

## REFERENCES

- Abdullah, A. R., Kumar, A., and Chapman, J. C. 1994. Inhibition of acetylcholinesterase in fresh water shrimp (*Paratya australiensis*) by profenofos. **Environmental Toxicology and Chemistry** 13:1861-1866
- Amer, S. M., and Fahmy, M. A. 1982. Cytogenetic effects of pesticides: I. Induction of micronuclei in mouse bone marrow by the insecticide Dursban. **Mutation Research-Genetic Toxicology** 101: 247-255.
- Anderson, D., Yu, T. W., and McGregor, D. 1998. Comet assay responses as indicators of carcinogen exposure. **Mutagenesis** 13: 539-555.
- APHA, AWWA, WEF. 1992. **Standard Methods for the Examination of Water and Wastewater**. Washington. DC.
- Ausubel, F. M., Brent, R., Kingston, R. E., Moore, D. D., Seidman, J. G., Smith, J. A., and Struhl, K. 1989. **Current protocols in molecular biology**. New York / John Wiley & Sons.
- Bagchi, D., Bagchi, M., Hassoun, E. A., and Stohs, S. J. 1995. In vitro and in vivo generation of reactive oxygen species, DNA damage, and lactate dehydrogenase leakage by selected pesticides. **Toxicology** 104: 129- 140.
- Bagchi, D., Bhattacharya, G., and Stohs, S. J. 1996. In vitro and in vivo induction of heat shock (stress) protein (Hsp) gene expression by selected pesticides. **Toxicology** 112: 57-68.
- Ballarin, P. 2004. Pesticides in water streams in Northern Thailand. **Diploma Thesis**. Department of Soil Science. Technische Universitat Berlin. Germany.
- Barata, C., Solayan, A., and Porte, C. 2004. Role of B-esterases in assessing toxicity of organophosphorus (chlorpyrifos, malathion) and carbamate (carbofuran) pesticides to *Daphnia magna*. **Aquatic Toxicology** 66:125-139.
- Basack, S. B., Oneto, M. L., Fuchs, J. S., Wood, J. S. and Kesten, E. M. 1998. Esterases of *Corbicula fluminea* as biomarker of exposure to organophosphorous pesticides. **Bulletin of Environmental Contamination and Toxicology** . 61: 569-576.
- Baskaran, S., Kookana, R. S., and Naidu, R. 1999. Degradation of bifenthrin, chlorpyrifos and imidacloprid in soil and bedding materials at termiticidal application rates. **Pesticide Science** 55: 1222-1228.

- Betti, C., Davini, T., Giannessi, L., Loprieno, N., and Barale, R. 1994. Microgel electrophoresis assay (comet test) and SCE analysis in human lymphocytes from 100 normal subjects. **Mutation Research-Fundamental and Molecular Mechanisms of Mutagenesis** 307: 323-333.
- Błasiak, J., Jałoszynski, P., Trzeciak, A., and Szyfter, K. 1999. In vitro studies on the genotoxicity of the organophosphorus insecticide malathion and its two analogues. **Mutation Research-Genetic Toxicology and Environmental Mutagenesis** 445: 275-283.
- Bocquené, G., Roig, A., Fournier, D. 1997. Evidence for the presence of a soluble acetylcholinesterase insensitive to organophosphate and carbamate indicators **FEBS Letters** 407: 261-266.
- Bocquene', G., Galgani, F., Burgeot, T., Le-Dean, L., and Truquet, P. 1993 Acetylcholinesterase levels in marine organisms along French coasts. **Marine Pollution Bulletin** 26: 101-106.
- Bradford, M. M. 1976. A rapid method and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. **Analytical Biochemistry** 72: 248-254.
- Buethong, A. 2004. Cloning and characterization of heat shock protein genes from the haemocytes of the Black Tiger Shrimp *Peneaus monodon*. **Master degree thesis**. Program Biotechnology, Faculty of Science, Chulalongkorn University. 200 pp. .
- Bulusu, S., and Chakravarty, I. 1986. Subacute Administration of Organophosphorus Pesticides and Hepatic Drug Metabolizing Enzyme Activity in Normal and Malnourished Rats **Bulletin of Environmental Contamination and Toxicology** 36: 73-80.
- Chakravarty I., and Sreedhar R. 1982. Interaction between parathion toxicity and protein malnutrition. **Environmental Research** 27:179-184.
- Choucroun, P., Gillet, D., Dorange, G., Sawicki, B., and Dewitte, J. D. 2001. Comet assay and early apoptosis. **Mutation Research** 478: 89-96.
- Ciglasch, H. 2003. Insecticide dynamics in the soil environment of a tropical lychee plantation: a case study from Northern Thailand. **Diploma Thesis**. Department of Soil Science. Technische Universitat Berlin. Germany.

- Clark, J. R., Patrick, J. M., Middaugh, D. P., and Moore, J. C. 1985. Relative sensitivity of six estuarine fishes to carbophenothion, chlorpyrifos, and fenvalerate. **Ecotoxicology and Environmental Safety** 10: 382-390.
- Colombo, A., Orsi, F., and Bonfanti, P. 2005. Exposure to the organophosphorus pesticide chlorpyrifos inhibits acetylcholinesterase activity and affects muscular integrity in *Xenopus laevis* larvae. **Chemosphere** 61: 1665-1671.
- Community Pharmacy Association Thailand. 2004. **Situation and Information Review of High Risk Insecticide : Chlorpyrifos and Diazinon**. Available from: [http://www.pharcpa.com/admin/info\\_report.asp?info\\_id=000238](http://www.pharcpa.com/admin/info_report.asp?info_id=000238)
- Cooper, N. L., Bidwell, J. R. 2006. Cholinesterase inhibition and impacts on behavior of the Asian clam, *Corbicula fluminea*, after exposure to an organophosphate insecticide. **Aquatic Toxicology** 76: 258-267.
- Danielson, P. B., Foster, J. L .M., McMahill, M. M., Smith, M. K., and Fogleman, J. C. 1998. Induction by alkaloids and phenobarbital of Family 4 Cytochrome P450s in *Drosophila*: evidence for involvement in host plant utilization. **Molecular and General Genetics**. 259: 54-59.
- Darnutra, V. 2004. **Knowledge platform on chemical safety: Hazardous substance import value**. Available from: <http://161.200.134.28/Stat-Import.asp>.
- Dauphin-Villemant, C., Bocking, D., Tom, M., and Malbec, M. 1999. Cloning of a novel cytochrome P450 (*CYP4C15*) differentially expressed in the steroidogenic glands of an arthropod. **Biochemical and Biophysical Research Communications** 264: 413-418.
- David, P., Dauphin-Villemant, C., Mesneau, A., and Meyran, J. C. 2003. Molecular approach to aquatic environmental bioreporting: differential response to environment inducers of cytochrome P450 monooxygenase genes in the detritivorous subalpine planktonic crustacean, *Daphnia pulex*. **Molecular Ecology** 12: 2473-2481.
- De Boeck, M., Touil, N., De Visscher, G., Vande, P. A., and Kirsch-Volders, M. 2000. Validation and implementation of an internal standard in comet assay **Mutation Research** 469: 181-197.
- Dearfield, K. L., Stack, H. F., Quest, J. A., Whiting, R. J., and Waters, M. D. 1993. A survey of EPA/OPP and open literature data on selected pesticide chemicals tested for mutagenicity. I. Introduction and first ten chemicals **Mutation Research** 297: 197-233.

- Department of agriculture, 2007. **Residue toxicity of chlorpyrifos.** Available from:  
<http://www.doa.go.th/th>ShowArticles.aspx?id=1679>.
- Devi, K. P., Pandian, K., and Kumar, N. S. 2005. Cholinesterase activity in clam *Meretrix casta*: possible biomarker for organophosphate pesticide pollution. **Bulletin of Environmental Contamination and Toxicology** 74: 250-255.
- Eder, K. J., Leutenegger, C. M., Wilson, B. W., and Werner, I. 2004. Molecular and cellular biomarker responses to pesticide exposure in juvenile chinook salmon (*Oncorhynchus tshawytscha*). **Marine Environmental Research** 58: 809-813.
- Ellman, G. L., Courtney, K. D., Andres, V., Featherstone, R. M. 1961. A new and rapid colorimetric determination of acetylcholinesterase activity. **Biochemical Pharmacology** 7: 88-95.
- Escartin, E., and Porte, C. 1996. Acetylcholinesterase inhibition in the crayfish *Procambarus clarkii* exposed to fenitrothion. **Ecotoxicology and Developmental Safety** 34: 160-164.
- Escartin, E., and Porte, C. 1997. The use of cholinesterase and carboxylesterase activities from *Mytilus galloprovincialis* in pollution monitoring. **Environmental Toxicology and Chemistry**. 16: 2090-2095.
- Fairbairn, D. W., Walburger, D. K., Fairbairn, J. J., and O'Neill, K. L. 1996. Key morphologic changes and DNA strand breaks in human lymphoid cells: discriminating apoptosis from necrosis. **Scanning** 18: 407-416.
- Forget, J., Beliaeff, B., and Bocquene, G. 2003. Acetylcholinesterase activity in copepods (*Tigriopus brevicornis*) from the Vilaine River Estuary, France, as biomarker of neurotoxic contaminant. **Aquatic Toxicology** 62(3): 195-204.
- Fossi, M. C., Savelli, C., and Casini, S. 1998. Mixed function oxidase induction in *Carcinus aetuari*. Field and experimental studies for the evaluation of toxicological risk due to Mediterranean contaminants. **Comparative Biochemistry and Physiology Part C** 121: 321-331.
- Fournier, D., Bride, J. M., Poirie, M., Berge, J. B., and Plapp, F. W. 1992. Insect glutathione S-transferases. Biochemical characteristics of the major forms of houseflies susceptible and resistant to insecticides. **Journal of Biological Chemistry** 267: 1840-1845.
- Frei, E., Kuchenmeister, F., Gliniorz, R., Breuer, A., and Schmezer, P. 2001. N-nitrosodimethylamine is activated in microsomes from hepatocytes to

- reactive metabolites which damage DNA of non-parenchymal cells in rat liver. **Toxicology Letters** 123: 227-234.
- Fu, J., Xie, P. 2006. The acute effects of microcystin LR on the transcription of nine glutathione S-transferase genes in common carp *Cyprinus carpio* L. **Aquatic Toxicology** 80: 261-266.
- Fujikawa, Y., Satoh, T., Suganuma, A., Suzuki, S., Niikura, Y., Yui, S., and Yamaura, Y. 2005. Extremely sensitive biomarker of acute organophosphorus insecticide exposure. **Human & Experimental Toxicology** 24(6): 333-336.
- Fulton, M. H., and Key, P. B. 2001. Acetylcholinesterase inhibition in estuarine fish and invertebrates as an indicator of organophosphorus insecticide exposure and effects. **Environmental Toxicology and Chemistry** 20: 37-45.
- Giddings, J. M., Biever, R. C., and Racke, K. D. 1997. Fate of chlorpyrifos in outdoor pond microcosms and effects on growth and survival of bluegill sunfish. **Environmental Toxicology and Chemistry** 16: 2353-2362.
- Giordano, G., Afsharinejad, Z., Guizzetti, M., Vitalone, A., Kavanagh, T. J., and Costa, L. G. 2007. Organophosphorus insecticides chlorpyrifos and diazinon and oxidative stress in neuronal cells in a genetic model of glutathione deficiency. **Toxicology and Applied Pharmacology** 19: 181-189.
- Giovanelli, L., Cozzi, A., Guarnieri, I., Dolara, P., and Moroni, F. 2002. Comet assay as a novel approach for studying DNA damage in focal cerebral Ischemia: Differential effects of NMDA receptor antagonists and poly (ADP-ribose) polymerase inhibitors. **Journal of Cerebral Blood Flow and Metabolism** 22: 697-704.
- Goel, A., Dani, V., and Dhawan, D. K. 2005. Protective effects of zinc on lipid peroxidation, antioxidant enzymes and hepatic histoarchitecture in chlorpyrifos-induced toxicity. **Chemico-Biological Interactions** 156: 131-140.
- Guilhermino, L., Lopes, M. C., Carvalho, A. P., and Soares, A. M. V. M. 1996. Inhibition of acetylcholinesterase activity as effect criterion in acute tests with juvenile *Daphnia magna*. **Chemosphere** 32: 727-738.
- Guillhermino, J., Reyes, G., Leyva, N. R., Millan, O. A., and Lazcano, G. A. 2002. Effects of pesticides on DNA and protein of shrimp larvae *Litopenaeus stylirostris* of the California gulf. **Ecotoxicology and Environmental Safety** 53: 191-195.

- Guizzetti, M., Pathak, S., Giordano, G., and Costa, L. G. 2005. Effect of organophosphorus insecticides and their metabolites on astroglial cell proliferation. **Toxicology** 215: 182-190.
- Habig, C., DiGiulio, R. T., Nomeir, A. A., and Abou-Donia, M. B. 1986. Comparative toxicity, cholinergic effects and tissue levels of *S,S,S-Tri-n-butyl phosphorotrichioate* (DEF) to Channel catfish (*Ictalurus punctatus*) and Blue crab (*Callinectes sapidus*). **Aquatic Toxicology** 9:193-206
- Halander, A. 2003. Biological markers in alcoholism. Journal of neural transmission. Supplementum 66: 15-32.
- Harford, A. J., O'Halloran, K., and Wright, P. F. A. 2005. The effects of in vitro pesticide exposures on the phagocytic function of four native Australian freshwater fish. **Aquatic Toxicology** 75: 330-342.
- Hartmann, A. et al. 2003. Recommendations for conducting the in vivo alkaline comet assay. **Mutagenesis** 18: 45-51.
- Hartmann, A., and Speit, G. 1997. The contribution of cytotoxicity to DNA-effects in the single cell gel test (comet assay). **Toxicology Letters** 90, 183-188.
- Hartmann, A., Herkommer, K., Gluck, M., and Speit, G. 1995. DNA-damaging effect of cyclophosphamide on human blood cells in vivo and in vitro studied with the single-cell gel test (comet assay). **Environmental and Molecular Mutagenesis** 25: 180-187.
- Hartmann, A., Kiskinis, E., Fjaellman, A., and Suter, W. 2001. Influence of cytotoxicity and compound precipitation on test results in the alkaline comet assay. **Mutation Research** 497: 199-212.
- Henderson, L., Wolfreys, A., Fedyk, J., Bourner, C., and Windebank, S. 1998. The ability of the comet assay to discriminate between genotoxins and cytotoxins **Mutagenesis** 13: 89-94.
- Hernández, A. F., Gómez, M. A., Pena, G., Gil, F., Rodrigo, L., Villanueva, E., and Pla, A. 2004. Effect of long-term exposure to pesticides on plasma esterase from plastic greenhouse workers. **Journal of Toxicology and Environmental Health, Part A** 67: 1095-1108.
- Herzberg, A.M. 1987. Toxicity of Chlorpyrifos (Dursban) in *Oreochromis aureus* and *O. niloticus* and Data on its Residues in *O. aureus*. **Bamidgeh** 39(1): 13-20.
- Hodge, S., Longley, M., Booth, L., Heppelthwaite V., and O'Halloran K. 2000. An Evaluation of glutathione -s-transferase activity in the Tasmanian lacewing

- (*Micromus tasmaniae*) as a biomarker of organophosphate contamination. **Bulletin of Environmental Contamination and Toxicology** 65: 8-15.
- Homola, E., and Chang, E. S. 1997. Distribution and regulation of methyl farnesoate esterase activity in *Homarus americanus* and other crustaceans. **General and Comparative Endocrinology**. 106: 62-72.
- Hook, S. E., and Lee, R. F. 2004. Genotoxicant induced DNA damage and repair in early and late developmental stages of the grass shrimp (*Paleomonetes pugio*) embryo as measured by the comet assay. **Aquatic Toxicology** 66: 1-14.
- Howard, A. S., Bucelli, R., Jett, D. A., Bruun, D. , Yang, D., and Lein, P. J. 2005. Chlorpyrifos exerts opposing effects on axonal and dendritic growth in primary neuronal cultures. **Toxicology and Applied Pharmacology** 207: 112– 124.
- Hughes, J. M., Griffiths, M. W., and Harrison, D. A. 1992. The Effects of an organophosphate insecticide on two enzyme loci in the shrimp *Caradina* sp. **Biochemical Systematics and Ecology**. 20(2): 89-97.
- Hulka, B. S., Wilcoxky, T. C., and Griffith, J. D. 1990. **Biological markers in epidemiology**. New York: Oxford University Press.
- Hyne, R. V., and Maher, W. A. 2003. Invertebrate biomarkers: links to toxicosis that predict population decline. **Ecotoxicology and Environmental Safety** 54(3): 366-374.
- Im, K. I., Park, K. M., Yong, T. S., Hong, Y. P., and Kim, T. E. 1999. Upregulated expression of the cDNA fragment possibly related to the virulence *Acanthamoeba culbertsoni*. **The Korean Journal of Parasitology** 37(4): 257-263.
- Ingelman-Sundberg, M. 2002. Polymorphism of cytochrome P450 and xenobiotic toxicity. **Toxicology** 181-182: 447-452.
- James, M. O., Boyle, S. M., Trapido-Rosenthal, H. G., Smith, W. C., Greenberg, R. M., and Shiverick, K. T. 1996. cDNA and protein sequence of a major form of P450 CYP2L, in the hepatopancreas of the spiny lobster, *Panulirus argus*. **Archives of Biochemistry and Biophysics** 329: 31-38.
- Jamil, K. 2001. **Bioindicators and Biomarkers of Environmental Pollution and Risk Assessment**. Enfield N. H. and Plymouth, Science Publishers, Inc., 204 pp.

- Jarvinen, A. W., Tanner, D. K., and Kline, E. R. 1988. Toxicity of chlorpyrifos, endrin, or fenvalerate to fathead minnows following episodic or continuous exposure. **Ecotoxicology and Environmental Safety** 15:78-95.
- Jasmin, B. J., Gisiger, V. 1990. Regulation by exercise of the pool of G4 acetylcholinesterase characterizing fast muscles: opposite effect of running training in antagonist muscles. **Journal of Neuroscience** 10: 1444-1454
- Kassie, F., Parazefal, L. W., and Knasmuller, S. 2000. Single cell gel electrophoresis assay: a new technique from human biomonitoring studies. **Mutation Research-Reviews in Mutation Research** 413: 33-38.
- Kawasaki, E. S., Innis, M. A., Gelfand, D. H., Sninsky, J. J., and White, T. J. 1990. Amplification of RNA. In: Innis MA (eds) **PCR protocols: A guide to methods and applications**. Academic Press, San Diego, pp 21-27
- Key, P. B., and Fulton, M. H. 2006. Correlation between 96-h mortality and 24-h acetylcholinesterase inhibition in three grass shrimp larval life stages. **Ecotoxicology and Environmental Safety** 63: 389-392.
- Kirby, M. F., Morris, S., Hurst, M., Kirby, S. J., Neall, P., Tylor, T., and Fagg, A. 2000. The use of cholinesterase activity in flounder (*Platichthys flesus*) muscle tissue as a biomarker of neurotoxic contamination in the UK estuaries. **Marine Pollution Bulletin** 40: 780-791.
- Kiskinis, E., Suter, W., and Hartmann, A. 2002. High throughput Comet assay using 96-well plates. **Mutagenesis** 17: 37-43.
- Klinbunga, S., Sodsuk, S., Penman, D. J., and McAndrew, B. J. 1996. An improved protocol for total DNA isolation and visualization of mtDNA RFLP(s) in tiger prawn, *Penaeus monodon*. **Thai. J Aquat Sci.** 3: 36-41.
- Kristensen, T., Edwards, T. M., Kohno, S., Baatrup, E., Guillette Jr, L. J. 2007. Fecundity, 17-estradiol concentrations and expression of vitellogenin and estrogen receptor genes throughout the ovarian cycle in female Eastern mosquitofish from three lakes in Florida. **Aquatic Toxicology** 81: 245-255.
- Kurelec, B., Britvic, S., Rijavec, M., Muller, W. E. G., and Zahn, R. K. 1977. Benzo(*a*)pyrene monooxygenase induction in marine fish-Molecular response to oil pollution. **Marine Biology** 44: 211-216.
- Laabs, V., Amelung, W., Pinto, A., Altstaedt, A. and Zech, W. 2000. Leaching and degradation of corn and soybean pesticides in an Oxisol of the Brazilian Cerrados. **Chemosphere** 41: 1441-1449.

- Ladaa, T., Bielmyer, G., and Murphy, K. L. 1998. EE&S 845 Environmental engineering chemistry II: Environmental organic chemistry spring 1998. EE&S Department, Clemson University .
- Lee, R. F., and Steinert, S. 2003. Use of the single cell gel electrophoresis/Comet assay for detecting DNA damage in aquatic (marine and freshwater) animals. **Mutation Research** 544(1): 43-64.
- Li, W. F., Costa, E. G., and Furlong, C. E. 1993. Serum paraoxonase status: A major factor in determining resistance to organophosphates. **Journal of Toxicology and Environmental Health.** 40: 337-346.
- Liang, P., and Pardee, A. B. 1992. Differential display of eukaryotic messenger RNA by means of the polymerase chain reaction. **Science** 14 (5072): 967-971.
- Limsuwan, C., and Chanrachchakul, P. 2004. **Shrimp aquaculture industry in Thailand.** National Research Council of Thailand. Thailand.
- Liu, B., McConnell, L. L., and Torrents, A. 2001. Hydrolysis of chlorpyrifos in natural waters of the Chesapeake Bay. **Chemosphere** 44(6): 1315-23.
- Liu, P., Ewis, H. E., Tai, P. C., Lu, C. D. and Crystal, I. T. W. 2007. Structure of the *Geobacillus stearothermophilus* Carboxylesterase Est55 and Its Activation of Prodrug CPT-11. **Journal of Molecular Biology** 367: 212-223.
- Lundebye, K. A., Langston, W. J., Depledge, M. H. 1997. Stress proteins and condition index as biomarkers of tributyltin exposure and effect in mussels. **Ecotoxicology** 6: 127-136.
- Macalady, D. L., and Wolf, N. L. 1985. Effects of sediment sorption and abiotic hydrolysis organophorothioate esters. **Journal of Agricultural and Food Chemistry.** 33: 167-173.
- Manisseril, M. K., and Menon, N. R. 1995. Copper-induced damage to the hepatopancreas of the penaeid shrimp *Metapenaeus dobsoni* - an ultrastructural study. **Diseases of Aquatic Organisms** 22: 51-57.
- Marone, M., Mozzetti, Si., Ritis, D. D., Pierelli, L., Scambia, G. 2001. Semiquantitative RT-PCR analysis to assess the expression levels of multiple transcripts from the same sample. **Biological Proceeding Online** 3(1): 19-25.
- Massoulie, J., Pezzementi, L., Bon, S., Krejci, E., and Vallette, F. M. 1993. Molecular and cellular biology of cholinesterases. **Progress in Neurobiology** 41: 31-91.

- Maxwell, D. M. 1992. The specificity of carboxylesterase protection against the toxicity of organophosphorus compounds. **Toxicology and Applied Pharmacology** 114: 306 -312.
- Mazanti, L., Rice ,C., Bialek, K., Sparling, D., Stevenson, C., Johnson, W. E., Kangas, P., and Rheinstein, J. 2003. Aqueous-Phase Disappearance of Atrazine, Metolachlor, and Chlorpyrifos in Laboratory Aquaria and Outdoor Macrocosms. **Archives of Environmental Contamination and Toxicology** 44: 67-76.
- McHenry, J. G., Saward, D., and Seaton, D. 1991. Lethal and sub-lethal effects of the salmon de-lousing agent dichlorovos on the larvae of the common lobster and herring. **Aquaculture** 98: 331-334.
- MedicineNet Inc. 2006. **Definition of Glutathione-S-transferase**. Available from: <http://www.medterms.com/script/main/art.asp?articlekey=31473>.
- Memorial Sloan-Kettering Cancer Center, 2006. **Heat-Shock Protein 90**. Available from: <http://www.mskcc.org/mskcc/html/10853.cfm>.
- Meshorer, E. et al. 2002. Alternative splicing and neuritic mRNA translocation under long-term neuronal hypersensitivity. **Science** 18;295(5554): 508-12.
- Mills, L. J., and Chichester, C., 2005. Review of evidence: are endocrine-disrupting chemicals in the aquatic environment impacting fish populations? **Science of the Total Environment** 343: 1-34.
- Mohan, S. V., Sirisha, K., Rao, N. C., Sarma, P. N., and Reddy, S. J. 2004. Degradation of chlorpyrifos contaminated soil by bioslurry reactor operated in sequencing batch mode: bioprocess monitoring. **Journal of Hazardous Materials** B116: 39-48.
- Montanes, J. F. C., Hattum, B. V., and Deneer, J. 1995. Bioconcentration of chlorpyrifos by the freshwater isopod *Asellus aquaticus* (L.) in outdoor experimental ditches. **Environmental Pollution** 88(2): 137-146.
- Moody, D. E. 2001. Genomics techniques: An overview of methods for the study of gene expression. **Journal of Animal Science** 79: E128-E135.
- Naravaneni, R., and Jamil, K. 2005. Evaluation of cytogenetic effects of lambda-cyhalothrin on human lymphocytes. **Journal of Biochemical and Molecular Toxicology** 19: 304-310.
- National Food Institute Information Center. 2004. **Shrimp production**. Available from: <http://www.nfi.or.th/stat/shrimp.asp>.

- Nayavon, R. 1996. Cabaryl and chlorpyrifos contamination in water and sediment of the golf course adjacent to Nong Klang Dong Reservoir, Changwat Chon Buri. **Diploma Thesis**. Interdepartment of Environmental Science. Chulalongkorn University. Thailand.
- Nazir, A., Mukhopadhyay, I., Saxena, D. K., Chowdhuri, D. K. 2001. Chlorpyrifos-Induced *hsp70* Expression and Effect on Reproductive Performance in Transgenic *Drosophila melanogaster* (*hsp70-lacZ*) Bg9. **Archives of Environmental Contamination and Toxicology**. 41: 443-449.
- NRC: Committee on Biological Markers of the National Research Council, 1987. Biological markers in environmental health research. **Environmental Health Perspectives** 74: 3-9.
- Office of agricultural economics, and the customs department. 2005. **Export value of black tiger shrimp, *Penaeus monodon* 2002-2004**. Thailand.
- Olima, C., Pablo, F. , and Lim, R. P. . 1997. Comparative tolerance of three populations of the freshwater shrimp (*Paratya australiensis*) to the organophosphate pesticide, chlorpyrifos. **Bulletin of Environment Contamination and Toxicology** 59: 321-328.
- Olive, P. L., Banáth, J. P., and Durand, R. E. 1990. Heterogeneity in radiation-induced DNA damage and repair in tumor and normal cells using the 'comet' assay. **Radiation Research** 122: 86-94.
- Olive, P. L., Frazer,G., and Banath, J. P. 1993. Radiation-induced apoptosis measured in TK6 human B lymphoblast cells using the Comet assay. **Radiation Research** 136: 130-136.
- Ostling, O., and Johanson, K. J. 1984. Micro-electrophoretic study of radiation-induced DNA damages in individual mammalian cells. **Biochemical and Biophysical Research Communications** 123: 291-298.
- Ozretic, B. and Krajnovic-Ozretic, M., 1992. Esterase heterogeneity in mussel *Mytilus galloprovincialis*: effects of organophosphate and carbamate pesticides in vitro. **Comparative Biochemistry and Physiology C** 103: 221-225.
- Patnaik, K. K., and Tripathy, N. K. 1992. Farm-grade chlorpyrifos (Durmet) is genotoxic in somatic and germ-line cells of *Drosophila*. **Mutation Research-Genetic Toxicology and Environmental Mutagenesis** 279: 15-20.
- Payne, J. F., Mathieu, A., Melvin, W., Fancey, L. L. 1996. Acetylcholinesterase, an old biomarker with a new future? Field trials in association with two urban

- rivers and a paper mill in Newfoundland. **Marine Pollution Bulletin** 32: 225-231.
- Perrier, N. A., Salani, M., Falasca, C., Bon, S., Augusti-Tocco, G., and Massoulie, J. 2005. The readthrough variant of acetylcholinesterase remains very minor after heat shock, organophosphate inhibition and stress, in cell culture and in vivo. **Journal of Neurochemistry** 94: 629-638.
- Phipps, G. L., and Holcombe, G. W. 1985. A Method for Aquatic Multiple Species Toxicant testing: Acute toxicity of 10 chemicals to 5 vertebrates and 2 invertebrates. **Environmental Pollution Series A, Ecological and Biological** 38(2):141-157.
- Piña-Guzmán, B., Solís-Heredia, M. J., and Quintanilla-Vega, B. 2005. Diazinon alters sperm chromatin structure in mice by phosphorylating nuclear protamines. **Toxicology and applied Pharmacology** 202: 189-198.
- Pittendrigh, B., Aronstein, K., Zinkovsky, E., Andreev, O., Campbell, B., Daly, J., Trowell, S., and French-Constant, R. H. 1997. Cytochrome P450 genes from *Helicoverpa armigera*: expression in a pyrethroid-susceptible and -resistant strain. **Insect Biochemistry and Molecular Biology** 27: 507-512.
- Poet, T. S., Wu, H., Kousba, A. A., and Timchalk, C. 2003. *In Vitro* Rat Hepatic and Intestinal Metabolism of the Organophosphate Pesticides Chlorpyrifos and Diazinon. **Toxicological Sciences** 72: 193-200.
- Prabhavathy, D., G., Pasha, S., A., Jamil, K. 2006. Cytotoxicity and Genotoxicity Induced by the Pesticide Profenofos on Cultured Human Peripheral Blood Lymphocytes. **Drug and Chemical Toxicology** 29: 313-322.
- Quintana, P. J., de Peyster, A., Klatzke, S., and Park, H. J. 2000. Gossypol-induced DNA breaks in rat lymphocytes are secondary to cytotoxicity. **Toxicology Letters** 117: 85-94.
- Radhakrishnaiah, K., Sivaramakrishna, B., Suresh, A., and Chamundeswari, P. 1995. Pesticidal impact on the protein metabolism of freshwater field crab, *Oziotelphusa senex senex* (Fabricius). **Biomedical and Environmental Sciences** 8(2): 137-148.
- Rahman, M. F., Mahboob, M., Danadevi, K., Banu, B. S., and Grover, P. 2002. Assessment of genotoxic effects of chloropyriphos and acephate by the comet assay in mice leucocytes. **Mutation Research** 516:139-147.

- Rao, J. V., Pavan, Y. S., and Madhavendrab, S. S. 2003. Toxic effects of chlorpyrifos on morphology and acetylcholinesterase activity in the earthworm, *Eisenia foetida*. **Ecotoxicology and Environmental Safety** 54: 296-30.
- Reddy, M. S., and Rao, K. V. R. 1988. Modulation of carbohydrate metabolism in the selected tissues of marine prawn, *Penaeus indicus* under phosphamidon induced stress. **Ecotoxicology and Environmental Safety** 15: 212-220.
- Rewitz, K., Styrihave, B., and Andersen, O. 2003. *CYP330A1* and *CYP4C39* enzymes in the shore crab *Carcinus maenas*: sequence and expression regulation by ecdysteroids and xenobiotics. **Biochemical and Biophysical Research Communications** 310: 252-260.
- Reyes, J. G. G., Leyva, N. R., Millan, O. A., and Lazcano, G. A. 2002. Effects of pesticides on DNA and protein of shrimp larvae *Litopenaeus stylirostris* of the California Gulf. **Ecotoxicology and Environmental Safety** 53: 191-195.
- Roast, S. D., Thompson, R. S., Donkin, P., Widdows, J., and Jones, M. B. 1999. Toxicity of the organophosphate pesticides chlorpyrifos and dimethoate to *Neomysis integer* (Crustacea: Mysidacea). **Water Research** 33: 319-326.
- Roser, S., Pool-Zobel, B. L., and Rechkemmer, G. 2001. Contribution of apoptosis to responses in the comet assay. **Mutation Research** 497: 169:175.
- Saleha, B. B., K., Danadevi, Rahman, M. F., Ahuja, Y. R., and Kaiser, J. 2001. Genotoxic effect of monocrotophos to sentinel species using comet assay. **Food and Chemical Toxicology** 39: 361-366.
- Sarkar, A., Ray, D., Shrivastava, A. N., and Sarkar, S. 2006. Molecular Biomarkers: Their significance and application in marine pollution monitoring. **Ecotoxicology** 15: 333-340.
- Scaps, P., Demuynck, S., Descamps, M., and Dhainaut, A. 1997. Effects of organophosphate and carbamate pesticides on acetylcholinesterase and cholinesterase activities of polychaete *Nereis diversicolor*. **Archives of Environmental Contamination and Toxicology** 33: 203-208.
- Scharf, M. E., Neal, J. J., and Bennett, G. W. 1998. Changes of insecticide resistance levels and detoxication enzymes following insecticide selection in the German cockroach, *Blattella germanica* (L.). **Pesticidebiochemistry and Physiology** 59: 67-79.
- Scott, J., and Redmond, M. S. 1986. **Acute toxicity tests with chloropyrifos.** SAIC, Narragansett R I:3.

- Siefert, R. E. 1987. **Effects of dursban (chlorpyrifos) on aquatic organisms in enclosures in a natural pond - Final report.** U.S.EPA, Duluth, MN :214.
- Singh, N. P. et al. 1988. A simple technique for quantitation of low levels of DNA damage in individual cells. **Experimental Cell Research** 175: 184-91.
- Sithisarankul, P. 2002. **Molecular epidemiology.** Bangkok, Thailand.
- Sobti, R. C., Krishan, A., and Pfaffenberger, C. D. 1982. Cytokinetic and cytogenetic effects of some agricultural chemicals on human lymphoid cells in vitro: organophosphates. **Mutation Research-Genetic Toxicology** 102: 89-102.
- Steevens, J. A., and Benson, W. H. 1999. Toxicological Interaction of chlorpyrifos and Methyl mercury in the Amphipod, *Hyalella azteca*. **Toxicological Science** 52: 168-177.
- Stien, X., Percic, P., Gnassia-Barelli, M., Roméo, M., and Lafaurie, M. 1998. Evaluation of different biomarkers in caged fish and mussels to assess the quality of waters in the Bay of Cannes (Côte d'Azur), S.E. France. **Environmental Pollution** 99: 339-345.
- Sturm, A., Radau, T. S., Hahn, T., and Schulz, R. 2007. Inhibition of rainbow trout acetylcholinesterase by aqueous and suspended particle-associated organophosphorous insecticides. **Chemosphere** 68: 605-612.
- The customs department, 2006. **Import/Export Statistics.** Available from: <http://www.customs.go.th/Statistic/StatisticIndex.jsp>.
- Tice, R. R., Agurell, E., Anderson, D., Burlinson, B., Hartmann, A., Kobayashi, H., Miyamae, Y., Rojas, E., Ryu, J. C., and Sasaki, Y. F. 2000. Single cell gel/Comet assay: guideline for in vitro and in vivo genetic toxicology testing. **Environmental and molecular mutagenesis** 35: 206-221366.
- Tilak, K. S., Veeraiah, K., and Rao, D. K. 2004. Toxicity and bioaccumulation of chlorpyrifos in Indian carp *Catla catla* (Hamilton), *Labeo rohita* (Hamilton), and *Cirrhinus mrigala* (Hamilton). **Bulletin of Environmental Contamination and Toxicology** 73: 933-41.
- Timchalk, C., Poet, T. S., and Kousba, A. A. 2006. Age-dependent pharmacokinetic and pharmacodynamic response in preweanling rats following oral exposure to the organophosphorus insecticide chlorpyrifos . **Toxicology** 220: 13-25.
- Toxic Substance Information Center. 2002. **Agricultural hazardous substance registered for production.** Department of Agriculture, Bangkok. 444 pp.
- US EPA. 1986. **Quality Criteria for Water.** EPA-440/5-86/001.

- US EPA. 2002a. **Interim reregistration eligibility decision for chlorpyrifos.** Available from: [http://www.epa.gov/oppsrrd1/REDs/chlorpyrifos\\_ired.pdf](http://www.epa.gov/oppsrrd1/REDs/chlorpyrifos_ired.pdf).
- US EPA. 2002b. **Reregistration Eligibility Science Chapter for Chlorpyrifos Fate and Environmental Risk Assessment Chapter.** Available from: <http://www.epa.gov/oppsrrd1/op/chlorpyrifos/efedassmnt1of3.pdf>.
- Varo', I., Navarro, J. C., Amat, F., and Guilhermino, L. 2002. Characterisation of cholinesterases and evaluation of the inhibitory potential of chlorpyrifos and dichlorvos to *Artemia salina* and *Artemia parthenogenetica*. **Chemosphere** 48: 563-569.
- Vaz, I. et al. 2004. Effect of acaricides on the activity of a *Boophilus microplus* glutathione-S-transferase. **Veterinary Parasitology** 119: 237-245.
- Vioque-Fernandez, A. et al. 2007. Doñana National Park survey using crayfish (*Procambarus clarkii*) as bioindicator: Esterase inhibition and pollutant levels. **Toxicology Letters** 168(3): 260-268.
- Walker, C. H. 1998. The Use of Biomarkers to Measure the Interactive Effects of Chemicals. **Ecotoxicology and Environmental Safety** 40: 65-70.
- Welsh, J., Chada, K., Dalal, S. S., Cheng, R., Ralph, D., and McClelland, M. 1992. **Nucleic Acid Research** 20: 4965-4970.
- Werck-Reichhart, D., and Feyereisen, R. 2000. Cytochromes P450: a success story. **Genome Biology** 1(6): 3003.1-3003.9.
- Wheelock, C. E., Eder, K. J., Werner, I., Huang, H., Jones, P. D., Brammell, B. F., Elskus, A. A., and Hammock, B. D. 2005. Individual variability in esterase activity and CYP1A levels in Chinook salmon (*Oncorhynchus tshawytscha*) exposed to esfenvalerate and chlorpyrifos. **Aquatic Toxicology** 74: 172-192.
- WHO International Programme on Chemical Safety (IPCS). 1993. Biomarkers and risk assessment: concepts and principles. **Environmental Health Criteria** 155. World Health Organization, Geneva.
- Wild, D., 1975. Mutagenicity studies on organophosphorus insecticides. **Mutation Research-Reviews in Genetic Toxicology** 32: 133-150.
- Williams, C. 1969. Glucuronidase activity in the serum and liver of rats administered pesticides and hepatotoxic agents. **Toxicology and Applied Pharmacology** 14:283-292.
- Wilson, J. A., and Clark, A. G. 1996. The role of E3 Esterase, glutathione-s-transferases and other nonoxidative mechanisms in resistance to diazinon and

- other organophosphate insecticides in *Lucilia cuprina*. **Pesticide Biochemistry and Physiology** 54: 85-95.
- Woodruff, R. C., Phillips, J. P., and Irwin, D. 1983. Pesticide-induced complete and partial chromosome loss in screens with repair-defective females of *Drosophila melanogaster*. **Environmental Mutagenesis** 5: 835-846.
- Zhu, Y. C., Snodgrass, G. L., and Chen, M. S. 2007. Comparative study on glutathione S-transferase activity, cDNA, and gene expression between malathion susceptible and resistant strains of the tarnished plant bug, *Lygus lineolaris*. **Pesticide Biochemistry and Physiology** 87: 62-72.
- Zucker, E. 1985. **Hazard evaluation division-Standard evaluation procedure- Acute toxicity test for fresh water fish.** U.S. EPA. Publication 540/9-85-006.

## **APPENDICES**

## APPENDIX A

### 1. LB Broth (per Liter)

10 g of NaCl

10 g of tryptone

5 g of yeast extract

Add deionized H<sub>2</sub>O to a final volume of 1 liter. Adjust to pH 7.0 with 5 N NaOH and autoclave.

### 2. LB Agar (per Liter)

- 10 g of NaCl

- 10 g of tryptone

- 5 g of yeast extract

- 20 g of agar

Add deionized H<sub>2</sub>O to a final volume of 1 liter. Adjust to pH 7.0 with 5 N NaOH and autoclave. After, pour into petri dishes (~25 ml/100-mm plate)

### 3. LB-Ampicillin Agar (per Liter)

- Prepare 1 liter of LB agar. Autoclave and cool to 55 °C

- Add 50 ml of filter-sterilized ampicillin

- Pour into petri dishes (~25 ml/100-mm plate)

### 4. 1x TAE Buffer

- 40 mM Tris-acetate

- 1 mM EDTA

**5. SOB Medium (Per liter) :**

- Bacto-tryptone 20 g
- Yeast extract 5 g
- NaCl 0.5 g

**6. Ampicillin**

Stock solution. 25 mg/ml of the sodium salt of ampicillin in water. Sterilize by filtration and store in aliquots at -20 °C

**7. 5 M NaCl**

Dissolve 292.2 g of NaCl in 800 ml of H<sub>2</sub>O. Adjust volume to 1 liter. Dispense into aliquots and sterilize by autoclaving.

**8. 1 M MgCl<sub>2</sub>**

Dissolve 203.3 g of MgCl<sub>2</sub> · 6H<sub>2</sub>O in 800 ml of H<sub>2</sub>O. Adjust volume to 1 liter. Dispense into aliquots and sterilize by autoclaving.

**9. 3 M Sodium acetate (pH 5.2)**

Dissolve 408.1 g of sodium acetate · 3H<sub>2</sub>O in 800 ml of H<sub>2</sub>O. Adjust pH to 5.2 with glacial acetic acid. Adjust volume to 1 liter. Dispense into aliquots and sterilize by autoclaving.

**10. 10% Sodium dodecyl sulfate (SDS) (also called sodium lauryl sulfate)**

Dissolve 100 g of electrophoresis-grade SDS in 900 ml of H<sub>2</sub>O. Heat to 68 °C to assist dissolution. Adjust the pH to 7.2 by adding a few drops of concentrated HCl. Adjust volume to 1 liter. Dispense into aliquots.

**11. Ethidium bromide 10 mg/ml**

Add 1 g of ethidium bromide to 100 ml of H<sub>2</sub>O. Stir on a magnetic stirrer for several hours to ensure that the dye has dissolved. Wrap the container in aluminum foil or transfer to a dark bottle and store at 4 °C.

**12. TE pH 7.4**

- 10 mM Tris · Cl (pH 7.4)

- 1 mM EDTA (pH 8.0)

**13. TE pH 7.6**

- 10 mM Tris · Cl (pH 7.6)

- 1 mM EDTA (pH 8.0)

**14. TE pH 8.0**

- 10 mM Tris · Cl (pH 8.0)

- 1 mM EDTA (pH 8.0)

**15. Tris-Borate (TBE)**

- Working solution

- 0.089 M Tris-borate

- 0.089 M boric acid

- 0.002 M EDTA

- Concentrated stock solution (5x)

Per liter:

- Tris base 54 g

- Boric acid 27.5 g

- 0.5 M EDTA (pH 8.0) 20 ml

**16. Gel-Loading Buffer Type II**

- 10x buffer

- 0.25% bromophenol blue
- 0.25% xylene cyanol
- 25% Ficoll (type 400) in H<sub>2</sub>O
- Store at room temperature.

### **17. 10x TEN buffer**

- 0.1 M Tris-Cl (pH 8.0)
- 0.01 M EDTA (pH 8.0)
- 0.1 M NaCl

### **18. Equilibration of Phenol**

Before use, phenol must be equilibrated to a pH of > 7.8 because the DNA partitions into the organic phase at acid pH. Wear gloves, full face protection, and a lab coat when carrying out this procedure.

1. Store liquefied phenol at -20°C. As needed, remove the phenol from the freezer, allow it to warm to room temperature, and then melt it at 68°C. Add hydroxyquinoline to a final concentration of 0.1 %. This compound is an antioxidant, a partial inhibitor of RNase, and a weak chelator of metal ions (Kirby 1956). In addition, its yellow color provides a convenient way to identify the organic phase.
2. To the melted phenol, add an equal volume of buffer (usually 0.5 M Tris-Cl [pH 8.0] at room temperature). Stir the mixture on a magnetic stirrer for 15 minutes. Turn off the stirrer, and when the phases have separated, aspirate as much as possible of the upper (aqueous) phase using a glass pipette attached to a vacuum line equipped with appropriate traps (please see Appendix 8, Figure A8-2).
3. Add an equal volume of 0.1 M Tris-Cl (pH 8.0) to the phenol. Stir the mixture on a magnetic stirrer for 15 minutes. Turn off the stirrer and remove

the upper aqueous phase as described in Step 2. Repeat the extractions the pH of the phenolic phase is >7.8 (as measured with pH paper)

4. After the phenol is equilibrated and the final aqueous phase has been removed, add 0.1 volume of 0.1 M Tris-Cl (pH 8.0) containing 0.2%  $\beta$ -mercaptoethanol<!>. The phenol solution may be stored in this form under 100 mM Tris-Cl (pH 8.0) in a light-tight bottle at 4°C for periods of up to 1 month.

#### **19. Phenol: Chloroform: Isoamyl Alcohol (25:24:1)**

A mixture consisting of equal parts of equilibrated phenol and chloroform: isoamyl alcohol<!> (24:1) is frequently used to remove proteins from preparation of nucleic acids. The chloroform denatures protein and facilitates the separation of the aqueous and organic phases, and the isoamyl alcohol reduces foaming during extraction. Neither chloroform nor isoamyl alcohol requires treatment before use. The phenol: isoamyl alcohol mixture may be stored under 100 mM Tris-Cl (pH 8.0) in a light-tight bottle at 4 °C for period of up to 1 month.

#### **20. Glycerol (10% v/v)**

Dilute 1 volume of molecular-biology-grade glycerol in 9 volume of sterile pure H<sub>2</sub>O. Sterilize the solution by passing it through a prerinse 0.22 uM filter. Store in 200-ml aliquots at 4 °C

#### **21. IPTG (20% w/v, 0.8 M)**

IPTG is isopropylthio-B-D-galactoside. Make a 20% solution of IPTG by dissolving 2 g of IPTG in 8 ml of distilled H<sub>2</sub>O. Adjust the volume of the solution to 10 ml with H<sub>2</sub>O and sterilize by passing it through a 0.22 uM disposable filter. Dispense the solution into 1-ml aliquots and store them at -20 °C

#### **22. X-gal solution (2% w/v)**

X-gal is 5-bromo-4-chloro-3-indolyl-B-d-galactoside. Make a stock solution by dissolving X-gal in dimethylformamide at a concentration of 20 mg/ml solution. Use a glass or polypropylene tube. Wrap the tube containing the solution in aluminum

foil to prevent damage by light and store at -20<sup>0</sup>C .It is not necessary to sterilize X-gal solution by filtration.

## APPENDIX B

**Table B1** Summary of partial gene sequences from RT-PCR using hepatopancreas first strand cDNA as template

Gene	Primer	Nucleotide Sequence
1. Cytochrome P450	F1R2	<u>TTCATGTTCGAAGGCCACGACACCACCGCGGCCATGA</u> ACTGGGTTCTTACCTCCCTGGGTCACTACCCAGAGATA <u>CAGGCTCGGT</u> CACCCAGGAGCTGG ACTCGATCTTCGGTGACGAGGACCGCCCGACGATGGACGACCTGCGCTCCATGAAGCTGCTGGAGAA <u>CTGCATCAAGGAGGGCTGAGG</u> CTATTCCCGTC CGTCCACAGGTTGCCAGGACGCTGCGAGAA <u>GACGTCCG</u> CATATGC <u>GACTACGT</u> CATCCCGCCGAACCAACATCATGCTTCGTACCGAATCCACCGC GACCCGAAGCAGTCCCCGACCCGGAGAGGTTCGACCCGGACC <u>GCTTCTGCCGAGAA</u> ACAGCAAACACCGC <u>CATCCTACCGT</u> TATGTTCC
2. Beta glucuronidase	F1R1	<u>GGTAAATCGCTACTATTCCTGGT</u> ACAGTGACACAGGGACCTAGAGCTGATTATAATCAGACTGTGAATGAGTTATTGAGTGGCATCTCCACAAACAG CCCGTTATGATCTCAGAGTATGGAGCTGGTCCATTGCTGGTTCCACATGGACCC <u>CTGCATT</u> CACATGGACT <u>GAAGAATACCAAGCAGAGTTG</u>
3. Glutathione-s-transferase	F1R1	<u>AGTCGGTAATGCTAACGCCAAGGCAGTGGC</u> ATCGAAC <u>CTGAAGCTGCTTAACCTGCCGGGGAGAGCATATGAAGCCGAGTTCGTGGCCATCA</u> ACCC <u>TCAGC</u> ACTGCATCCCCACCTGGTCAGGGAAC <u>CTGAAGCTG</u> TTGGAGAGGCCGCCAT <u>CTGCACCTAC</u> CTGATGCCAA <u>ATACCCGAGGACACTC</u> <u>ACTCTACCCGTCCGACCCG</u>

Remark: Underlined letter indicates location of primer

**Table B2** Summary of 5' and 3' nucleotide sequences from RACE PCR

Gene	RACE PCR Product	Nucleotide Sequence
1. Carboxylesterase	5'	ACGCGGGGGGGGAGTCGTGGCTGCGCTCCGGAGCGGAAAGATGGTAGTCGAAGTACGCAATGAAGCTGGGTCCTTCTCTGACGGCTG GATGGCAACACTGTCGAAGGCCAGCAGCAGGAACCA <u>CGACATCCACAGGCC</u> TTCGATTGAGGTGGCTCCGGCAGGGCTGATCACAGGGCC CAGTCAGAGGCCGAAACGGTAGGGTCTTCA <u>AGCTTCAAGACC</u> ATTCCCTCGCCAGGCTCTGAGGACTAAGGTTAGGGACCCGTTC CTGCAAGGCCATGGCAGGAGTAAGAA <u>ATGGATCCATGCCACACCGAA</u> ATGCCACAGTGGAA <u>ATGCTACTGTTGAGGGCAGGAAGACTGTCT</u> CTATCTCCGT <u>TACACACCTCGCC</u> TTACCGCTGGACTTGCCTGT <u>CATGGTGTGATT</u> ACGGGGAGGATTACGAACGGTCAAGGGAGGTC TTCGGGCCCCCTCTCTCAGAAGGATGTGGTCTCTGTT <u>CATACAGTATCGCTGGCCACGCTGGGATTCTTATCGACTGAGGACAATGAGC</u> TGCCTGGCAATCTAGGACT <u>CAAGGACCAAGGATGGCT</u> CCCTGTGGGT <u>CAAGACAACATCCGT</u> GACCT
	3'	<u>ATTTCTACCGACTACCCGCCGTCTG</u> TGAGAAGAGGACGGTATAAC <u>AAGGTGGACATTAT</u> ATCTGGGATTACGCAAGATGAGGCAGCCGTGAT TGGCTGATTT <u>TCACCTGG</u> ACAAAGCCGCTGCAACAGCCTGGTCCAGAA <u>CTTCTGT</u> CAACGGAC <u>AGTTC</u> ACTGAT <u>CTCGAAGCTGGAA</u> GATGACCCGGAGTAC <u>CTGGCACCCGAGC</u> CTCCACACTAC <u>CTGGG</u> CCATTGAAGT <u>GACAGAAGAGAAACGGGATT</u> ACTTACGGCTTTCA GTGATAGAATGTCGACATGTGTC <u>ATCTTGACGCGGT</u> CGGGCAAC <u>ACCTCAGAACATCTCATCAGAAGCT</u> GTGTTACATCAA <u>ACTG</u> CAGCATGACGG AGAACACCAGTTTTGGTTTTCC <u>ACTACCCG</u> ATTGGTACAA <u>AGCTATGCGGT</u> ACGGCAG <u>ATGATTTG</u> TACCTGGTACCGTCAA GCTGAGGGCAACAGAAC <u>GTGAGCGAGACGAGG</u> ATCTGTC <u>GGTAGCC</u> TAT <u>GGTAGAGCTG</u> GGACCA <u>ACTTCGCTGGACACCCGA</u> CGCCTGACAT <u>GTCCCTCGG</u> CTCAATGGAA <u>ACCCGACGT</u> CTTCCC <u>AAACAGACTCCTACTGTCC</u> AT <u>ACCTCCTCACCCACCATG</u> AAA <u>ACCTCGA</u> AGACT <u>TCGAGACCGT</u> GAATT <u>CTGGAAGAACATG</u> CT <u>ACCAAGA</u> AA <u>ACAAATG</u> CTGAC <u>CCCTGAGCG</u> TT <u>ACAAGTGT</u> ACT <u>ACCTGGCT</u> GC

Remark: Underlined letter indicates location of primer

**Table B3** Summary of partial gene sequences from mRNA DDRT-PCR using A anchored oligodT first strand cDNA as template

Product	Nucleotide Sequence
OPA07A-350-7-1	<b>GAAACGGGTGTTGGTGTGCCCTCAGCGGAACATCCGCCACGTGCTGATGTC</b> TAACCTCACGCACCCAAAGAGGTATA <b>CAGCTGGACTTGAGAACAGGTACGTGACTGGACTGACGTTAACCGGTCGACCTTGGAGCGAGCTAGCCTGGATGGCTAGTC</b> ATCGCCAGGATATA <b>CATAGCGGATGCTCATACCAACGAGGCTTCCATGATTG</b> <b>CTTGGAGCGAGCTAGCCTGGATGGGTAGGTGCAACAGTCCATCTCGGAGGGCGTGAGGTGGCCAATGGACTGTGGATAGTC</b> GTCCAGCGCGAGGATATA <b>CATAGCGGATGCTCATACCAACGAGGCTTCCATGATTG</b>
OPA18A-500-19-1	<b>AGGTGACCGTGACTTCGATTGCCATGACCGATCTGATGAGGCTA</b> CTGCTTCTGACAGTAAGTGAAGCATAATTGTTCTGGCTGATATAACGTAACATTTCGATTATATCTGTTCCATGATTG CCTCAAACACGAGGAAACGATCCTAGTATGATTAATATAAGTCAGGCGATTCAATAGACTTCAGGAATTACAATACTTTAGGTTAAATCATTCAACATCGTAGCTGAATAATAGTTCTAGAAATACC AAATCACATTAACTGTATAATGACCTCAAGTATAATAACCCCTACACTCTGAAACGACACTGAGGTCAAATCTCATCCTGAAACTAAAACCAGAAAACACACTTAATACGACTGAAAGTTTTTTT ATATCACATGTGACCTTTTCAGCACCTGTGACCTTTCTGATATAAGACCTTTTATAGCACACGCGAGATTTCAGCACATGTGACTTTTCAGCACATGTGACTTTTCAGCACATGTGTTCTTACCATCGCTG <b>CGGAGACGGTCACCT</b>
OPA18A-550-18-1	<b>AGGTGACCGTGAGCAGTTCTTAGTCAGCCTCCACTGTATGCTCTGCCACACCAGTCTGTAACAGGAAGTGTCTCGCATCCATGATATCAGTTGAAACATAACCATCCAGATTCA</b> CAGATCGAGTTAC ATCTCCTCAGTTTATCAGTATCAAGTGCTGCCCTAAGCAAAAGACTGGAATGGGACTCTGGGCTGACGTTGGCTATGCAGGCAACACACCAGAACAAATGACAGCATCCTTGAGTACTCTAGAGGAGAT TGCTATTGAAACTATGCATCTGTGTTAAGAACAGAACCTGAAGGGACAACCCAAAGTGAAGAAAAAAACTAAAGAATTCTAAGAAGAATTACAAAGCAAGTGGAGCAAAGGTGGTGGAAAGCCTATGAGTGT TAAGCGAACATTCTGAGGTTGTGGGTCATTGCTACTCGATCACTTAGAGTCCCACAGATTGCTGAAAGAAAATTGTCTTCTAGTGTGATGAGGAATTGACAGCAGACGGACAAGTCCCTCTCAAAG <b>TCCAAGACGGAAGAACAGCGTCACCT</b>
OPA18A-550-18-2	<b>AGGTGACCGTCTACCTCGGCCTGCGCTTGCACTCGCTCAAACCTCGAGAAAATTACGCTGAATAATGTCAGTCTCAGCGATACAGTCGACAGTATCAAGCAGACAGCAGCTAGCAAGGGCAAGACGTCGGCGG</b> ATAAAATCGAACTAGTCCACTGTGGTGTCAATTCTGAATGGATCGAGACGCTGGAAGAACAGCAGGAATCACCTCGGTGCTATGATTCACTGTCTCCAAAAAAAGAGGATCCTGTAACTGTGGCGTCCCAT TAAGCCTCCCTGAAGTCAGACATTACTGGTGTCTGAGAAATTGTCGGCTCACCCAAATTCCGTGCTTTTATACAAAATCTGCAACCCCAAGAGATGGTGGAGAACGGTGTCTCATCGTGTCCGATTG CTCGCAGTGATCCATCAGCTCTCCATGCTCAAGACCCCTGAGCTGTTTATGGGACTCACTAATCCAGATAACTTAAGCAAGGTCGTAGATAGTAATCCTGCACTGGGAAGGGTGGCAGAGTATGTGACAT <b>CCCTCATGACGTCGGACACCACCTCACCTGACGGTCACCT</b>
OPA18A-600-17-1	<b>AGGTGACCGTCTACCTCGGCCTGCGCTTGCACTCGCTCAAACCTCGAGAAAATTACGCTGAATAATGTCAGTCTCAGCGATACAGTCGACAGTATCAAGCAGACAGCAGCTAGCAAGGGCAAGACGTCGGCGG</b> ATAAAATCGAACTAGTCCACTGTGGTGTCAATTCTGAATGGATCGAGACGCTGGAAGAACAGCAGGAATCACCTCGGTGCTATGATTCACTGTCTCCAAAAAAAGAGGATCCTGTAACTGTGGCGTCCCAT TAAGCCTCCCTGAAGTCAGACATTACTGGTGTCTGAGAAATTGTCGGCTCACCCAAATTCCGTGCTTTTATACAAAATCTGCAACCCCAAGAGATGGTGGAGAACGGTGTCTCATCGTGTCCGATTG CTCGCAGTGATCCATCAGCTCTCCATGCTCAAGACCCCTGAGCTGTTTATGGGACTCACTAATCCAGATAACTTAAGCAAGGTCGTAGATAGTAATCCTGCACTGGGAAGGGTGGCAGAGTATGTGACAT <b>CCCTCATGACGTCGGACACCACCTCACCTGACGGTCACCT</b>
OPA18A-600-17-3	<b>AGGTGACCGTGAGCAGTTCTTAGTCAGCCTCCACTGTATGCTCTGCCACACCAGTCTGTAACAGGAAGTGTCTCGCATCCATGATATCAGTTGAAACATAACCATCCAGATTCA</b> CAGATCGAGTTAC ATCTCCTCAGTTTATCAGTATCAAGTGCTGCCCTAAGCAAAAGACTGGAATGGGACTCTGGGCTGACGTTGGCTATGCAGGCAACACACCAGAACAAATGACAGCATCCTTGAGTACTCTAGAGGAGAT TGCTATTGAAACTATGCATCTGTGTTAAGAACAGAACCTGAAGGGACAACCCAAAGTGAAGAAAAAAACTAAAGAATTCTAAGAAGAATTACAAAGCAAGTGGAGCAAAGGTGGTGGAAAGCCTATGAGTGT TAAGCGAACATTCTGAGGTTGTGGGTCATTGCTACTCGATCACTTAGAGTCCCACAGATTGCTGAAAGAAAATTGTCTTCTAGTGTGATGAGGAATTGACAGCAGACGGACAAGTCCCTCTCAAAG <b>TCCAAGACGGAAGAACAGCGTCACCT</b>
OPA18A-650-16-3	<b>AGGTGACCGTCCTCGCAGAACGGTGGCGAGGTGGAATTCA</b> GGAGTCAGTGTGAAATTGACAACGGCTTGTGGCATTGATGACGTCATTCTGAGTCCCGACGCCGTGCCCCGATCTGTTCCGTGACT TCGAAGATGGCAGTTGCTGTGGCTGAACCTCGGGGAAATAGTGGATCACGGGCCCGTGACGGACGCCAGCTCTCTGCCCTGGGTTCGCTCCGCCGTACGCCACAGCAGGGACTCCAT ATGGCCACTACCTTACTCTACGATGCCCGGACTTGCCTGCAAGCGGTACGCCATCGTCTCTGAGGCTTCAGCGTCAACGGCAGTCGTTGTGCTTGGATCCATATGTCGGTGTATA AGGTTGGTCACTCACCTCAAGACCATACAAGAACGACCGGGTACCCCTGCTCTTAATCACTAGAGGGCTCACAAGGACTTCGCTGGAAAAACATACGGGCGAATTAGCAGTCACCCGACCACGGCTAT CTTCACGGCAGAGGGCGTTGCTGGCCACACCAACGTTGTGGCCATAGATGACATTCACTCTATGCAAGGACCTGTGGATAATTCT <b>ACGGTCACCT</b>



Product	Nucleotide Sequence
	<u>CCAAAT</u>

Remark: Bald letter indicates primer location

**Table B4** Summary of partial gene sequences from mRNA DDRT-PCR using C anchored oligodT first strand cDNA as template

Remark: Bold letter indicates primer location

**Table B5** Summary of partial gene sequences from mRNA DDRT-PCR using G anchored oligodT first strand cDNA as template

Product	Nucleotide Sequence
OPA01G 415 1	<u>CAGGCCCTCTGTATCTGCTAGCTTCCCTCCAAAGCCTTAGGCCTACCATTTGTCAGCCACTGCACTGCACTAACAGGGACTGCAGCAAGACCAGGACAAGGAAGAAATCTTATGTCAGCTATGTAAT</u> GCTCTGTGTA <u>ACTGGAACTCGTGAGATGACATCTTGGAACTGGCAACAGAATGGCTGAATGAAGGTTTAGAGCTGGTACTACAAAGTAGTAAGGAACGCAGAAAAGCCGAAAGGTGCGATTCCAAGAGGCACTTACACCGAACCTCTGGCTACGTCTGGCTATCCATAACATCCATTAAATAACCTGCTGCCCTGGGAGAACAGACATTTTAGTACAATGTCAGAGAACATGGCTGAA <u>GTGAAGGCCCTG</u></u>
OPA01G 415 3	GATTCCCCCCTCCCTCCAAAAAAAAAAAAAAATATGTAAGAATGATGTTATCACTTTTTAAGACAAAAAAAAAGAAGAAAAGAAAAAGAACGGACGCCAGCATTAA CCAACCAATCATAAAAGAAGAATAGGAATAAGGATTAAGAAATGAGAAAGAGAACGATATAGAAAAAGACAGGGAGATAGAGAGATAGAGATAGAGTAAAGGTATAAAAAAAATCATCCC TATAATACATTTCCAGAGTGAATATTCACTCCGTTCCAAAACGAGCGTTAAAACCTCTAAAAACTGCCGGGAAACAAAGTTACAAAATGCCATAAACACGCAATCATACAGTTACTGCTCGGA <u>AATATGAGGGAAAGGCCCTG</u>
OPA01G 600 3	<u>CAGGCCCTCAGAAAGTACCCGTGGCCCCCTGGGGACCATGACCTCCAGGGACAGTGGAAATGGAAAATAGCAAAACATAATAACAGAGATAAAGGAGAATGAATAAATACGACATTAAATGGT</u> TCTTATAATGATAAGAACATTAAACAAGCATGATGCTAAAGAACAGTAACGTGTTAAGATGAGTATAATATAATGATGATGATAATATTAGTAATAATATAATCTACTGATCCATGTGCCATGCCAACCC ACCCATAAACATTCTACTATCATGGCTGATAAGGCAATTCAAAGAAGATGGATTAGTGGAAACATTAGTAATGAGCAGAACATTGGTTACAATAATCCCTCGCCAGTGGAAATAACGATA GTGATAGCAATTACAATGATAGAAACAAAGACAGTTATACAATAACAAGAACAAATTCGTGATCATACAGAAGAGGAGCTCCCTACAGAAAATGGCGGTACTAAATCAACAGCTTGATTTAGTA ATGAATCCCATCGGTACCTGTTACCTCTTTCACTTGATGAGTTAAGGATTATGAATCANAGTTAACGACATGCTAT <u>GGAAAGGCCCTG</u>

Product	Nucleotide Sequence
OPA02G 450 2	<b>TGCCGAGCTGGAAACTCTGAGCGAAAACCGAAGAGCAGGAAAAAAATAGCACACAACTACTGTGGGCTTGAAGAAAATTCCAGCTGCTGAAGGGGGAGTTAGGAAGATGAGTCCTCCACCAGGACTCGA GGCGTCCCCTGACTCCCAGTCTCATCAATTCTGCAGCAGGCCCCGGCACAGCAGCCAGAGATGCCAATACACAGAGAAGAGAGAAGGCCACAAGGAGAATCGAGAAAGAGATGAGAGGGC AGCAGGAGGCAGGCTGGTGGACCTGTCCGCACACGAATAAGAAAAACCTCAAGAGCCAATTGTTCATGGCAACACCACAGAGCGATACCAATAAGCATGGTGGTCACAGTCGGCATCCCCGTGAGCAA AGTTGGCACGAGCAGTCCAGTAGCAGTAGCAG<u>CAGCTCGGCA</u></b>
OPA02G 450 3	<b>TGCCGAGCTGATCATCCCCTCATTACAAAGCTACTTGTGGACTCACTCCTTCCACCATGAAGAAGTCTCTGCATGATGACAACCCCTGGCATCAATGACATCACAGGGAGAGTGAATGTGAAGTGGT GGTGTCTGACATCACTGAGATCTTCTCTGGATGGCACTGCCTGGTAGCATCACCCCAACACAAACACTTCTATGCTGAGTCTTGAGGTGCTTACACAGCTAAACCTTTATGTCGAGGGAAAGCAGCCCA GTGTGGAGGCCAGGCCAGTTGTGGGGATTGGCAGTGCAGCACAGAGGAGCTTTGAAGTGTCTTACACGTATTCAAGCCACAGCCTCCACACATCTACATTCCATTGGCACAGGATCTGAGTGACAG TATCAGAGAGGAGAAGACCTTGCACTGTCAGTGCAGTCAACACATCAAGCTGTTGCCACCATGGCAG<u>CTCGGCA</u></b>
OPA07G- 350-27-1	<b>GAAACGGGTGTTGGTGTGCTCCCTCAGCGGAACCTCTGCCACCGTGTATGCTAACCTCACGCACCCAAAGAGGTATACAGCTGGACTTGGAGAACAGGTACGTGTACTGGACTGACGTTAACCGTCGAC CTTGGACGGAGCTAGCTGGATGGGTCAAGTCGACAGTCTCATCTGGAGGGCGTGAGGTGCCAAATGGACTGTGGATAGATGCTCAGCGCGGAGGATATACATAGCGATGCTCATACCAACGAGT CTTCATGTGAACATATAATGGACCGATAAAACACCTTGCGGAGGGCGTCACTGGAT<u>CACCCGTTTC</u></b>
OPA09G- 3-5	<b>GTAACGCCATCTGACTCTGGCAGTTATAGCCGACAACACTGACTGCATCCGTATTCAGAAAAATGGAGTCAGGGCTTAGCACGTAGTATGCCACAGGATGTGCCATTGACAGAGTTGCTGCCAAG AAAGGAATGAAATATTTGAGGTTCTACTGGCTGAAATACTTTGAAATCTTATGGATGCTGGAAGGGTGTCCATTGTTGGTAAGAGAGCTTGGACCGGCTG</b>
OPA11G- 350-21-2	<b>CAATCGCCGTCATCTCATTGCGAAATCATTACTGTAATTTCATATCACTGTTTTGTAAATGGAGGAGTGTGAGAGAAATTATAATATAAGGAGTCAGGAGGCGTGTGTGTGTGTGTGT GTGTGTGTGTATGTGTGTTGTATATGTTGGTGTGTGTTGGGGAGGGGGTGCAGGTGTGGTGCAGTCTTGTACCATCTCAACTATGCTTTCCACATCTGGATGCTATGTCTGGCTA GCAGCTAGGGTGCCAGCACAGCAGCAAAGCCAGGGAGAAGAAAGACAGTCTGGC<u>ACGGCGATTG</u></b>
OPA18G- 600-4-1	<b>AGGTGACCGTCTACCTCGGCTTGCCTGCACTCGCCAAACTCGAGAAAATTACGCTGAATAATGTCAGTCTCAGCGATACTCGATAGTATCAAGCAGACAGCAGCTAGCAAGGGCAAGACGTCGGCGG ATAAAATCGAACTAGTCCACTGTGGTGTATTCTGAATGGATCGGAGACGCTGGAAAGAAGCAGGAATCACCTCGGGTGTATGATTCACTGTCTCCAAAAAAAGAGGATCCTGTAACGTAGCGTCCCAT TAAGCCTCCCTGAAGTGCAGACATTACTGGTGTCTGAGAAATGTTGCCGCTCACCCAAATTCCGTGTTTATACAAAAACTGCGAACCCAGAGATGGTGGAGAAGGTGTCTCATCGTGTCCGATG CTCGCAGTGTCCATCAGCTCTCCATGCTCAAGACCCCTGAGCTGTTATGGGACTCACTAATCCAGATAACTTAAGCAAGGTGCTAGATAGTAATCCTGCAGTGGGAAGGGTGGCAGAGTATGTGACAT CCCTCATGACGTCGGACACCACTCACCTCG<u>ACGGTCACCT</u></b>
OPA18G- 650-5-1	<b>AGGTGACCGTGGTATTAAACCATTCAAGCTGGAGGACGTGCGTGAAGCCCTATCCTCTGAGGTATTCAAGGGCTGACCGTAAGCGAAGTGAAGGGTTGGCGCCAGAAAGGCATGCTGAGCTGTATC GGCGCCTGAGTACAGCGTCACCTTACCCAAAGTTAAATCGACATTGCTATCGCTGACGACAGTTAGATGAAGTAGTTGATGTCATTAGTAAAGCTGCCTATACCGAAAAAATAGGTGACGCCAAA TTTCGTTGCCGACTTGCACAGTGTCAATTGCAATTGCACTCGCACCGGGAAACAGATGAAGCAGCACTGTAATCACAGGCTCAGTAAAGTAATGGGATGGATTGATAATGAAAAAATTTCATGTTGGCC TGAGTACGGTAACCATGGTCTCTTGGCCCTGGCCGCTCCGGCGTGCAGACAAGGTGACAACGCCCTCATGATGATTGACCGCTTGGTGTTCATGACCTGCCGGTGTGGCGTTTT ACGGTGGTTGCTGCTTCAAAACGTGCTTCCATGTTGACCC<u>AGTGAACCGTCACCT</u></b>
OPA18G- 650-5-2	<b>AGGTGACCGTCTACCTCGGCTTGCCTGCACTCGCCAAACTCGAGAAAATTACGCTGAATAATGTCAGTCTCAGCGATACTCGATAGTATCAAGCAGACAGCAGCTAGCAAGGGCAAGACGTCGGCGG ATAAAATCGAACTAGTCCACTGTGGTGTATTCTGAATGGATGGAGACGCTGGAAAGAAGCAGGAATCACCTCGGGTGTATGATTCACTGTCTCCAAAAAAAGAGGATCCTGTAACGTGGCGTCCCAT TAAGCCTCCCTGAAGTGCAGACATTACTGGTGTCTGAGAAATGTTGCCGCTCACCCAAATTCCGTGTTTATACAAAAACTGCGAACCCAGAGATGGTGGAGAAGGTGTCTCATCGTGTCCGATG CTCGCAGTGTCCATCAGCTCTCCATGCTCAAGACCCCTGAGCTGTTATGGGACTCACTAATCCAGATAACTTAAGCAAGGTGCTAGATAGTAATCCTGCAGTGGGAAGGGTGGCAGAGTATGTGACAT CCCTCATGACGTCGGACACCACTCACCTCG<u>ACGGTCACCT</u></b>
OPB04G- 6-1	<b>GGACTGGAGTCCCAAACGCTTTATCTGAGGTCTCGGTCACTGCGTCTATCTGGTCAGCTTACACCTCCAGGTACAGGACTTGTGATCTCTCCCCAGATGGAGACACCGTCACATCAGTGTCTG GTCAGAAAGAGGGAGTCAGGTGCGTGGTGTGTTGACACACAGAGGCCGGTCCAGGTGTGGGACGTAGCAGTAGTAAAGAGTGTGTGTTGAATGGTCACACAGCCGAGTTGGAGCGTGGCAATGG TGACGTACTGTCCTCAGGTCACCGCAGACAGACTTCTCAGAGGGATGTGCGCACCCCATGCAATTACCCAGATAGAAGATTAGCTGGACATAGACAGGAGGTATGTGGATTAAAGTGGTCTCCAGACAG CCAGTACTTAGCCTCTGGGAAATGACAACAGTTATGTTGAACTTGCAATTCCACTCCAGT</b>

Product	Nucleotide Sequence
OPB10G-700-23-2	<u>CTGCTGGGACTCTCGGGGATCTAGGGCGGAAAGAACGACACAGTGAAGGGATTAACTTATTGAATATGGTCGGTGGCAGTGTAGGGAGAGGAAGAGGGCTCCGAGGAATGGAGGAATGATGC</u> AAAGGAGAGGAGAGTAGACGGGACGAACTGTAAGTCGATTAAAGGACTTAGAAGAAGAGCGAAGATATAAGAAGAGGAAATGGCGTGTGAGTAAGTTGGAGATGCAGGAAGCAGAGAGGGAGTAGTAG TATTACCCCTCCAGCTCAGTAATTAACAGTGGTCCGACAGATGAGCGAAGGAACAAGAGGAGCGAAGGGCGCCGAGAGGGCCTTACGTGCTCCAGCGCTCTCTCGCTGTGCTAAAGAGGCA AGCATTATGCGTGGGGATGGCTTATGCTGGACCCAAAACAAAATAATAGAATAAAAGTGGAGTGTGCTGGCAAGGCCTGTAACAAACGGTGGAAATTTAAAGAAACTGAGTGTAATTTAGT ATTTAGATTGTCGCCGGGGAAAACCTTAACAAGGACGCCTTCAGTTCCCTCGAGCTCCACCATGTAGTATTAAAGATTAAAGATTGGAGCTTGAGAGTACCGCGAATAATGAATCCGGG AGATGTTCTGAGCAGTGC <u>CCACGTCCCACAG</u>
OPB10G-700-23-3	<u>CTGCTGGACAGACAATTACGCACACGATACAGGAAATCCAACACAAATTACACGAAATTAAAGTAACAAACACCACTGGACACACTCGCAGAACAGACTATGTCAAACACTCTAAAGACCCCAG</u> AAATTTGGAGTTCTTCAAAAACATCGGGGTCGAACAAACAGATAATCACATACATATAACACACAGACAGAAGGTTGACTCGTGGAGGAGATGGAGCTCTCCATAGGGAAATTCTGGCAACAG AATTCCAAAATCTCCCCACCGAGAACCGCCGATTCCACCACATTACAGAGACGGAGTTACAAACTACATACAACACATAGGAACATAATTCTCCAGAAAACATCATTACACTAAACACTCAGAG GACAGTCCCCCTCCACACAAATCACTCAGAAGAAATAACAAACACAATCAAACCGACTAAAACACAAACACCAGGAGCTAGCAACATAAAACAAACATCATGAACATCTCCACCCAGAATGATCAGC AGACTACGCCATATCTCAATGCAGCACTGGCAACAGGATATTCCCTGACAAGTTCAAACATGCCACACTCACCTTAATACCAAACCAACAGGGAAATCTGCACACGAGGTGGGAAATTACGCCCATCTCA <u>CTCTAGAAGTCCACAGCAG</u>
OPB10G-850-22-1	<u>CTGCTGGACATCATGGATGGTCAAGTAGTCCTGTTTACAGGATTCCCTCTCCACCCATTATCTTACTCCAGACATGTCAACCGAGAAATCATGAGTCTAGCCCTATGGACGTGCCGGTGTGGA</u> ACACGGTCTAGCGGCAACAAATCTGGTCTAGCAGCCCTGGACTTCCATTCCATGGTCAAAGGAAAGTCTCTGAAGGGGAAGGGGGGCACTGCTACACCATTCTGCACCCCCCTCGATT TTATCACCAAGCATCAGGCAAAGGAGTCGAATAATTCAATATGAGTGCACAGGGAGATATAACCCCTGAATTGCAACTGGCACATGCCAAGGACAGGACAGAAAGAAAAGAAAATCATTCCG CTCAAAGACACCTGACCCCTCCGCAATGCGAGGAGGGAGAGAGATCCGCACACTGGAACCATGGCAGTCTACAGACCCCATGCACGACACGCCCTAAAAACCTATTATCCAGTGCC AAACCGCCCATGCAAGGGTAGTCCTGTAGTCACTCCAAAACAAAGCAGAAAGGCCCTACTACATCGAAGGGCAGTGGTACAGAAAGCATTCAAGTATAGCCAGTAGTGAGAGTGGCAGTAGCAC AGGGAGCAGTAGTAGTTACTGGTAGTGAAGTGAAGTGAGAGTGAAGATGAAAAGCAGGACATCTGGAGTACGGACTTAGGACAGGAATATAACCAAGTCTCATCTCGCCT
OPB10G-850-22-2	<u>CAGTAGATGATGTCATTACGAATGAGAGTAGAAAGATGTACAGGGAGAGAGAGAACAGCGGTATCAAACCTGGACGAGTTCGAGGCTCAGGAGCGCTCAAGAACCTCCGCCGGACGCCAGC</u> CCGAAACTCCTCTGGACACAGATAATCTCCAAAATCCATACAAATATACATTCCCTGAGTGAACAGAGAACGAGAATCTTTAATTGATAATGTCGAAGTGAATTTCCTCGCTGAAGGAAATT GTTAAGAAGTAAAGTATCACATTGCAAAATAATTTCGCAAGTAAACAGATGGAAAGGAAACAGACAAAAGATTAAGAAGGACAGACAGAGAGGAAAAGGAGGAAGGGAGCGGATT GGAAAGAAGTGGAGAGGGAGGGAGAGAGAGAGAGAGAGACAGGGAG GATTTAGCAGCTGGAGTGCCTACCGATGCGAAGGGCTCGGGATACAATAATCGTAAGAACGTGGCTTAAATCTCTCTATCGTAAAGTGCATAGAAGTAAAGAAGAATTAACAA AATACCGAAACTTCAAAATCTGTGTAAGAACAGTAACACACTGATAATAGGATCACAAGATAGACGCCATGCCAATTATGCCAACAATAAGGTTCACATTGGTCC <u>CAGCAG</u>
UBC101G-225-A-1	<u>GCGCCTGGAGGTACACAGAGCAGGATTGGTGCACATGTTATGTCGCACATGGGAGTTCATGACTGCACTGGAAGGAATAGTTGCTCTATTATAACCAAGAGCGCATTACAAAATATATTGA</u> GAAAATATATATTGATCTAACGTACCAAACATAATGAAAGTTCAAGGAGAGGGTAGC
UBC119G-225-29-3	<u>ATTGGGCGATGACCTTGACAATCCCAGCTACCTTCAGGTAGTAGCAGGAGAACATAACCTAGAACGAGTTGACGAGGGCAACGAGCAGCGTGTCTCCAGATCATTCAACACGAGGACTACACGGCTT</u> CTCCATCAGTAACGACATCTCCCTGCTCAAGTCTCTCACTCTGTCTTACGTAATGTGCTCATGAGTTCTGG <u>ATCGCCCAAT</u>
UBC119G-275-C-1	<u>TTGGGCGATGGGGGGAGGATGAGTGTGAAGAATGATAGTCATTGGACTTGTGCAATCATTGGTAAATTGTTACTTCCATTTCCTTCTCCGTGGATGGTAGTTAAGACGTGGA</u> TTTCACITGGCTGTCAAGCTACCAATTTCGAAACATCACGTAATTGTTACTACCTTCTGTCTTATTGTCATGATTGTCATGTCATTGACAGCGTGGTGG <u>CAAAAAAA</u> <u>AAAAAAA</u>
UBC119G-275-C-3	<u>GTTATTCACTCAGTATTGATTGTTAACGACAATGGTGAAGAAATTCAAAATCGTTGAACCTGGACCCAGAACAGGCAAGTGGAGGGTGAACGAGGACTAAAGCCTGTTGGCTAACGAGGGAGGGAGTGAATAGTAA</u> GAGGACACCAAGTGTAGTGTAGTATAGTGGTTTATTTCAGTAACCAAAGTAATTATAAGTTTATGTCATGAGTGGAGAACAGGACTAAAGCCTGTTGGCTAACGAGGGAGGGAGTGAATAGTAA TATAAGAGATTATGCCACGGAGCGTGAACACGGCAATTAGGGTATGCTAAGTGGCTAAAGGATATAAAACAAAACAGGAATTGGTATGAGGAGTGTAGCAGTATTGGATGGACCCATAAAGTGC AAATATGGATGTTAACCAATGGCATGATAGGTAACAGGAGGGTCAATTGTCACCGGGTAAACACCAAGTAATTGAGTGTGGAAACATATATGGCAGCACCATAAAGGAGGAGGGTAGGTGTTTTT

Product	Nucleotide Sequence
	GCTTCTCCCAGAACAAATTATAAGGTCCAATGTGAAGTATTGTTACAGATGATGTACCAACTAAGACATGTTAAGGACTTCCACTAACTAAAAGATGAAAGTTCTGGTTCAAAAGGAATTGATT TTGCACAAAAAAAGAGTTTGCAATTGCCATGAAGAACATCAGAACAGATAGGTCAAGTTACATAAGTCTCGTTATCTTCNTTNCCNTGCCCAAT
UBC119G -350-5-3	<u>ATGGGGCGATGACTTTGACAATCCCAGCTACCTTCAGGTAGTAGCAGGAGAACATAACCTAGAAGTTGACGAGGGCAACGAGCACAGGTCGTCTCTCCAAAGATCATTCAACACGAGGACTACAACGGCTT</u> CTCCATCAGTAACGACATCTCCCTGCTCAAGTTCTCACTCTGTCCCTCAACGACTTCGTGCACTACGTCGACCTGAGCAAGAACCCGCTGGAGTGTGATTGCGCCGTGAGTGG <u>ATCGCCCAAT</u>
UBC122G -7-2	<u>GTAGACGAGCCAGACTATGGTTGCAACGGGTCGACTATTGCGAGCAGCCCCGATGACATCACCTGCTTCTGACGTCAGACCACACTACACTGACGAAGGAGTCGAGATCGGAATTCCCGACCTGCTCGCAT</u> TACTCACGCCAGTATTACGATGGAAAGGACATGAACGCTTACGCCACAGCAGGCAGTACACAACAGCCAGGAGCAGA <u>AGCTCGTCTAC</u>
UBC135G -200-24-2	<u>AAGCTGCGAGGGAGAGCCTGGCGATGCTGGCGAACAGACCCACTCCTGGTAGGCCTAACGAGGAGTCCAATTCCCTTGACCAACATCTTGAAACAGAGTCCTCACGTTCTAAAAAAAGAAAAATAAA</u> TCGCATGTATGTTAATTTTTTGTTTTAGAGTCCCGATTGTTCTCGCTTCTCG <u>CTCGCAGCTT</u>
UBC135G -200-25-2	<u>AAGCTGCGAGAGGTGGAGGTGAAGTCCAAATCATCTGATGAGCATAGTGGAGTTAGTTGGATCAAGAGGAACAGCTGGTAGAGGAGAGAGATTGCCAGGAGAGAATAATATTGGCATGGTGCTGGC</u> GCATGACACTACACACACCCAGAGTATCCGATGCCGTGATATCATTGTGATCTGTAACGACATCACCTACCAAAATAGGATCCTTGCCACATGAGGATATTCTGTTCTCAGAGC <u>CTCGCAGCTT</u>

Remark: Bald-underlined letter indicates primer location

## APPENDIX C

### Blast X result for carboxylesterase

Sequences producing significant alignments:

		Score (Bits)	E Value
gi 91084115 ref XP_967137.1	PREDICTED: similar to CG6414-PA [Tr	311	1e-82
gi 91086415 ref XP_967183.1	PREDICTED: similar to CG10175-PC...	294	1e-77
gi 48097744 ref XP_391943.1	PREDICTED: similar to CG6414-PA [Ap	287	2e-75
gi 54311783 emb CAH64510.1	putative esterase [Tribolium castane	282	5e-74
gi 100811805 dbj BAE94685.1	juvenile hormone esterase [Psacotho	281	7e-74
gi 86515416 ref NP_001034534.1	putative esterase [Tribolium ...	280	2e-73
gi 54311781 emb CAH64509.1	putative esterase [Tribolium castane	279	4e-73
gi 62086395 dbj BAD91555.1	carboxylesterase [Athalia rosae]	278	7e-73
gi 54311777 emb CAH64507.1	putative esterase [Tribolium castane	273	2e-71
gi 108876577 gb EAT40802.1	alpha-esterase [Aedes aegypti]	270	2e-70
gi 11761909 gb AAG40239.1 AF302777.1	carboxylesterase precursor	270	2e-70
gi 54019713 emb CAH60164.1	esterase [Tribolium castaneum]	270	2e-70
gi 54019717 emb CAH60166.1	putative esterase [Tribolium castane	270	3e-70
gi 54019719 emb CAH60167.1	putative esterase [Tribolium confusu	270	3e-70
gi 54311779 emb CAH64508.1	putative esterase [Tribolium castane	268	1e-69
gi 54019715 emb CAH60165.1	putative esterase [Tribolium castane	266	3e-69
gi 86515386 ref NP_001034512.1	esterase 1 [Tribolium castane...	265	5e-69
gi 91086429 ref XP_967916.1	PREDICTED: similar to CG1128-PB...	262	5e-68
gi 17646748 gb ALA41023.1 AF448479.1	juvenile hormone esterase [	262	5e-68
gi 54019721 emb CAH60168.1	putative esterase [Tribolium freeman	262	5e-68
gi 544256 sp P35502 ESTF_MYZPE	Esterase FE4 precursor (Carbox...	262	5e-68
gi 91084517 ref XP_972335.1	PREDICTED: similar to CG10175-PC...	261	9e-68
gi 91086427 ref XP_967835.1	PREDICTED: similar to CG10175-PC...	260	2e-67
gi 115650767 ref XP_782312.2	PREDICTED: similar to acetylcho...	259	4e-67
gi 91086425 ref XP_967598.1	PREDICTED: similar to CG10175-PC...	259	4e-67
gi 91089215 ref XP_967444.1	PREDICTED: similar to CG10175-PC...	259	5e-67
gi 544255 sp P35501 ESTE_MYZPE	Esterase E4 precursor (Carboxy...	258	1e-66
gi 62086393 dbj BAD91554.1	juvenile hormone esterase [Athalia r	257	2e-66
gi 108881320 gb EAT45545.1	carboxylesterase [Aedes aegypti]	256	2e-66
gi 72077750 ref XP_782249.1	PREDICTED: similar to acetylchol...	255	5e-66
gi 37574080 ref NP_932116.1	hypothetical protein LOC72361 [M...	255	5e-66
gi 115653078 ref XP_788440.2	PREDICTED: similar to acetylcho...	254	9e-66
gi 108881319 gb EAT45544.1	carboxylesterase [Aedes aegypti]	254	1e-65
gi 66560187 ref XP_392698.2	PREDICTED: similar to CG10175-PC, i	253	2e-65
gi 91086417 ref XP_967268.1	PREDICTED: similar to CG10175-PC...	252	4e-65
gi 91084421 ref XP_968215.1	PREDICTED: similar to CG6414-PA [Tr	252	4e-65
gi 118782888 ref XP_312564.3	ENSANGP00000014256 [Anopheles g...	252	6e-65
gi 68697266 emb CAJ14159.1	putative esterase [Anopheles gambiae	251	7e-65
gi 91091806 ref XP_970836.1	PREDICTED: similar to CG6414-PA [Tr	251	1e-64
gi 77735475 ref NP_001029432.1	carboxylesterase 2 (intestine...	251	1e-64
gi 116829962 gb ABK27874.1	carboxylesterase [Bombyx mori]	249	4e-64
gi 91083857 ref XP_974072.1	PREDICTED: similar to CG4382-PA [Tr	249	4e-64
gi 37718991 ref NP_937814.1	hypothetical protein LOC234669 [...	249	4e-64
gi 119721182 gb ABL98071.1	carboxylesterase [Bombyx mandarina]	249	5e-64
gi 91086419 ref XP_967349.1	PREDICTED: similar to CG10175-PC...	249	5e-64
gi 108881318 gb EAT45543.1	carboxylesterase [Aedes aegypti]	248	6e-64
gi 84095052 dbj BAE66716.1	carboxylesterase [Aphis gossypii]	248	6e-64
gi 84095050 dbj BAE66715.1	carboxylesterase [Aphis gossypii]	248	8e-64
gi 119699079 gb ABL96242.1	carboxylesterase [Bombyx mori]	247	1e-63
gi 91076732 ref XP_972864.1	PREDICTED: similar to CG10175-PC...	247	1e-63
gi 108881317 gb EAT45542.1	carboxylesterase [Aedes aegypti]	247	2e-63
gi 28317060 gb AAO39549.1	RE03380p [Drosophila melanogaster]	247	2e-63
gi 24649333 ref NP_651151.1	CG10175-PC, isoform C [Drosophil...	247	2e-63
gi 24649335 ref NP_732874.1	CG10175-PA, isoform A [Drosophil...	247	2e-63
gi 3426006 dbj BAA32385.1	carboxylesterase precursor [Aphis ...	246	2e-63
gi 42412531 gb AAS15642.1	carboxylesterase [Aphis gossypii] ...	246	3e-63
gi 27658990 ref XP_226397.1	PREDICTED: similar to carboxyles...	246	4e-63
gi 72007966 ref XP_786979.1	PREDICTED: similar to acetylchol...	246	4e-63
gi 118782091 ref XP_312052.3	ENSANGP00000016214 [Anopheles g...	245	5e-63
gi 91084505 ref XP_972277.1	PREDICTED: similar to CG10175-PC...	245	5e-63
gi 125773103 ref XP_001357810.1	GA10132-PA [Drosophila pseud...	245	5e-63
gi 42412533 gb AAS15643.1	carboxylesterase [Aphis gossypii]	245	7e-63
gi 54043019 gb AAV28503.1	acetylcholinesterase [Culex pipiens p	244	9e-63
gi 34222522 sp Q86GC8 ACES_CULPI	Acetylcholinesterase precursor	244	9e-63
gi 32968054 emb CAD33707.2	acetylcholinesterase [Culex pipiens]	244	9e-63
gi 91084423 ref XP_968291.1	PREDICTED: similar to CG6414-PA [Tr	244	1e-62

## APPENDIX C

## Blast X result for carboxylesterase

Sequences producing significant alignments:

		Score (Bits)	E Value
gi 91084115 ref XP 967137.1	PREDICTED: similar to CG6414-PA [Tr]	311	1e-82
gi 91086415 ref XP 967183.1	PREDICTED: similar to CG10175-PC...	294	1e-77
gi 48097744 ref XP 391943.1	PREDICTED: similar to CG6414-PA [Ap	287	2e-75
gi 54311783 emb CAH64510.1	putative esterase [Tribolium castane	282	5e-74
gi 100811805 dbj BAE94685.1	juvenile hormone esterase [Psacoth	281	7e-74
gi 86515416 ref NP 001034534.1	putative esterase [Tribolium ...	280	2e-73
gi 54311781 emb CAH64509.1	putative esterase [Tribolium castane	279	4e-73
gi 62086395 dbj BAD91555.1	carboxylesterase [Athalia rosae]	278	7e-73
gi 54311777 emb CAH64507.1	putative esterase [Tribolium castane	273	2e-71
gi 108876577 gb EAT40802.1	alpha-esterase [Aedes aegypti]	270	2e-70
gi 11761909 gb AAG40239.1 AF302777_1	carboxylesterase precursor	270	2e-70
gi 54019713 emb CAH60164.1	esterase [Tribolium castaneum]	270	2e-70
gi 54019717 emb CAH60166.1	putative esterase [Tribolium castane	270	3e-70
gi 54019719 emb CAH60167.1	putative esterase [Tribolium confusu	270	3e-70
gi 54311779 emb CAH64508.1	putative esterase [Tribolium castane	268	1e-69
gi 54019715 emb CAH60165.1	putative esterase [Tribolium castane	266	3e-69
gi 86515386 ref NP 001034512.1	esterase 1 [Tribolium castane...	265	5e-69
gi 91086429 ref XP 967916.1	PREDICTED: similar to CG1128-PB...	262	5e-68
gi 17646748 gb AAL41023.1 AF448479_1	juvenile hormone esterase [	262	5e-68
gi 54019721 emb CAH60168.1	putative esterase [Tribolium freeman	262	5e-68
gi 544256 sp P35502 ESTF MYZPE	Esterase FE4 precursor (Carbox...	262	5e-68
gi 91084517 ref XP 972335.1	PREDICTED: similar to CG10175-PC...	261	9e-68
gi 91086427 ref XP 967835.1	PREDICTED: similar to CG10175-PC...	260	2e-67
gi 115650767 ref XP 782312.2	PREDICTED: similar to acetylcho...	259	4e-67
gi 91086425 ref XP 967598.1	PREDICTED: similar to CG10175-PC...	259	4e-67
gi 91089215 ref XP 967444.1	PREDICTED: similar to CG10175-PC...	259	5e-67
gi 544255 sp P35501 ESTE MYZPE	Esterase E4 precursor (Carboxy...	258	1e-66
gi 62086393 dbj BAD91554.1	juvenile hormone esterase [Athalia r	257	2e-66
gi 108881320 gb EAT45545.1	carboxylesterase [Aedes aegypti]	256	2e-66
gi 72077750 ref XP 782249.1	PREDICTED: similar to acetylchol...	255	5e-66
gi 37574080 ref NP 932116.1	hypothetical protein LOC72361 [M...	255	5e-66
gi 115653078 ref XP 788440.2	PREDICTED: similar to acetylcho...	254	9e-66
gi 108881319 gb EAT45544.1	carboxylesterase [Aedes aegypti]	254	1e-65
gi 66560187 ref XP 392698.2	PREDICTED: similar to CG10175-PC, i	253	2e-65
gi 91086417 ref XP 967268.1	PREDICTED: similar to CG10175-PC...	252	4e-65
gi 91084421 ref XP 968215.1	PREDICTED: similar to CG6414-PA [Tr	252	4e-65
gi 118782888 ref XP 312564.3	ENSANGP00000014256 [Anopheles g...	252	6e-65
gi 68697266 emb CAJ14159.1	putative esterase [Anopheles gambiae	251	7e-65
gi 91091806 ref XP 970836.1	PREDICTED: similar to CG6414-PA [Tr	251	1e-64
gi 77735475 ref NP 001029432.1	carboxylesterase 2 (intestine...	251	1e-64
gi 116829962 gb ABK27874.1	carboxylesterase [Bombyx mori]	249	4e-64
gi 91083857 ref XP 974072.1	PREDICTED: similar to CG4382-PA [Tr	249	4e-64
gi 37718991 ref NP 937814.1	hypothetical protein LOC234669 [...	249	4e-64
gi 119721182 gb ABL98071.1	carboxylesterase [Bombyx mandarina]	249	5e-64
gi 91086419 ref XP 967349.1	PREDICTED: similar to CG10175-PC...	249	5e-64
gi 108881318 gb EAT45543.1	carboxylesterase [Aedes aegypti]	248	6e-64
gi 84095052 dbj BAE66716.1	carboxylesterase [Aphis gossypii]	248	8e-64
gi 84095050 dbj BAE66715.1	carboxylesterase [Aphis gossypii]	248	1e-63
gi 119699079 gb ABL96242.1	carboxylesterase [Bombyx mori]	247	1e-63
gi 91076732 ref XP 972864.1	PREDICTED: similar to CG10175-PC...	247	1e-63
gi 108881317 gb EAT45542.1	carboxylesterase [Aedes aegypti]	247	2e-63
gi 28317060 gb AAO39549.1	RE03380p [Drosophila melanogaster]	247	2e-63
gi 24649333 ref NP 651151.1	CG10175-PC, isoform C [Drosophil...	247	2e-63
gi 24649335 ref NP 732874.1	CG10175-PA, isoform A [Drosophil...	247	2e-63
gi 3426006 dbj BAA32385.1	carboxylesterase precursor [Aphis ...	246	2e-63
gi 42412531 gb AAS15642.1	carboxylesterase [Aphis gossypii] ...	246	3e-63
gi 27658990 ref XP 226397.1	PREDICTED: similar to carboxyles...	246	4e-63
gi 72007966 ref XP 786979.1	PREDICTED: similar to acetylchol...	246	4e-63
gi 118782091 ref XP 312052.3	ENSANGP00000016214 [Anopheles g...	245	5e-63
gi 91084505 ref XP 972277.1	PREDICTED: similar to CG10175-PC...	245	5e-63
gi 125773103 ref XP 001357810.1	GA10132-PA [Drosophila pseud...	245	5e-63
gi 42412533 gb AAS15643.1	carboxylesterase [Aphis gossypii]	245	7e-63
gi 54043019 gb AAV28503.1	acetylcholinesterase [Culex pipiens p	244	9e-63
gi 34222522 sp Q86GC8 ACES_CULPI	Acetylcholinesterase precursor	244	9e-63
gi 32968054 emb CAD33707.2	acetylcholinesterase [Culex pipiens]	244	9e-63
gi 91084423 ref XP 968291.1	PREDICTED: similar to CG6414-PA [Tr	244	1e-62

gi 66505864 ref XP_394404.2	PREDICTED: similar to Esterase-6...	244	2e-62
gi 24583086 ref NP_609301.1	CG4382-PA [Drosophila melanogast...	244	2e-62
gi 71834068 dbj BAE16975.1	juvenile hormone esterase isoform A	243	2e-62
gi 67678086 gb AAH97486.1	LOC679149 protein [Rattus norvegicus]	243	3e-62
gi 115653076 ref XP_788459.2	PREDICTED: similar to cholinest...	243	3e-62
gi 29120004 emb CAD56155.1	acetylcholinesterase [Culex pipiens]	242	4e-62
gi 71834070 dbj BAE16976.1	juvenile hormone esterase isoform B	242	6e-62
gi 42412529 gb AAS15641.1	carboxylesterase [Aphis gossypii]	241	8e-62
gi 15983755 gb AAL09822.1	carboxylesterase [Aphis gossypii]	241	8e-62
gi 62002225 gb AAX58712.1	pheromone-degrading enzyme 2 [Anthera	241	8e-62
gi 40363516 dbj BAD06210.1	acetylcholinesterase [Culex tritaeni	241	8e-62
gi 108881316 gb EAT45541.1	carboxylesterase [Aedes aegypti]	241	1e-61
gi 62002223 gb AAX58711.1	pheromone-degrading enzyme 1 [Anthera	241	1e-61
gi 21064383 gb AAM29421.1	RE16761p [Drosophila melanogaster]	240	2e-61
gi 108881321 gb EAT45546.1	carboxylesterase [Aedes aegypti]	240	2e-61
gi 24639611 ref NP_570089.1	CG6414-PA [Drosophila melanogast...	240	2e-61
gi 47213516 emb CAF96163.1	unnamed protein product [Tetraodon n	240	2e-61
gi 91091808 ref XP_970896.1	PREDICTED: similar to CG6414-PA [Tr	239	3e-61
gi 120474987 ref NP_001073334.2	hypothetical protein LOC7190...	239	3e-61
gi 88192514 pdb 2COP A	Chain A, Aged Form Of Mouse Acetylchol...	239	3e-61
gi 46015343 pdb 1Q83 A	Chain A, Crystal Structure Of The Mous...	239	3e-61
gi 40889078 pdb 1KU6 A	Chain A, Fasciculin 2-Mouse Acetylcholine	239	3e-61
gi 1421161 pdb 1MAH A	Chain A, Fasciculin2 - Mouse Acetylchol...	239	3e-61
gi 13928664 ref NP_033729.1	acetylcholinesterase [Mus muscul...	239	3e-61
gi 6980490 pdb 1C20 A	Chain A, Electrophorus Electricus Acety...	239	3e-61
gi 6730113 pdb 1C2B A	Chain A, Electrophorus Electricus Acetylch	239	3e-61
gi 28373898 pdb 1N5M A	Chain A, Crystal Structure Of The Mous...	239	3e-61
gi 3062827 dbj BAA25691.1	carboxylesterase precursor [Rattus no	239	4e-61
gi 2641986 dbj BAA23605.1	carboxylesterase precursor [Mesocricet...	239	4e-61
gi 112491234 pdb 2HA4 A	Chain A, Crystal Structure Of Mutant ...	238	6e-61
gi 24649337 ref NP_732875.1	CG10175-PB, isoform B [Drosophil...	238	6e-61
gi 89148031 gb ABD62772.1	esterase [Chilo suppressalis]	238	8e-61
gi 91095115 ref XP_969956.1	PREDICTED: similar to Esterase-6...	238	8e-61
gi 62087113 dbj BAD92015.1	carboxylesterase [Athalia rosae]	238	8e-61

## Blast X result for glutathione-s-transferase

Sequences producing significant alignments:

		Score (Bits)	E Value
gi 91080623 ref XP_974273.1	PREDICTED: similar to Glutathione...	189	1e-46
gi 22218855 pdb 1JLV A	Chain A, Anopheles Dirus Species B Glu...	182	2e-44
gi 53828193 emb CAH58743.1	putative glutathione S-transferase...	181	4e-44
gi 31208165 ref XP_313049.1	ENSANGP00000011661 [Anopheles ga...	178	3e-43
gi 1632771 emb CAB03592.1	GSTD1-5 protein [Anopheles gambiae]	177	4e-43
gi 2738075 gb AAB94639.1	glutathione S transferase-1 [Culicoide	177	6e-43
gi 60678789 gb AAX33729.1	Per 5 allergen [Periplaneta americana	176	1e-42
gi 14517793 gb AAK64362.1 AF336288_1	glutathione-S-transferase-l	174	6e-42
gi 11596154 gb AAG38507.1 AF273041_1	glutathione transferase ...	174	6e-42
gi 108883615 gb EAT47840.1	glutathione-s-transferase theta, gst	173	8e-42
gi 1786091 gb AAB41104.1	glutathione S-transferase I [Anopheles	172	2e-41
gi 112983444 ref NP_001036974.1	glutathione S-transferase th...	171	3e-41
gi 3511227 gb AAC79994.1	glutathione S-transferase [Anophele...	171	4e-41
gi 1346214 sp P42860 GSTT1 LUCCU	Glutathione S-transferase 1-1 (	171	4e-41
gi 31208163 ref XP_313048.1	ENSANGP00000023508 [Anopheles ga...	171	4e-41
gi 21464458 gb AAM52032.1	RH47312p [Drosophila melanogaster]	171	5e-41
gi 2842718 sp Q93113 GSTT6 ANOGA	Glutathione S-transferase 1-...	170	7e-41
gi 121696 sp P28338 GSTT1 MUSDO	Glutathione S-transferase 1 (...)	170	7e-41
gi 58384153 ref XP_313050.2	ENSANGP00000024808 [Anopheles gambi	170	7e-41
gi 14538008 gb AAK66764.1 AF386788_1	glutathione S-transferase D	169	2e-40
gi 385883 gb AAB26519.1	glutathione S-transferase D1, DmGST1...	169	2e-40
gi 17737923 ref NP_524326.1	Glutathione S transferase D1 CG1...	169	2e-40
gi 54637753 gb EAL27155.1	GA10031-PA [Drosophila pseudoobscura]	169	2e-40
gi 1495235 emb CAA96105.1	GSTD2 protein [Anopheles gambiae]	169	2e-40
gi 108873197 gb EAT37422.1	glutathione-s-transferase theta, gst	167	8e-40
gi 116128452 gb EAA09273.4	ENSANGP00000013097 [Anopheles gambia	166	1e-39
gi 21435007 gb AAM53609.1 AF513637_1	glutathione S-transferase D	166	1e-39
gi 10443882 gb AAG17624.1	glutathione transferase [Anopheles...	166	1e-39
gi 58385036 ref XP_313665.2	ENSANGP00000013097 [Anopheles gambi	166	1e-39
gi 1125671 emb CAA63946.1	GST-3/GST-5; glutathione transferase	166	2e-39
gi 20386063 gb AAM21563.1 AF448500_1	glutathione S-transferase [	164	4e-39
gi 1079182 pir IS43851	glutathione transferase (EC 2.5.1.18) ...	163	9e-39
gi 1125669 emb CAA63945.1	GST-3; glutathione transferase [Musca	163	1e-38
gi 242503 gb AAB20908.1	glutathione S-transferase 27, GST27=...	163	1e-38
gi 1125663 emb CAA63948.1	GST-3/GST-5; glutathione transferase	163	1e-38
gi 17864598 ref NP_524916.1	Glutathione S transferase D8 CG4...	163	1e-38
gi 385882 gb AAB26518.1	glutathione S-transferase D27, DmGST...	162	1e-38

gi 1170120 sp P46433 GSTT4 MUSDO	Glutathione S-transferase 4 ...	162	3e-38
gi 54637754 gb EAL27156.1	GA10065-PA [Drosophila pseudoobscura]	162	3e-38
gi 1708068 sp P46431 GSTT2 MUSDO	Glutathione S-transferase 2 ...	161	3e-38
gi 1170119 sp P46432 GSTT3 MUSDO	Glutathione S-transferase 3 ...	161	3e-38
gi 232194 sp P30107 GSTT1 DROTE	Glutathione S-transferase 1-1 (G	159	1e-37
gi 232191 sp P30104 GSTT1 DROER	Glutathione S-transferase 1-1 (G	159	2e-37
gi 1125661 emb CAA63950.1	GST-4; glutathione transferase [Musca	159	2e-37
gi 54037234 sp P67804 GSTT1 DROMA	Glutathione S-transferase 1...	158	3e-37
gi 232195 sp P30108 GSTT1 DROYA	Glutathione S-transferase 1-1 (G	158	3e-37
gi 27752553 gb AAO19738.1	glutathione S-transferase [Bactrocera	158	4e-37
gi 232193 sp P30106 GSTT1 DROSE	Glutathione S-transferase 1-1 (G	158	4e-37
gi 2117755 pir S51566	glutathione transferase (EC 2.5.1.18) 2 -	157	8e-37
gi 22218861 pdb 1JLW A	Chain A, Anopheles Dirus Species B Glu...	156	1e-36
gi 56462176 gb AAV91371.1	hypothetical protein 3 [Lonomia obliqua]	155	2e-36
gi 38493021 pdb 1R5A A	Chain A, Glutathione S-Transferase >gi...	155	2e-36
gi 54637757 gb EAL27159.1	GA14590-PA [Drosophila pseudoobscura]	155	3e-36
gi 54637755 gb EAL27157.1	GA18009-PA [Drosophila pseudoobscura]	155	3e-36
gi 24646251 ref NP 650181.1	Glutathione S transferase D9 CG1...	154	5e-36
gi 1125665 emb CAA63952.1	GST-5; glutathione transferase [Musca	154	7e-36
gi 24646259 ref NP 650183.1	CG17639-PA [Drosophila melanogaster]	153	9e-36
gi 110763730 ref XP 392997.3	PREDICTED: similar to Glutathione-S-...	153	1e-35
gi 76262439 gb AAT39512.2	glutathione-S-transferase 1 [Apis mellifera]	153	1e-35
gi 17864592 ref NP 524912.1	Glutathione S transferase D2 CG4...	153	1e-35
gi 116128908 gb EAA08622.3	ENSANGP00000011261 [Anopheles gambiae]	152	2e-35
gi 116128909 gb EAL40657.2	ENSANGP00000028057 [Anopheles gambiae]	152	2e-35
gi 385880 gb AAB26516.1	glutathione S-transferase D21, DmGST...	152	2e-35
gi 57967576 ref XP 562675.1	ENSANGP00000011261 [Anopheles gambiae]	152	2e-35
gi 3511225 gb AACT79992.1	glutathione S-transferase [Anopheles gambiae]	151	3e-35
gi 57967586 ref XP 562680.1	ENSANGP00000028492 [Anopheles gambiae]	151	3e-35
gi 58384149 ref XP 313047.2	ENSANGP00000023440 [Anopheles gambiae]	151	3e-35
gi 45549270 ref NP 524914.3	Glutathione S transferase D5 CG1...	151	4e-35
gi 54637756 gb EAL27158.1	GA18171-PA [Drosophila pseudoobscura]	151	4e-35
gi 385879 gb AAB26515.1	glutathione S-transferase D24, DmGST...	150	6e-35
gi 1125675 emb CAA63951.1	GST-5; glutathione transferase [Musca	150	8e-35
gi 17864594 ref NP 524913.1	Glutathione S transferase D4 CG1...	149	2e-34
gi 385881 gb AAB26517.1	glutathione S-transferase D23, DmGST...	149	2e-34
gi 108883616 gb EAT47841.1	glutathione-s-transferase theta, gst	147	5e-34
gi 108883614 gb EAT47839.1	glutathione-s-transferase theta, gst	146	1e-33
gi 91085767 ref XP 974204.1	PREDICTED: similar to Glutathione-S-...	144	7e-33
gi 54637752 gb EAL27154.1	GA14986-PA [Drosophila pseudoobscura]	143	9e-33
gi 24646249 ref NP 652713.1	Glutathione S transferase D10 CG...	143	1e-32
gi 116128453 gb EAL40527.2	ENSANGP00000029040 [Anopheles gambiae]	142	2e-32
gi 112984484 ref NP 001037183.1	glutathione S-transferase [Bacillus subtilis]	141	5e-32
gi 78172883 gb ABB29466.1	glutathione S-transferase [Corcyra cephalonica]	140	8e-32
gi 55794094 gb AAV65948.1	delta glutathione-S-transferase [Saccharomyces cerevisiae]	139	1e-31
gi 14495354 gb AAK64286.1 AF384858_1	glutathione S-transferases	133	9e-31
gi 6560681 gb AAF16718.1 AF117596_1	glutathione S-transferase [Mycobacterium tuberculosis]	135	2e-30
gi 57966486 ref XP 562128.1	ENSANGP00000029040 [Anopheles gambiae]	135	2e-30
gi 85861087 gb ABC86493.1	IP02541p [Drosophila melanogaster]	135	3e-30
gi 477693 pir  B46681	glutathione transferase (EC 2.5.1.18) D...	135	3e-30
gi 17933730 ref NP 525114.1	Glutathione S transferase D7 CG4...	135	3e-30
gi 478238 pir  H46681	glutathione transferase (EC 2.5.1.18) D...	135	3e-30
gi 112982796 ref NP 001037546.1	glutathione S-transferase 3 ...	135	3e-30
gi 17864596 ref NP 524915.1	Glutathione S transferase D6 CG4...	135	3e-30
gi 12007376 gb AAG45165.1 AF316637_1	glutathione S-transferase D	133	1e-29
gi 57967606 ref XP 562690.1	ENSANGP00000028085 [Anopheles gambiae]	133	1e-29
gi 58384160 ref XP 313057.2	ENSANGP00000000321 [Anopheles gambiae]	133	1e-29
gi 4704804 gb AAD28279.1 AF133268_1	glutathione S-transferase GS	132	3e-29
gi 46391804 gb AAS90947.1	glutathione S-transferase [Aedes aegypti]	131	5e-29
gi 108883613 gb EAT47838.1	glutathione-s-transferase theta, gst	130	8e-29
gi 91080625 ref XP 974300.1	PREDICTED: similar to Glutathione-S-...	130	8e-29
gi 91076556 ref XP 966702.1	PREDICTED: similar to CG17531-PA [Tetrahymena thermophila]	129	2e-28
gi 91078560 ref XP 971136.1	PREDICTED: similar to CG17533-PA [Tetrahymena thermophila]	128	3e-28

## Blast X result for glutathione-s-transferase

Sequences producing significant alignments:	Score (Bits)	E Value	
gi 60729680 pir  JC8026	cytochrome P450 enzyme, CYP4C39 enzym...	535	4e-150
gi 6456874 gb AAF09264.1 AF091117_1	cytochrome P450 [Orconectes limosus]	482	3e-134
gi 18032259 gb AAL56662.1 AF263607_1	cytochrome P450 CYP4 [Cherry tree hornworm, Manduca sexta]	471	8e-131
gi 50657412 ref NP 001001879.1	cytochrome P450, family 4, subfamily 4, polypeptide 1 [Aedes vexans]	434	1e-119
gi 108871345 gb EAT35570.1	cytochrome P450 [Aedes aegypti]	424	6e-117
gi 231885 sp P29981 CP4C1 BLADI	Cytochrome P450 4C1 (CYPIVC1)...	423	2e-116
gi 108876007 gb EAT40232.1	cytochrome P450 [Aedes aegypti]	412	3e-113

gi 17864130 ref NP_524598.1	Cytochrome P450-4c3 CG1438-PA [Drosophila pseudoobscura]	407	1e-111
gi 54637319 gb EAL26721.1	GA12945-PA [Drosophila pseudoobscura]	402	4e-110
gi 118404542 ref NP_001072667.1	hypothetical protein LOC7801...	397	1e-108
gi 109076402 ref XP_001088961.1	PREDICTED: similar to cytochrome...	394	9e-108
gi 121583883 ref NP_001073465.1	hypothetical protein LOC5620...	394	1e-107
gi 114597206 ref XP_001165629.1	PREDICTED: hypothetical protein	392	3e-107
gi 114597208 ref XP_001165592.1	PREDICTED: hypothetical protein	392	3e-107
gi 68373569 ref XP_687753.1	PREDICTED: similar to cytochrome...	392	3e-107
gi 119625029 gb EAX04624.1	cytochrome P450, family 4, subfamily A	392	5e-107
gi 34532967 dbj BAC86562.1	unnamed protein product [Homo sapien]	392	5e-107
gi 61743922 ref NP_997235.2	cytochrome P450, family 4, subfamily A	392	5e-107
gi 116642350 dbj BAF35771.1	cytochrome P450 4 family [Daphnia magna]	391	8e-107
gi 38603630 dbj BAD02915.1	Cytochrome P450 [Xenopus laevis]	390	2e-106
gi 115625651 ref XP_783244.2	PREDICTED: similar to ENSANGP00...	389	4e-106
gi 73979556 ref XP_849364.1	PREDICTED: similar to cytochrome...	388	5e-106
gi 117606212 ref NP_001071070.1	hypothetical protein LOC5588...	387	9e-106
gi 71648657 sp Q5RCN6 CP4V2_PONPY	Cytochrome P450 4V2 >gi 557...	387	9e-106
gi 74151909 dbj BAE29740.1	unnamed protein product [Mus musculus]	385	6e-105
gi 19527190 ref NP_598730.1	family 4 cytochrome P450 [Mus musculus]	384	7e-105
gi 109503398 ref XP_001064152.1	PREDICTED: similar to family...	383	2e-104
gi 49257971 gb AAH74131.1	MGC81840 protein [Xenopus laevis]	383	2e-104
gi 108876008 gb EAT40233.1	cytochrome P450 [Aedes aegypti]	380	2e-103
gi 77735695 ref NP_001029545.1	hypothetical protein LOC51015...	379	2e-103
gi 108876009 gb EAT40234.1	cytochrome P450 [Aedes aegypti]	374	8e-102
gi 47216297 emb CAF96593.1	unnamed protein product [Tetraodon nigriventer]	371	8e-101
gi 47605530 sp Q964T1 CP4CU_BLAZE	Cytochrome P450 4c21 (CYP4...	355	6e-96
gi 2431938 gb AAB71169.1	cytochrome P450 [Drosophila melanogaster]	352	3e-95
gi 12644424 sp Q27589 CP4D2_DROME	Cytochrome P450 4d2 (CYP4VD2)	352	3e-95
gi 2431964 gb AAB71182.1	cytochrome P450 [Drosophila simulans]	351	7e-95
gi 3249041 gb AAC69184.1	corpora allata cytochrome P450 [Diplopoda]	351	7e-95
gi 2431960 gb AAB71180.1	cytochrome P450 [Drosophila melanogaster]	350	2e-94
gi 17933518 ref NP_525043.1	Cytochrome P450-4d2 CG3466-PA [Drosophila melanogaster]	350	2e-94
gi 91085551 ref XP_966563.1	PREDICTED: similar to Cytochrome...	349	3e-94
gi 542555 pir S41192	cytochrome P450 4D2 - fruit fly (Drosophila melanogaster)	348	6e-94
gi 2894114 emb CAA15698.1	EG:152A3.4 [Drosophila melanogaster]	346	3e-93
gi 93278141 gb ABF06549.1	CYP4BF1 [Ips paraconfusus]	345	4e-93
gi 312904 emb CAA80549.1	cytochrome P-450 [Drosophila melanogaster]	344	1e-92
gi 47779228 gb AAT38512.1	pheromone-degrading enzyme [Phyllopertha horticola]	338	5e-91
gi 108876251 gb EAT40476.1	cytochrome P450 [Aedes aegypti]	337	1e-90
gi 108876247 gb EAT40472.1	cytochrome P450 [Aedes aegypti]	337	2e-90
gi 94158626 ref NP_001035323.1	cytochrome P450 monooxygenase...	335	5e-90
gi 91078612 ref XP_966411.1	PREDICTED: similar to Cytochrome...	332	6e-89
gi 21355669 ref NP_652020.1	Cyp4d14 CG3540-PA [Drosophila melanogaster]	332	6e-89
gi 38679391 gb AAR26517.1	antennal cytochrome P450 CYP4 [Mamestra brassicae]	329	4e-88
gi 5263306 gb AAC03111.2	family 4 cytochrome P450 [Coptotermes acinaciformis]	329	4e-88
gi 37287641 gb AAQ90477.1	cytochrome P450 CYP4AB2 [Solenopsis invicta]	328	6e-88
gi 108876250 gb EAT40475.1	cytochrome P450 [Aedes aegypti]	326	3e-87
gi 93278137 gb ABF06547.1	CYP4BE1 [Ips paraconfusus]	325	7e-87
gi 91094839 ref XP_971612.1	PREDICTED: similar to CG3466-PA [Trichoplax adhaerens]	324	1e-86
gi 115625653 ref XP_783176.2	PREDICTED: hypothetical protein...	323	2e-86
gi 5230695 gb AAD40966.1 AF081807_1	cytochrome P450 4W1 [Boophilus microplus]	323	3e-86
gi 91078618 ref XP_967724.1	PREDICTED: similar to Cytochrome...	322	3e-86
gi 72014091 ref XP_786946.1	PREDICTED: hypothetical protein ...	321	1e-85
gi 56710314 dbj BAD81026.1	cytochrome P450 CYP4G25 [Antheraea yamamai]	320	2e-85
gi 91082471 ref XP_971854.1	PREDICTED: similar to Cytochrome...	319	3e-85
gi 54643537 gb EAL32280.1	GA17510-PA [Drosophila pseudoobscura]	318	5e-85
gi 71990269 ref NP_502152.3	Cytochrome P450 family member (caterpillar)	318	6e-85
gi 95102948 gb ABF51415.1	cytochrome P450 CYP4G25 [Bombyx mori]	318	8e-85
gi 71985409 ref NP_496939.2	Cytochrome P450 family member (caterpillar)	318	8e-85
gi 39587668 emb CAE58606.1	Hypothetical protein CBG01773 [Caenorhabditis elegans]	317	1e-84
gi 72001484 ref NP_507688.2	Cytochrome P450 family member (caterpillar)	317	2e-84
gi 93278153 gb ABF06555.1	CYP31B1 [uncultured nematode]	316	2e-84
gi 39580225 emb CAE72981.1	Hypothetical protein CBG20323 [Caenorhabditis elegans]	316	2e-84
gi 33150238 gb AAP97090.1	cytochrome P450 CYP4AB1 [Solenopsis invicta]	315	7e-84
gi 93448327 gb ABC72321.2	cytochrome P450 [Spodoptera littoralis]	313	2e-83
gi 39580246 emb CAE69638.1	Hypothetical protein CBG15879 [Caenorhabditis elegans]	313	2e-83
gi 93278139 gb ABF06548.1	CYP4BE2 [Ips paraconfusus]	311	6e-83
gi 17560320 ref NP_507109.1	Cytochrome P450 family member (caterpillar)	311	8e-83
gi 17542994 ref NP_500637.1	Cytochrome P450 family member (caterpillar)	311	1e-82
gi 27763613 gb AAO20251.1	cytochrome P450 monooxygenase CYP4G19	310	1e-82
gi 108877330 gb EAT41555.1	cytochrome P450 [Aedes aegypti]	309	4e-82
gi 108875660 gb EAT39885.1	cytochrome P450 [Aedes aegypti]	309	4e-82
gi 33518703 gb AAQ20834.1	p450 enzyme precursor [Rhodnius prolixus]	308	9e-82
gi 91081697 ref XP_975951.1	PREDICTED: similar to Cytochrome...	307	1e-81
gi 91093475 ref XP_967939.1	PREDICTED: similar to Cytochrome...	306	3e-81
gi 91082475 ref XP_971963.1	PREDICTED: similar to Cytochrome...	306	3e-81
gi 47779230 gb AAT38513.1	ubiquitous cytochrome P450 [Phyllopertha horticola]	306	3e-81

<u>gi 21552585 gb AAM54722.1 </u>	cytochrome P450 monooxygenase CYP4M6	<u>305</u>	<u>7e-81</u>
<u>gi 24642101 ref NP_573003.2 </u>	Cyp4s3 CG9081-PA [Drosophila mel...]	<u>303</u>	<u>2e-80</u>
<u>gi 17565220 ref NP_503598.1 </u>	CYtochrome P450 family member (c...)	<u>303</u>	<u>2e-80</u>
<u>gi 91081695 ref XP_966683.1 </u>	PREDICTED: similar to Cytochrome...	<u>302</u>	<u>5e-80</u>
<u>gi 17946332 gb AAL49205.1 </u>	RE63964p [Drosophila melanogaster]	<u>301</u>	<u>6e-80</u>
<u>gi 91082459 ref XP_966858.1 </u>	PREDICTED: similar to Cytochrome...	<u>300</u>	<u>1e-79</u>
<u>gi 115534628 ref NP_505009.4 </u>	CYtochrome P450 family member (...)	<u>300</u>	<u>1e-79</u>
<u>gi 66562674 ref XP_625057.1 </u>	PREDICTED: similar to Cytochrome...	<u>300</u>	<u>2e-79</u>
<u>gi 95103020 gb ABF51451.1 </u>	cytochrome P450 [Bombyx mori]	<u>300</u>	<u>2e-79</u>
<u>gi 39591336 emb CAE73389.1 </u>	Hypothetical protein CBG20829 [Caeno	<u>298</u>	<u>5e-79</u>
<u>gi 93278147 gb ABF06552.1 </u>	CYP4BJ1 [Ips paraconfusus]	<u>298</u>	<u>7e-79</u>
<u>gi 54643625 gb EAL32368.1 </u>	GA21527-PA [Drosophila pseudoobscura]	<u>297</u>	<u>1e-78</u>
<u>gi 115647018 ref XP_784930.2 </u>	PREDICTED: similar to cytochrom...	<u>297</u>	<u>2e-78</u>
<u>gi 72098778 ref XP_799260.1 </u>	PREDICTED: similar to cytochrome...	<u>296</u>	<u>2e-78</u>
<u>gi 91090792 ref XP_970404.1 </u>	PREDICTED: similar to Cytochrome...	<u>296</u>	<u>3e-78</u>
<u>gi 9652058 gb AAF91384.1 AF261080_1</u>	P450 CYP319A1 [Boophilus mic	<u>295</u>	<u>6e-78</u>

## APPENDIX D

**Table D1** Residue concentration of chlorpyrifos in treatment water (1 h post treatment).

<b>Specimen</b>	<b>Chorpyrifos contration (µg/l)</b>					
	<b>0</b>	<b>6.81</b>	<b>13.62</b>	<b>27.24</b>	<b>54.48</b>	<b>68.10</b>
Rep1	0.00	1.36	3.37	5.56	14.46	14.76
Rep2	0.00	2.95	3.22	5.48	10.93	12.28
Rep3	0.00	1.75	2.53	12.29	11.71	28.07
Rep4	0.00	2.02	3.04	7.78	12.37	18.37
Rep5	0.00	0.83	0.45	3.91	1.86	8.49
mean	0.00	1.36	3.37	5.56	14.46	14.76
SD	0.00	2.95	3.22	5.48	10.93	12.28

**Table D2** Residue concentration of chlorpyrifos in treatment water (24 h post treatment).

<b>Specimen</b>	<b>Chorpyrifos contration (µg/l)</b>					
	<b>0</b>	<b>6.81</b>	<b>13.62</b>	<b>27.24</b>	<b>54.48</b>	<b>68.10</b>
Rep1	0.00	0.02	0.56	0.38	1.16	1.67
Rep2	0.00	0.06	0.12	0.31	1.19	0.98
Rep3	0.00	0.00	0.26	0.74	0.89	1.47
Rep4	0.00	0.03	0.31	0.48	1.08	1.37
Rep5	0.00	0.03	0.23	0.23	0.16	0.35
mean	0.00	0.02	0.56	0.38	1.16	1.67
SD	0.00	0.06	0.12	0.31	1.19	0.98

**Table D3** Residue concentration of chlorpyrifos in treatment water (48 h post treatment).

<b>Specimen</b>	<b>Chorpyrifos contration (µg/l)</b>					
	<b>0</b>	<b>6.81</b>	<b>13.62</b>	<b>27.24</b>	<b>54.48</b>	<b>68.10</b>
Rep1	0.00	0.00	0.06	0.13	0.49	0.89
Rep2	0.00	0.00	0.04	0.11	0.63	0.38
Rep3	0.00	0.00	0.00	0.24	0.46	0.78
Rep4	0.00	0.00	0.03	0.16	0.53	0.68
Rep5	0.00		0.03	0.07	0.09	0.27
mean	0.00	0.00	0.06	0.13	0.49	0.89
SD	0.00	0.06	0.12	0.31	1.19	0.98

**Table D4** Inhibitory effects of chlorpyrifos on AChE (mean $\pm$ S.D.) in gills of juvenile *P. monodon* at the lethal concentration of chlorpyrifos (30 min post treatment).

Specimen	Chlorpyrifos contration ( $\mu\text{g/l}$ )				
	0	0.0681	6.81	68.1	681
Rep1	4.85128	3.55815	4.34531	3.51016	1.92816
Rep2	3.78904	4.48589	4.90191	3.02807	0
Rep3	6.05623	3.97086	2.06462	1.53839	0.19357
Rep4	4.93096	2.71356	2.97339	2.45759	2.60125
Rep5	2.03864	3.04978	5.55079	1.79295	1.68405
mean	4.33323	3.55565	3.9672	2.46543	1.28141
SD	1.51292	0.7076	1.42582	0.82458	1.13444

**Table D5** Inhibitory effects of chlorpyrifos on AChE (mean $\pm$ S.D.) in gills of juvenile *P. monodon* at the sub-lethal concentration of chlorpyrifos (24 h post treatment).

Specimen	Chlorpyrifos contration ( $\mu\text{g/l}$ )			
	0	0.00681	0.0681	0.681
Rep1	5.34439	6.20337	8.52766	6.90408
Rep2	6.71449	4.40453	2.71462	5.46918
Rep3	5.57484	7.39289	8.94145	3.92896
Rep4	8.62909	6.84910	6.48230	5.07045
Rep5	7.02023	10.94697	4.35934	8.51315
mean	6.65661	7.15937	6.20507	5.97716
SD	1.31514	2.39790	2.67156	1.77241

**Table D6** Inhibitory effects of chlorpyrifos on AChE (mean $\pm$ S.D.) in gills of juvenile *P. monodon* at the sub-lethal concentration of chlorpyrifos (48 h post treatment).

Specimen	Chlorpyrifos contration ( $\mu\text{g/l}$ )			
	0	0.00681	0.0681	0.681
Rep1	2.99903	2.07333	4.46586	1.10501
Rep2	3.91211	2.21492	12.61716	0.82891
Rep3	3.00903	3.17947	1.80544	1.16192
Rep4	4.13912	3.19576	2.82747	1.69885
Rep5	2.18494	23.57416	2.60909	6.46199
mean	3.24885	6.84753	4.86500	2.25134
SD	0.78809	9.36515	4.44007	2.37482

**Table D7** Inhibitory effects of chlorpyrifos on AChE (mean $\pm$ S.D.) in gills of juvenile *P. monodon* at the sub-lethal concentration of chlorpyrifos (72 h post treatment).

Specimen	Chlorpyrifos contration ( $\mu\text{g/l}$ )			
	0	0.00681	0.0681	0.681
Rep1	5.94171	5.60530	2.91295	1.49516
Rep2	2.92083	5.07247	4.65568	1.49997
Rep3	3.57659	0.49330	1.99221	3.33831
Rep4	3.68519	2.91377	1.60298	2.74348
Rep5	4.64339	3.65047	3.00776	1.54238
mean	4.15354	3.54706	2.83432	2.12386
SD	1.17360	2.01869	1.18099	0.86334

**Table D8** Inhibitory effects of chlorpyrifos on AChE (mean $\pm$ S.D.) in gills of juvenile *P. monodon* at the sub-lethal concentration of chlorpyrifos (96 h post treatment).

Specimen	Chlorpyrifos contration ( $\mu\text{g/l}$ )			
	0	0.00681	0.0681	0.681
Rep1	1.55925	2.13577	1.86149	2.21919
Rep2	5.89584	1.14548	2.29556	1.93445
Rep3	5.53862	3.00097	3.01399	1.69486
Rep4	1.64003		2.34469	1.92354
Rep5	3.40132		4.17288	
mean	3.60701	2.09407	2.73772	1.94301
SD	2.06606	0.92845	0.90191	0.21471

**Table D9** DNA tail length ( $\mu\text{m}$ ) (mean  $\pm$  SD) from haemocytes after 1 h of chlorpyrifos exposure.

Specimen	Chlorpyrifos concentration ( $\mu\text{g/l}$ )			
	0	0.007	0.034	0.170
Rep1	9.87	14.45	25.52	20.58
Rep2	7.10	10.50	21.95	21.16
Rep3	6.84	16.70	22.06	24.60
mean	7.94	13.88	23.17	22.11
SD	1.68	3.14	2.03	2.17

**Table D10** DNA tail length ( $\mu\text{m}$ ) (mean  $\pm$  SD) from haemocytes after 6 h of chlorpyrifos exposure.

Specimen	Chlorpyrifos concentration ( $\mu\text{g/l}$ )			
	0	0.007	0.034	0.170
Rep1	10.20	9.60	13.41	21.04
Rep2	7.12	18.71	14.54	21.57
Rep3	14.45	17.29	16.11	21.27
mean	10.59	15.20	14.69	21.29
SD	3.68	4.90	1.35	0.27

**Table D11** DNA tail moment (mean  $\pm$  SD) representing DNA damage after 1 h of chlorpyrifos exposure.

Specimen	Chlorpyrifos concentration ( $\mu\text{g/l}$ )			
	0	0.007	0.034	0.170
Rep1	3.98	3.13	9.74	5.35
Rep2	2.46	2.05	7.61	6.04
Rep3	1.85	4.51	10.78	10.69
mean	2.76	3.23	9.38	7.36
SD	1.09	1.23	1.61	2.90

**Table D12** DNA tail moment (mean  $\pm$  SD) representing DNA damage after 6 h of chlorpyrifos exposure.

Specimen	Chlorpyrifos concentration ( $\mu\text{g/l}$ )			
	0	0.007	0.034	0.170
Rep1	4.90	3.85	4.94	10.14
Rep2	3.32	6.71	4.92	7.86
Rep3	4.43	6.19	6.28	8.57
mean	4.22	5.58	5.38	8.86
SD	0.81	1.53	0.78	1.17

**Table D13** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration (µg/l)				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.69	NA*	NA*	NA*	NA*
Rep2	1.09	NA*	NA*	NA*	NA*
Rep3	0.88	NA*	NA*	NA*	NA*
mean	0.89	NA*	NA*	NA*	NA*
SD	0.20	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D14** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration (µg/l)				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.94	1.47	1.42	1.00	1.18
Rep2	0.99	1.14	1.05	1.13	1.02
Rep3	1.08	0.89	1.27	0.53	0.84
mean	1.00	1.16	1.25	0.88	1.01
SD	0.07	0.29	0.18	0.31	0.17

**Table D15** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration (µg/l)				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.18	1.10	0.10	0.87	NA**
Rep2	0.76	1.03	0.93	0.76	NA**
Rep3	0.92	0.42	0.09	0.65	NA**
mean	0.95	0.85	0.37	0.76	NA**
SD	0.21	0.37	0.48	0.11	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24 µg/l was 100% within 12 h.

**Table D16** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.13	1.20	0.77	1.13	NA**
Rep2	1.09	0.60	0.97	0.97	NA**
Rep3	0.54	1.17	0.93	1.01	NA**
mean	0.58	0.99	0.89	1.04	NA**
SD	0.48	0.34	0.10	0.08	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D17** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.98	1.30	0.93	0.88	NA**
Rep2	1.04	0.74	0.88	1.00	NA**
Rep3	0.70	0.21	0.82	0.98	NA**
mean	0.91	0.75	0.87	0.96	NA**
SD	0.18	0.54	0.06	0.06	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D18** Relative expression level of cytochrome P450 (*CYP4C39*) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.04	0.09	0.22	0.03	NA**
Rep2	0.74	0.95	0.07	0.93	NA**
Rep3	0.88	0.70	0.73	0.82	NA**
mean	0.89	0.58	0.34	0.59	NA**
SD	0.15	0.44	0.34	0.49	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D19** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.06	NA*	NA*	NA*	NA*
Rep2	1.09	NA*	NA*	NA*	NA*
Rep3	1.09	NA*	NA*	NA*	NA*
mean	1.08	NA*	NA*	NA*	NA*
SD	0.02	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D20** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.64	0.94	0.75	0.72	0.82
Rep2	0.78	0.53	0.85	0.79	0.92
Rep3	0.81	0.86	0.94	0.81	0.70
mean	0.74	0.78	0.84	0.77	0.81
SD	0.09	0.22	0.09	0.05	0.11

**Table D21** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.90	0.87	0.94	1.03	NA**
Rep2	1.11	0.82	0.78	1.03	NA**
Rep3	0.89	1.04	1.06	1.08	NA**
mean	0.96	0.91	0.92	1.05	NA**
SD	0.13	0.11	0.14	0.03	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D22** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.03	0.72	1.03	0.87	NA**
Rep2	1.08	1.04	1.18	0.94	NA**
Rep3	0.99	0.80	0.81	0.80	NA**
mean	1.03	0.85	1.01	0.87	NA**
SD	0.04	0.16	0.19	0.07	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D23** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.89	0.92	0.59	0.80	NA**
Rep2	0.96	0.96	0.86	0.86	NA**
Rep3	0.97	0.99	0.96	0.81	NA**
mean	0.94	0.96	0.80	0.82	NA**
SD	0.05	0.03	0.19	0.03	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D24** Relative expression level of beta glucuronidase in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.50	0.78	1.00	0.85	NA**
Rep2	1.13	0.96	1.00	0.75	NA**
Rep3	0.73	0.96	1.01	0.36	NA**
mean	0.78	0.90	1.00	0.65	NA**
SD	0.32	0.11	0.01	0.26	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D25** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.93	NA*	NA*	NA*	NA*
Rep2	1.03	NA*	NA*	NA*	NA*
Rep3	0.97	NA*	NA*	NA*	NA*
mean	0.97	NA*	NA*	NA*	NA*
SD	0.05	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D26** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.18	1.18	1.06	1.30	0.78
Rep2	1.05	1.14	1.07	1.13	1.22
Rep3	1.05	1.06	1.05	1.02	1.11
mean	1.09	1.13	1.06	1.15	1.04
SD	0.07	0.06	0.01	0.14	0.23

**Table D27** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.88	0.95	0.92	0.92	NA**
Rep2	0.86	0.81	0.89	0.91	NA**
Rep3	0.77	0.88	0.86	0.89	NA**
mean	0.84	0.88	0.89	0.91	NA**
SD	0.06	0.07	0.03	0.02	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D28** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.84	0.91	0.89	0.89	NA**
Rep2	0.93	0.88	0.94	0.84	NA**
Rep3	0.87	0.74	0.75	0.80	NA**
mean	0.88	0.84	0.86	0.84	NA**
SD	0.05	0.09	0.10	0.04	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D29** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.14	1.30	1.29	0.92	NA**
Rep2	1.08	1.11	1.15	1.10	NA**
Rep3	1.07	1.17	1.13	1.11	NA**
mean	1.09	1.19	1.19	1.04	NA**
SD	0.04	0.10	0.09	0.11	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D30** Relative expression level of heat shock protein 70 in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.94	0.94	0.92	0.96	NA**
Rep2	0.97	0.88	0.89	0.90	NA**
Rep3	0.91	0.92	0.94	0.82	NA**
mean	0.94	0.91	0.92	0.89	NA**
SD	0.03	0.03	0.03	0.07	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D31** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.87	NA*	NA*	NA*	NA*
Rep2	0.73	NA*	NA*	NA*	NA*
Rep3	0.73	NA*	NA*	NA*	NA*
mean	0.78	NA*	NA*	NA*	NA*
SD	0.08	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D32** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.10	0.18	0.05	0.45	0.05
Rep2	0.47	0.09	0.20	0.54	0.37
Rep3	0.04	0.06	0.04	0.58	0.16
mean	0.21	0.11	0.10	0.52	0.19
SD	0.23	0.06	0.09	0.06	0.16

**Table D33** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.87	0.14	0.47	0.68	NA**
Rep2	0.49	0.14	0.18	0.51	NA**
Rep3	0.15	0.68	0.78	0.66	NA**
mean	0.50	0.32	0.48	0.62	NA**
SD	0.36	0.31	0.30	0.09	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D34** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.85	0.76	0.81	0.62	NA**
Rep2	0.68	0.44	0.78	0.75	NA**
Rep3	0.88	0.62	0.72	0.51	NA**
mean	0.80	0.61	0.77	0.63	NA**
SD	0.11	0.16	0.04	0.12	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D35** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.64	0.07	0.67	0.71	NA**
Rep2	0.73	0.31	0.68	0.70	NA**
Rep3	0.73	0.43	0.08	0.37	NA**
mean	0.70	0.27	0.48	0.59	NA**
SD	0.05	0.18	0.35	0.19	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D36** Relative expression level of heat shock protein 90 in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.41	0.92	0.63	0.73	NA**
Rep2	0.75	0.85	0.85	0.96	NA**
Rep3	0.61	0.77	0.71	0.57	NA**
mean	0.59	0.85	0.73	0.75	NA**
SD	0.17	0.07	0.11	0.20	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D37** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.40	NA*	NA*	NA*	NA*
Rep2	0.02	NA*	NA*	NA*	NA*
Rep3	1.04	NA*	NA*	NA*	NA*
mean	0.49	NA*	NA*	NA*	NA*
SD	0.51	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D38** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.01	0.02	0.02	0.00	0.94
Rep2	1.05	0.01	0.00	0.02	0.00
Rep3	0.70	0.00	0.28	0.00	0.01
mean	0.58	0.01	0.10	0.01	0.32
SD	0.53	0.01	0.15	0.01	0.54

**Table D39** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.06	0.73	0.00	0.05	NA**
Rep2	0.00	0.94	1.03	0.85	NA**
Rep3	1.04	0.19	0.46	0.50	NA**
mean	0.37	0.62	0.50	0.47	NA**
SD	0.59	0.39	0.52	0.40	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D40** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.57	0.81	0.10	0.86	NA**
Rep2	0.91	0.00	0.94	0.01	NA**
Rep3	0.89	0.04	0.03	0.02	NA**
mean	0.79	0.28	0.35	0.30	NA**
SD	0.19	0.45	0.51	0.49	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D41** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.80	1.06	0.02	0.95	NA**
Rep2	0.37	0.75	0.93	0.97	NA**
Rep3	1.00	0.78	0.94	0.32	NA**
mean	0.72	0.87	0.63	0.75	NA**
SD	0.32	0.17	0.53	0.37	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D42** Relative expression level of UBC101C-1,000-D-3 (Esterase) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.24	0.92	1.06	0.90	NA**
Rep2	0.01	0.00	0.82	0.91	NA**
Rep3	1.00	0.91	0.86	0.01	NA**
mean	0.75	0.61	0.91	0.60	NA**
SD	0.65	0.53	0.13	0.51	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D43** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.86	NA*	NA*	NA*	NA*
Rep2	0.83	NA*	NA*	NA*	NA*
Rep3	0.82	NA*	NA*	NA*	NA*
mean	0.84	NA*	NA*	NA*	NA*
SD	0.02	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D44** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.29	0.77	0.94	0.98	0.97
Rep2	1.04	0.76	0.55	1.10	0.88
Rep3	0.95	0.67	0.83	0.81	0.83
mean	0.76	0.73	0.78	0.97	0.89
SD	0.41	0.06	0.20	0.14	0.08

**Table D45** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.03	1.03	0.83	0.95	NA**
Rep2	0.57	1.06	0.94	0.85	NA**
Rep3	1.03	0.81	0.85	1.01	NA**
mean	0.88	0.96	0.87	0.93	NA**
SD	0.26	0.14	0.06	0.08	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D46** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.04	0.89	0.74	1.00	NA**
Rep2	0.94	0.78	0.91	0.85	NA**
Rep3	0.59	0.83	0.83	0.74	NA**
mean	0.52	0.83	0.82	0.86	NA**
SD	0.45	0.06	0.08	0.13	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D47** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.83	0.70	0.88	0.84	NA**
Rep2	1.06	0.86	0.83	1.04	NA**
Rep3	0.85	0.89	0.91	1.12	NA**
mean	0.91	0.82	0.87	1.00	NA**
SD	0.13	0.10	0.04	0.14	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D48** Relative expression level of UBC119A-650-F-5 (CYP330A1) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.62	0.54	0.77	0.20	NA**
Rep2	0.86	0.77	0.66	0.88	NA**
Rep3	0.80	0.37	0.81	0.96	NA**
mean	0.76	0.56	0.75	0.68	NA**
SD	0.12	0.20	0.08	0.42	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D49** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.96	NA*	NA*	NA*	NA*
Rep2	0.45	NA*	NA*	NA*	NA*
Rep3	1.13	NA*	NA*	NA*	NA*
mean	0.85	NA*	NA*	NA*	NA*
SD	0.34	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D50** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.36	0.07	0.63	0.69	0.93
Rep2	0.47	0.71	0.92	0.93	0.63
Rep3	0.94	0.04	0.66	0.08	0.40
mean	0.59	0.27	0.74	0.57	0.65
SD	0.31	0.38	0.16	0.44	0.27

**Table D51** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.85	0.99	0.53	0.49	NA**
Rep2	0.00	1.42	0.97	0.59	NA**
Rep3	0.90	0.26	0.28	0.74	NA**
mean	0.58	0.89	0.59	0.61	NA**
SD	0.51	0.58	0.35	0.12	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D52** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.01	0.66	0.62	0.68	NA**
Rep2	0.82	0.40	0.54	0.16	NA**
Rep3	0.22	0.52	0.10	0.68	NA**
mean	0.35	0.53	0.42	0.51	NA**
SD	0.42	0.13	0.28	0.30	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D53** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.03	0.29	0.13	0.74	NA**
Rep2	0.67	0.67	0.10	0.64	NA**
Rep3	0.20	0.03	0.07	0.13	NA**
mean	0.64	0.33	0.10	0.51	NA**
SD	0.41	0.32	0.03	0.33	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D54** Relative expression level of glutathione-s-transferase in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.34	0.03	0.38	0.01	NA**
Rep2	0.57	0.09	0.01	0.08	NA**
Rep3	0.92	0.42	0.42	0.76	NA**
mean	0.61	0.18	0.27	0.28	NA**
SD	0.29	0.21	0.22	0.41	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D55** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.92	NA*	NA*	NA*	NA*
Rep2	0.68	NA*	NA*	NA*	NA*
Rep3	0.76	NA*	NA*	NA*	NA*
mean	0.79	NA*	NA*	NA*	NA*
SD	0.05	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D56** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.40	0.19	0.22	0.07	0.35
Rep2	0.62	0.23	0.33	0.72	1.39
Rep3	0.61	0.59	0.34	0.53	0.22
mean	0.54	0.34	0.30	0.44	0.65
SD	0.12	0.22	0.07	0.33	0.64

**Table D57** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.45	0.14	0.91	1.07	NA**
Rep2	1.03	0.22	0.68	0.49	NA**
Rep3	0.15	2.25	0.97	0.92	NA**
mean	0.88	0.87	0.85	0.83	NA**
SD	0.66	1.20	0.15	0.30	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D58** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.25	0.13	0.97	0.10	NA**
Rep2	0.23	0.75	0.59	0.61	NA**
Rep3	0.26	0.11	0.20	0.16	NA**
mean	0.24	0.33	0.59	0.29	NA**
SD	0.02	0.36	0.38	0.28	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D59** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.18	0.31	0.15	0.34	NA**
Rep2	0.72	0.80	0.46	0.41	NA**
Rep3	0.62	0.42	0.59	0.06	NA**
mean	0.51	0.51	0.40	0.27	NA**
SD	0.29	0.26	0.22	0.18	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D60** Relative expression level of OPA18G-600-4-1 (Ubiquitin-like-7) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.01	0.31	0.60	0.72	NA**
Rep2	0.57	0.38	0.52	0.02	NA**
Rep3	0.00	0.46	0.56	0.42	NA**
mean	0.19	0.38	0.56	0.39	NA**
SD	0.33	0.08	0.04	0.35	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D61** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.29	NA*	NA*	NA*	NA*
Rep2	0.91	NA*	NA*	NA*	NA*
Rep3	0.97	NA*	NA*	NA*	NA*
mean	1.05	NA*	NA*	NA*	NA*
SD	0.07	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D62** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.11	0.23	0.08	0.30	0.30
Rep2	0.47	0.04	0.15	0.13	0.13
Rep3	0.21	0.12	0.25	0.12	0.12
mean	0.26	0.13	0.16	0.18	0.18
SD	0.19	0.10	0.09	0.10	0.10

**Table D63** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.26	0.07	0.15	0.24	NA**
Rep2	0.11	0.06	0.01	0.24	NA**
Rep3	0.02	0.09	0.23	0.22	NA**
mean	0.13	0.07	0.13	0.23	NA**
SD	0.12	0.02	0.11	0.01	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D64** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.23	0.02	0.16	0.09	NA**
Rep2	0.37	0.60	0.36	0.58	NA**
Rep3	0.46	0.00	0.50	0.11	NA**
mean	0.35	0.21	0.34	0.26	NA**
SD	0.12	0.34	0.17	0.28	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D65** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	1.15	0.83	0.48	0.52	NA**
Rep2	0.51	0.60	0.34	0.51	NA**
Rep3	0.88	0.70	0.56	0.18	NA**
mean	0.85	0.71	0.46	0.40	NA**
SD	0.32	0.12	0.11	0.19	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D66** Relative expression level of OPA01G-415-1 (Leucine zipper protein 5) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ )				
	(N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.14	0.58	0.94	1.10	NA**
Rep2	1.01	0.22	0.46	0.24	NA**
Rep3	0.20	0.34	0.29	0.24	NA**
mean	0.45	0.38	0.56	0.53	NA**
SD	0.48	0.18	0.34	0.50	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D67** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 0 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.25	NA*	NA*	NA*	NA*
Rep2	0.17	NA*	NA*	NA*	NA*
Rep3	0.16	NA*	NA*	NA*	NA*
mean	0.19	NA*	NA*	NA*	NA*
SD	0.02	NA*	NA*	NA*	NA*

Remark: \* data was not available according to 0 h exposure was specifically set for control group.

**Table D68** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 12 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.08	0.00	0.00	0.09	0.00
Rep2	0.01	0.16	0.05	0.00	0.18
Rep3	0.21	0.12	0.00	0.00	0.00
mean	0.10	0.09	0.02	0.03	0.06
SD	0.10	0.08	0.03	0.05	0.10

**Table D69** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 24 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.03	0.03	0.00	0.00	NA**
Rep2	0.05	0.00	0.00	0.00	NA**
Rep3	0.01	0.12	0.04	0.01	NA**
mean	0.03	0.05	0.02	0.00	NA**
SD	0.02	0.07	0.02	0.01	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D70** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 48 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.11	0.00	0.00	0.00	NA**
Rep2	0.09	0.07	0.09	0.04	NA**
Rep3	0.07	0.07	0.00	0.10	NA**
mean	0.09	0.05	0.03	0.05	NA**
SD	0.02	0.04	0.05	0.05	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D71** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 72 h of chlorpyrifos exposure

Time of Exposure (h)	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.45	0.07	0.06	0.24	NA**
Rep2	0.20	0.25	0.25	0.15	NA**
Rep3	0.18	0.28	0.41	0.00	NA**
mean	0.28	0.20	0.24	0.13	NA**
SD	0.15	0.11	0.18	0.12	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

**Table D72** Relative expression level of OPA02G-450-2 (sequence of unknown gene) in hepatopancreas of *P. monodon* after 96 h of chlorpyrifos exposure

Specimen	Chlorpyrifos Concentration ( $\mu\text{g/l}$ ) (N=3)				
	0	0.0681	6.81	13.62	27.24
Rep1	0.04	0.42	0.54	0.01	NA**
Rep2	0.16	0.03	0.53	0.06	NA**
Rep3	0.00	0.20	0.00	0.00	NA**
mean	0.07	0.22	0.36	0.02	NA**
SD	0.08	0.20	0.31	0.03	NA**

Remark: \*\* data was not available according to mortality of shrimp exposed to 27.24  $\mu\text{g/l}$  was 100% within 12 h.

## BIOGRAPHY

Miss Tassanee Eamakmon was born on September 18, 1976 in Chaiyaphume Province, Thailand. She graduated with the degree of Master of Science in Zoology from Chulalongkorn University. She has studied for a degree of doctoral degree of science at faculty of graduate school, Chulalongkorn University.

Publication from this thesis

1. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., Khonsue, W. and Menasveta, P. 2004. Effect of Chlorpyrifos on DNA damage in Haemocyte of Black Tiger Shrimp (*Penaeus monodon*). The 30<sup>th</sup> Congress on Science and Technology of Thailand. Bangkok, Thailand. (Poster presentation on).
2. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2005. Expression of cytochrome P450 gene in Black Tiger Shrimp, (*Penaeus monodon*) exposed to chlorpyrifos. The 31<sup>st</sup> Congress on Science and Technology of Thailand. Nakorn Ratchasima, Thailand. (Oral Presentation).
3. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2006. Toxicity of Chlorpyrifos to Giant Tiger Shrimp, *Penaeus monodon*. International Conference on Environment and Public Health Management: Aquaculture and Environment. Hong Kong Baptist University. Hong Kong. (Oral Presentation).
4. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2006. The Effects of Chlorpyrifos on Acetylcholinesterase Activity in Giant Tiger Shrimp, *Penaeus monodon*. International Conference on Hazardous Waste Management for Sustainable Future. Bangkok, Thailand. (Oral Presentation).
5. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2006. Molecular response of Giant Tiger Shrimp, *Penaeus monodon* exposed to sub-lethal concentration of chlorpyrifos. The 32<sup>nd</sup> Congress on Science and Technology of Thailand. Bangkok, Thailand. (Oral Presentation).
6. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2007. Effects of Chlorpyrifos on Giant Tiger Shrimp, *Penaeus monodon*. The 6<sup>th</sup> National Symposium on Marine Shrimp. Bangkok, Thailand. (Poster presentation on).
7. **Eamkamon, T.**, Puanglarp, N., Thirakhupt, K., and Menasveta, P. 2007. Evaluation of DNA Damage by Single Cell Gel Electrophoresis in the Chlorpyrifos-Exposed Haemocytes of Giant Tiger Shrimp, *Penaeus monodon*. ***Journal of Scientific Research*** 32(1): (in press).