.1633	1.96975	.934	-4.0647	3.7380
	3	-.9500	1.96975	.631	-4.8513	2.9513
	4	-3.8267	1.96975	.054	-7.7280	.0747
3	1	.7867	1.96975	.690	-3.1147	4.6880
	2	.9500	1.96975	.631	-2.9513	4.8513
	4	-2.8767	1.96975	.147	-6.7780	1.0247
4	1	3.6633	1.96975	.065	-.2380	7.5647
	2	3.8267	1.96975	.054	-.0747	7.7280
	3	2.8767	1.96975	.147	-1.0247	6.7780

The survival of shrimp feeding period for 4 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

Group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	23.0000	1.63299	.81650	20.4015	25.5985	21.00	25.00
2	4	23.0000	1.82574	.91287	20.0948	25.9052	21.00	25.00
3	4	23.5000	.57735	.28868	22.5813	24.4187	23.00	24.00
4	4	22.2500	1.50000	.75000	19.8632	24.6368	21.00	24.00
Total	16	22.9375	1.38894	.34724	22.1974	23.6776	21.00	25.00

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	3.188	3	1.063	.495	.692
Within Groups	25.750	12	2.146		
Total	28.938	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	.0000	1.03582	1.000	-2.2569	2.2569
	3	-.5000	1.03582	.638	-2.7569	1.7569
	4	.7500	1.03582	.483	-1.5069	3.0069
2	1	.0000	1.03582	1.000	-2.2569	2.2569
	3	-.5000	1.03582	.638	-2.7569	1.7569
	4	.7500	1.03582	.483	-1.5069	3.0069
3	1	.5000	1.03582	.638	-1.7569	2.7569
	2	.5000	1.03582	.638	-1.7569	2.7569
	4	1.2500	1.03582	.251	-1.0069	3.5069
4	1	-.7500	1.03582	.483	-3.0069	1.5069
	2	-.7500	1.03582	.483	-3.0069	1.5069
	3	-1.2500	1.03582	.251	-3.5069	1.0069

The survival of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	14.25	1.26	.63	12.25	16.25	13	16
2	4	18.75	2.22	1.11	15.22	22.28	16	21
3	4	17.25	1.50	.75	14.86	19.64	16	19
4	4	15.75	1.71	.85	13.03	18.47	14	18
Total	16	16.50	2.31	.58	15.27	17.73	13	21

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	45.000	3	15.000	5.143	.016
Within Groups	35.000	12	2.917		
Total	80.000	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	-4.50(*)	1.21	.003	-7.13	-1.87
	3	-3.00(*)	1.21	.029	-5.63	-.37
	4	-1.50	1.21	.238	-4.13	1.13
2	1	4.50(*)	1.21	.003	1.87	7.13
	3	1.50	1.21	.238	-1.13	4.13
	4	3.00(*)	1.21	.029	.37	5.63
3	1	3.00(*)	1.21	.029	.37	5.63
	2	-1.50	1.21	.238	-4.13	1.13
	4	1.50	1.21	.238	-1.13	4.13
4	1	1.50	1.21	.238	-1.13	4.13
	2	-3.00(*)	1.21	.029	-5.63	-.37
	3	-1.50	1.21	.238	-4.13	1.13

* The mean difference is significant at the 0.05 level.

The body weight of shrimp feeding period for 4 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	1.7675	.21407	.10703	1.4269	2.1081	1.46	1.95
2	4	1.8875	.18062	.09031	1.6001	2.1749	1.74	2.13
3	4	2.0400	.09866	.04933	1.8830	2.1970	1.93	2.15
4	4	2.0325	.20759	.10379	1.7022	2.3628	1.73	2.19
Total	16	1.9319	.19951	.04988	1.8256	2.0382	1.46	2.19

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	.203	3	.068	2.064	.159
Within Groups	.394	12	.033		
Total	.597	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2.00	-.1200	.12810	.367	-.3991	.1591
	3.00	-.2725	.12810	.055	-.5516	.0066
	4.00	-.2650	.12810	.061	-.5441	.0141
2	1.00	.1200	.12810	.367	-.1591	.3991
	3.00	-.1525	.12810	.257	-.4316	.1266
	4.00	-.1450	.12810	.280	-.4241	.1341
3	1.00	.2725	.12810	.055	-.0066	.5516
	2.00	.1525	.12810	.257	-.1266	.4316
	4.00	.0075	.12810	.954	-.2716	.2866
4	1.00	.2650	.12810	.061	-.0141	.5441
	2.00	.1450	.12810	.280	-.1341	.4241
	3.00	-.0075	.12810	.954	-.2866	.2716

The body weight of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	4	5.8975	.3689	.1845	5.3105	6.4845	5.63	6.43
2	4	5.6500	.1885	9.425E-02	5.3500	5.9500	5.52	5.93
3	4	6.0625	.1638	8.189E-02	5.8019	6.3231	5.84	6.20
4	4	6.5475	.4758	.2379	5.7903	7.3047	5.87	6.98
Total	16	6.0394	.4470	.1117	5.8012	6.2775	5.52	6.98

ANOVA

Analysis

	Sum of Squares	Df	Mean Square	F	Sig.
Between Groups	1.722	3	.574	5.404	.014
Within Groups	1.275	12	.106		
Total	2.996	15			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	.2475	.2305	.304	-.2546	.7496
	3	-.1650	.2305	.488	-.6671	.3371
	4	-.6500(*)	.2305	.015	-1.1521	-.1479
2	1	-.2475	.2305	.304	-.7496	.2546
	3	-.4125	.2305	.099	-.9146	8.962E-02
	4	-.8975(*)	.2305	.002	-1.3996	-.3954
3	1	.1650	.2305	.488	-.3371	.6671
	2	.4125	.2305	.099	-8.9618E-02	.9146
	4	-.4850	.2305	.057	-.9871	1.712E-02
4	1	.6500(*)	.2305	.015	.1479	1.1521
	2	.8975(*)	.2305	.002	.3954	1.3996
	3	.4850	.2305	.057	-1.7118E-02	.9871

* The mean difference is significant at the 0.05 level.

The total hemocyte count (THC) of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

One way

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	12	1.1592	.4484	.1294	.8743	1.4441	.54	1.75
2	12	1.2017	.4490	.1296	.9164	1.4870	.58	2.04
3	12	1.5592	.4313	.1245	1.2851	1.8332	1.00	2.13
4	12	1.6833	.5668	.1636	1.3232	2.0434	.83	2.46
Total	48	1.4008	.5146	7.427E-02	1.2514	1.5502	.54	2.46

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	2.435	3	.812	3.569	.021
Within Groups	10.009	44	.227		
Total	12.445	47			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) group	(J) group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	-4.2500E-02	.1947	.828	-.4349	.3499
	3	-.4000(*)	.1947	.046	-.7924	-7.5789E-03
	4	-.5242(*)	.1947	.010	-.9166	-.1317
2	1	4.2500E-02	.1947	.828	-.3499	.4349
	3	-.3575	.1947	.073	-.7499	3.492E-02
	4	-.4817(*)	.1947	.017	-.8741	-8.9246E-02
3	1	.4000(*)	.1947	.046	7.579E-03	.7924
	2	.3575	.1947	.073	-3.4921E-02	.7499
	4	-.1242	.1947	.527	-.5166	.2683
4	1	.5242(*)	.1947	.010	.1317	.9166
	2	.4817(*)	.1947	.017	8.925E-02	.8741
	3	.1242	.1947	.527	-.2683	.5166

* The mean difference is significant at the 0.05 level.

The phenoloxidase activity of shrimp feeding period for 8 weeks was statistical analyses by using one way ANOVA test in version 11.5.

Descriptives

Analysis

group	N	Mean	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
1	12	609.3692	117.71273	33.98074	534.5781	684.1603	411.76	839.29
2	12	705.9867	208.96827	60.32394	573.2146	838.7588	207.79	980.77
3	12	900.1108	281.85145	81.36351	721.0310	1079.1907	518.52	1319.15
4	12	893.3042	252.58072	72.91377	732.8220	1053.7863	511.11	1235.29
Total	48	777.1927	250.54339	36.16282	704.4425	849.9429	207.79	1319.15

ANOVA

Analysis

	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	741909.33 3	3	247303.111	4.927	.005
Within Groups	2208374.1 44	44	50190.321		
Total	2950283.4 77	47			

Post Hoc Tests

Multiple Comparisons

Dependent Variable: Analysis

LSD

(I) Group	(J) Group	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
1	2	-96.6175	91.46067	.297	-280.9444	87.7094
	3	-290.7417(*)	91.46067	.003	-475.0685	-106.4148
	4	-283.9350(*)	91.46067	.003	-468.2619	-99.6081
2	1	96.6175	91.46067	.297	-87.7094	280.9444
	3	-194.1242(*)	91.46067	.039	-378.4510	-9.7973
	4	-187.3175(*)	91.46067	.047	-371.6444	-2.9906
3	1	290.7417(*)	91.46067	.003	106.4148	475.0685
	2	194.1242(*)	91.46067	.039	9.7973	378.4510
	4	6.8067	91.46067	.941	-177.5202	191.1335
4	1	283.9350(*)	91.46067	.003	99.6081	468.2619
	2	187.3175(*)	91.46067	.047	2.9906	371.6444
	3	-6.8067	91.46067	.941	-191.1335	177.5202

* The mean difference is significant at the 0.05 level.

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