

## REFERENCES



1. Zimmerman, M.R., " Pulmonary and osseous tuberculosis in an Egyptain mummy," Bull. NY. Acad. Med., No. 55, PP. 604-608, New York, 1979.
2. Koch, R., " Die Aeteologie der Tuberculose," Berlin Klin.Wochenschr., 15, 221-230, 1882.
3. Koch, R., " Weitere Mitteilung Uber ein Heilmittel gegen Tuberculose," Dtsch. Med. Wochenschr., 16, 1029-1032, 1890.
4. Hershfield, E.S., " Tuberculosis in the world," Chest, 76, 805-811, 1979.
5. Smith, D.T., " Mycobacterium tuberculosis," Zinsser 's Microbiology (Joklik, W.K., and D.T., Smith, eds.), pp. 445-452, Appleton century Crofts, New York, 15th. ed., 1972.
6. Grange, J.M., " The humoral immune response in tuberculosis," Adv. Tiberc. Res., 21, 1-78, 1984.
7. Runyon, E.H., Karlson, A.G., Kubica, Wayne, L.G., "Mycobacterium," Manual of Clinical Microbiology (Lennette, E.H., Balows, A., Hausler, W., and J.P., Trauant, eds.), pp. 150-174, American Society for Microbiology, Washington D.C., 3rd. ed., 1980.
8. Wolinsky, E., " Mycobacteria," Microbiology (Davis, B.D., Dulbecco, R., Eisen, H.N., and H.S. Ginsberg, eds.), pp. 724-741, Harper International Edition, Pennsylvania, 3rd. ed., 1980.



9. Bloch, H., " Studies on the virulence of tubercle bacilli: Isolation and biological properties of a constituent of virulent organism," J. Exp. Med., 91, 197-217, 1950.
10. Steenken, W., Raleigh, J.W., Smith, M.M., " Pathogenicity of attenuated and avirulent tubercle bacilli," Yale J. Biol. Med., 15, 393-402, 1943.
11. Chaparas, S.D., " Immunity in tuberculosis," Bull. WHO., vol. 60, No. 4, pp. 449-462, 1982.
12. Daniel, T.M., " The immunology of tuberculosis," Clinics Chest Med., 1, 189-201, 1980.
13. Zeitz, S.J., Ostrow, J.H., Van Arsdell, P.P., " Humoral and cellular immunity in the anergy tuberculosis patients," J. Allerg. Clin. Immunol., 53, 20-26, 1974.
14. Wieszorek, Z., Szibinski, G., Zwolinski, J., " Characterization of lymphocyte E-rosette inhibitory factor in sera from tuberculosis patients," Arch. Immunol. Theor. Exp., 25, 63-68, 1977.
15. Zinsser, H., " The studies on the tuberculin reaction and on specific hypersensitiveness in bacterial infection," J. Exp. Med., 34, 495-524, 1921.
16. Lurie, M.B., " Studies on the mechanism of immunity in tuberculosis. The fate of tubercle bacilli ingested

- by mononuclear phagocytes derived from normal and immunized animals," J. Exp. Med., 247-267, 1942.
17. Collins, F.M., " Cellular antimicrobial immunity," CRC Crit. Rev. Microbio., 7, 27-91, 1979.
  18. Collins, F.M., Morrison, N.E., " Restoration of T-cell responsiveness by thymosin. Expression of anti-tuberculosis immunity in mouse lungs," Infect. Immun. 23, 330-335, 1979.
  19. Fox, J.S., Ho, R.S., Aroma, K., Harding, G.E., Smith, D.W., " Host-parasite relationships in experimental tuberculosis. V. Lack of hematogenous dissemination of M. tuberculosis to the lungs in animals vaccinated with BCG," J. Infect. Dis., 137-144, 1976.
  20. Mackaness, G.B., " The induction of expression of cell-mediated hypersensitivity in the lung, Am. Rev. Respir. Dis., 104, 813-828, 1971.
  21. Wing, E.J., Remington, J.S., " Delayed hypersensitivity and macrophages functions, " Basic and Clinical Immunology (Fudenburg, H.H., Sites, D.P., Cardwell, J.L., and J.V. Wells, eds.), pp. 129-143, Lange Medical Publications, California, 3rd ed., 1980.
  22. Cantor, H., Weissman, I., " Development and function of subpopulations of thymocytes and T-lymphocytes," Prog. Allergy, 20, 1-64, 1976.
  23. Rocklin, R.E., Bendtzen, K., Greineder, D., " Mediators

- of immunity; Lymphokines and Monokines," Adv. Immunol., 29, 55-136, 1980.
24. Waksman, B.H., " Overview: Biology of the lymphokines," The biology of lymphokines (Cohen, S. Pick, E., and J.J., Oppenheim, eds.), pp. 585-615, Academic press, New York, 1979.
25. Yoshida, T., Cohen, S., " Biological control of lymphokine function," Feeder. Proc., 41(8), 2480-2483, 1982.
26. Rocklin, R.E., " Mediators of cellular immunity," Basic and clinical Immunology (Fudenburg, H.H., Sites D.P., Cardwell, J.L., and J.V. Wells, eds.), pp. 129-143, Lange Medical Publications, California, 3rd ed., 1980.
27. Rocklin, R.E., " Products of activated lymphocytes: LIF distinct from MIF," J. Immunol., 112, 1461-1466, 1974.
28. Bloom, B.R., " In vitro approaches to mechanism of cell-mediated immunity reaction," Adv. Immunol., 13, 101-208, 1971.
29. Bendtzen, K., " In vitro assay of LIF activity from con A stimulated human lymphokines," Acta. Allergol., 30, 133-149, 1975.
30. Goetzl, E.J., Austen, K.F., " Neutrophil immobilizing factor derived from human leukocytes, J. Exp. Med. , 136, 1564-1580, 1972.

31. Rocklin, R.E., " Partial characterization of LIF by con A stimulated human lymphocytes," J. Immunol., 114, 1161-1165, 1975.
32. Krambovitis, E.,Holzel, H., " Rapid diagnosis of tuberculous meningitis by Latex agglutination," Lancet , ii, 1229-1232, 1984.
33. Gautheir-Rahman, S., " Leukocyte migration test in BCG vaccinated healthy adult by a rapid photoelectric procedure," Clin. Imm. Immunopatho., 41, 75-90, 1986.
34. Orme, I.M., " The kinetics of emergence and loss of mediator T-lymphocytes acquired in response to infection with M. tuberculosis," J. Immunol., 138(1), 293-298, 1987.
35. Berger, H.W., Mejia, E., " Tuberculous Pleurisy," Chest, 63(5), 88-92,1973.
36. Yam, L.T., " Diagnostic significance of lymphocytes in pleural effusions," Ann. Intern. Med., 66, 972-982, 1967.
37. Raabo, E., Rasmussen, K.N., Terkilden, T.C. " A study of isoenzymes of LDH in pleural effusions," Scand. J. Resp. Dis., 47, 150-156, 1966.
38. Falk, A., " Tuberculous pleurisy: diagnosis and results of chemotherapy," Postgrad. Med. 38, 631-635, 1965.
39. Richert, J.H., Wier, J.A., Salyer, J.M., " The reliability of tissue diagnosis of pleurisy," Ann.



Intern. Med., 52, 320-325, 1960.

40. Mestitz, P., Purves, M.J., Pollard, A.C., "Pleural biopsy in the diagnosis of pleural effusion," Lancet, 2, 1349-1353, 1958.
41. Holden, M., Dubin, M.R., Diamond, P.H., "Frequency of negative intermediate-strength tuberculin sensitivity in patients with active TB," N. Engl. J. Med., 285, 1506-1509, 1971.
42. Stead, W.W., Bates, J., "Tuberculosis," Harrison's Principle of Medicine (Thorn, G. W., and R.G., Peterdorf, eds.), p.900, McGraw-Hill, New York, 8th ed., 1977.
43. Ellner, J.J., "Pleural fluid and peripheral blood lymphocytes," Ann. Int. Med., 89(6), 932-933, 1978.
44. Bendixon, G., Sorborg, M., "A leukocyte migration technique for in vitro detection of delayed hypersensitivity in man," Dan. Med. Bull., No. 16, pp. 1-6, Copenhagen, 1969.
45. Harrington, J., Stastny, P., "Macrophage migration from agarose droplet : Development of a micro method for assay of delayed hypersensitivity," J. Immunol., 110, 752-759, 1973.
46. McCoy, J., Dean, J., Herberman, R., "Human cellular immunity to tuberculin as assayed by the agarose micro-droplet technique," Clin. Imm. Immunopatho., 41, 75-90, 1986.

47. Bauer, J.D., " Numerical evaluation of red blood cell and platelets," Gradwohl 's clinical laboratory methods and diagnosis (Frankel, S., and A. Sonnenwirth , eds.), vol. 1, pp. 483-505, CV mosby company , Saint Louis, 7th ed., 1974.
48. Boyum, A., " Isolation of Lymphocytes , Granulocytes and Macrophages," Scand. J. Immunol., 5, 9-15, 1976.
49. Hoffman, T., Kunkel,H., " The E-rosette test" In vitro methods in Cell-mediated and Tumor immunity, (Bloom, B., and J., David, eds.), pp. 71-82. Academic press, New York, 1976.
50. Reiherz, E.L., Kung, P.C., Goldstein, G., Schlossman, S.F., " A monoclonal antibody with selective reactivity with functionally mature human thymocytes and all peripheral human T-cells." J.Immunol., 123, 1312-1317, 1979.
51. Reiherz, E.L., Kung, P.C., Goldstein, G., Schlossman, S.F., " Seperation of functional subsets of human T-cells by a monoclonal antibodies," Proc. Natl. Acad. Sci. USA., 123(6), 2894-2896, 1979.
52. Winchester, R.J., Fu, J.M., " Lymphocyte surface membrane immunoglobulin," Scand. J. Immunol., 5, 77-82, 1976.
53. Hosoda, S., Takase, S., " Non-specific esterase activity in histiocytes," Nature, 190, 927, 1961.
54. Yam, L.T., Li, C.Y., Cosby, W.H.," Cytochemical

- identification of monocytes and granulocytes." Am. J. Clin. Pathol., 55, 283-290, 1971.
55. Koski, I.R., Popack, D.G., Blaese, R.M., " A nonspecific esterase stain for identification of monocytes and macrophages," In vitro methods in Cell-mediated and Tumor immunity, (Bloom, B., and J., David, eds.), pp. 359-362. Academic press New York, 1976.
56. Oppenheim, J.J., Schecter, B., " Lymphocyte transformation " Manual of Clinical Immunology (rose, N.R., and H. Friendman eds.), pp. 233-245, American Society for Microbiology, Washington, D.C., 2nd.ed., 1980.
57. Clausen, J.E., " Tuberculin induced migration inhibition of human peripheral leukocyte in agarose medium," Acta Allergologica, 26, 56-80, 1971.
58. Bendixon, G., Bendtzen, K., Clausen, J.E., Soborg, M., " Human leukocyte migration inhibition" Scand. J. Immunol. , 5(5), 175-184, 1976.
59. Koster, F.T. Mc Gregor, D.D., Maokaness, G.B., " The mediator of cellular immunity: Migration of immunological committed lymphocytes into inflammatory exudates," J. Exp. Med., 133, 400-409, 1971.
60. Koster, F.T., McGregor, D.D., " Lymphocyte traffic from blood into inflamed peritoneal cavity," J. Exp. Med., 133, 846-876, 1971.



61. Jakubusek, P.M., Janicka, G., Zugorecka, A., Pregowski, W., " Leukocyte migration inhibition and rosette tests in pleural effusion," Eur. J. Respir. Dis., 61, 67-70, 1980.
62. Djeu, J.Y., McCoy, J.L., Cannon, G.B., Reeves, W.J., Lymphocyte forming rosette with SRBC in metastatic pleural effusion," J. Natl. Cancer. Insti., 56, 1051-1052, 1976.
63. Petterson, T., Klockars, M., Hellstrom, P., " T and B lymphocyte in pleural effusions," Chest, 73, 49-51, 1978.
64. Donagala, W., Emerson, E.E., Koss, L.G., " Distribution of T and B lymphocyte in blood and effusions of patients with cancer." J. Natl. Cancer. Inst., 61, 295-300, 1978.
65. Domagala, W., Emerson, E.E., Koss, L.G., " T and B lymphocyte enumeration in diagnosis of lymphocyte riched pleural fluid," Acta Cytologica ,25, 108-110, 1981.
66. Falcao, R.P., Bottura, C., " A comparative study of lymphocyte in effusion of tuberculous and malignant disease," Clin. Exp. Immunol., 45, 201-204, 1981.
67. Shimokata, K., Kawakachi, H., Kishimoto, H., Maeda, F., Ito, Y., " Local cellular immunity in tuberculous pleurisy," Am. Rev. Respir. Dis., 126, 822-824, 1982.
68. Shiratsuchi, H., Tsuyuguchi, I., " Analysis of T cell

- subset in patients with tuberculosis after in vitro stimulation with purified protein derivative of tuberculin," Clin.Exp. Immunol., 57,271-278, 1984.
69. Ko, H.S., Fu, S.M., Winchester, R.J., Yu, D., Kunkel, H.G. , " Ia determinants on stimulated human T-lymphocytes," Cell. Immunol., 7, 166-170, 1972.
70. Okubo, Y., Kasama,S., Yono, A., " PPD-specific proliferative cells from tuberculous pleurisy patients and healthy controls with tuberculous pleurisy," Microbiol. Immunol., 26, 511-521, 1982.
71. May, J.J., " The purification and identification of circulating immune complexes in TB," Am. Rev. Respir. Dis., 128(5), 920-925,1982.
72. Simon,M.R., Desai, S.G., Jening, J., Engel, D., " T-cell differentiation antigens and antigenic lymphocyte reactivity in pleural effusions," Asian Pac. J. Aller. Immunol., 4, 19-27, 1986.
73. Groman, G.S., Castele, R.J., Altose, M.D., Scillian, J., Ehlers, R., " Lymphocyte subpopulation in sacoid pleural effusion." Ann. Int. Med., 100, 75-76, 1984.
74. Fujiwara, H., Okuda, Y., Fukukawa, T., Tsuyugushi, I., " In vitro tuberculin reactivity of lymphocyte from patients with tuberculous pleurisy," Infect. Immun. 35, 402-409, 1982.
75. Petterson, T., Welin, M., Weber, T.H., " In vitro production of leukocyte migration inhibition factor



- by lymphocytes in exudative pleural effusions," J. Clin. Lab. Immunol., 8, 107-111. 1982.
76. Rook, G., " The immunological consequence of antigen overload in experinemtal infection mice," Clin. Exp. Immunol., 19, 167-178, 1975.
77. Kanton, F., " Infection ,anergy and cell-mediated immunity," N. Engl. J. Med., 292, 629-634, 1975.
78. Meuer, S.C., Hodgdon, J.C., Cooper, A.D., " Human cytotoxic T cell clones directed at autologous virus-transformed targets: further evidence for linkage of genetic restric tions to T4 and T8 surface glycoprotiens," J. Immunol., 131, 186-190, 1983.
79. Zatz, M., Lance, E., " The distribution of 51 Cr-labelled lymphocyte into antigen stimulated mice," J. Exp. Med., 134, 224-241, 1971.
80. Roaley, D., Gowan, J., Atkins, R., Ford, W., Smith, M., " The specific selection of recirculating lymphocytes by antigen in normal and preimmunized rats," J. Exp. Med., 136, 499-513, 1972.
81. Rosenstreich, D, Blake, T., Rosental, A., " The peritoneal exudate lymphocyte. Difference in antigen responsesiveness between peritoneal exudate and lymphocytes from immunized guinea pigs," J. Exp. Med., 134, 1170-1186, 1971.
82. Rook, G., Carswell, J., Stanford, J., " Preliminary

- evidence for the trapping of antigen specific lymphocytes in the lymphoid tissue of anergic tuberculosis patients," Clin. Exp. Immunol., 26, 129-132, 1976.
83. Stead, W., Bates, J., "Tuberculosis" Harrison's Principles of Internal Medicine (Thorn, G., Adams, R., Braunwald, E., Isselbacher, K., Petersdorf, R., eds.), p. 900, McGraw-Hill, New York, 8th ed. 1977.
84. Acuto, o., Reinherz, E., "The human T-cell receptor: structure and funtion," N. Engl. J. Med., 312, 1100-1111, 1985.
85. Lew, D., Perrin, L., Vassalli, D., Lambert, P. "High levels of complement brakedown products in tuberculous pleural effusion," Clin. Exp. Immunol., 52, 569-574, 1983.
86. Poryhus, A., Steinmann, G., Stein, E., Mertelsmann, R., "T and B cell responses in patients with malicnant pleural effusions," Br. J. Cancer., 43, 471-477, 1981.
87. Cohen, S., Ward, P., Yoshida, T., Burek, C., "Biological activity of delayed hypersensitivity skin reaction sites," Cell. Immunol., 9, 363-376, 1973.
88. Stastny, P., Rosenthal, M., Andreis, M., Ziff, M., "Lymphokines in the reumatic joints," Arthritis Rheum., 18, 237-241, 1975.
89. Kinnman,, Fryden, A., Eriksson, S., Moller, E., Link,



- H., " Tuberculous meningitis : immune reaction with  
in the central nervous system," Scand. J. Immunol.,  
13, 289-296, 1981.
90. Rocklin, R., " The leukocyte migration inhibition test  
and leukocyte adherancing factor test," Mannaul of  
Clinical Immunology ( Rose, N.R., and H.Friendman,  
eds.), pp278-295, American Society for Microbiology,  
Washington D.C., 2nd. ed., 1980.
91. Falk, A., " Tuberculous pleurisy with effusion,  
diagnosis and result of chemoterapy," Postgrad.  
Med., 38 ,631-635, 1965.
92. Mestitz, P., Purves, M.J., Pollard, A.C., " Pleural  
biopsy in the diagnosis of pleural effusion.  
Areport of 200 cases, " Lancet, 2, 1349, 1958.
93. Schools, G.S., " Needle biopsy of the parietal pleura  
: current status," Texas J. Med., 59, 1056, 1963.



APPENDIX I

CHEMICAL AGENTS AND INSTRUMENTS

A. Chemical substances.

Agarose type III :high EEO ( Sigma, Mo., USA. )

Absolute methanol ( CH OH ) ( Mallinckrodt, Paris, France

Ammonium chloride ( NH Cl ) ( May & Baker, Dagenham, UK. )

Acetone ( C H O ) ( E.merck, Darmstadt, W.Germany )

$\alpha$ -naphthylbutyrate N-8000 ( Sigma, Mo. USA. )

Citric acid ;AR grade ( C H O .H O ) ( E.merck, Darmstadt

W.Germany )

di-sodium hydrogen phosphate ( Na HPO ) ( May & Baker  
Dagenham, UK. )

Dextran T500 ( Phamacia Fine Chemicals, Upsala, Sweden )

Ficoll; M.W. 400,000 ( Sigma, Mo., USA. )

Formaldehyde ( Riedel, Hannover, W.Germany )

Fluted filter paper; No.1 ( Whatman, N.J., USA. )

Glacial acetic acid ( CH COOH ) ( E.merck, Darmstadt

W.Germany )

Glass fiber filter paper No.934-AH ( Whatman, N.J., USA. )

Glucose ( C H O ); AR. grade ( BDH, Poole, UK. )

Glycerine ( E.merck, Darmstadt, W.Germany )

Heparin 5000 I.U./ml.;sterile for injection. ( Leo  
Bellerup, Denmark )

Hypaque sodium 50 %, diatrizoate sodium injection.

(Winthrop, N.Y., USA. ).

- HEPES ( N-2-hydroxythypiperazine-N-2-ethanesulfonic acid  
 ( Sigma, Mo., USA. )
- Hydrochloric acid ( HCl )( E.merck, Darmstadt, W.Germany )
- Hank 's balance salt solution ( HBSS ); without NaHCO<sub>3</sub>  
 ( Gibco; Grand Island, N.Y., USA. )
- Methyl green ( BDH, Poole, UK. )
- Millipore membrane filter, pore size 0.22 Um ( Millipore  
 Ma., USA. )
- N,N-dimethyl formamide, spectrophotometer grade. ( C H NO  
 3 7  
 ( Mallinckrodt, Paris, France )
- Oil immersion ( E.merck, Darmstadt, W.Germany )
- Penicillin G; 1,000,000 Units/vial ( Dumex, Bangkok  
 Thailand )
- Paraformaldehyde ( Sigma, Mo., USA. )
- Pararosaniline HCl; No. P 3750 ( Sigma, Mo. USA. )
- Purified protein derivative RT23 (PPD), stock solution  
 5,000 TU/ml. or 1 mg/ml. with 0.01% chinisol. (statens  
 seruminstitut, Copenhagen, Denmark)
- Phytohemagglutinin (PHA M-form). (GIBCO;Grand Island,  
 N.Y., USA.)
- PPO (2,5-Diphynenylloxazole). (Sigma,Mo., USA.)
- POPOP [p-bis(2-(5-phynyloxazolyl)-benzene)], Scintillation  
 grade. (NEN., Edinburgh, Scotland)
- Picric acid (E.merck, Damstadt, W.Germany)
- Permout (Fisher Scientific company, N.J., USA.)

Potassium di hydrogen phosphate ( $\text{KH}_2\text{PO}_4$ ). (E.Merck, Darmstadt, W.Germany)

RPMI 1640 (Rosewell Park Memorial Institute formular 1640), with L-glutamine, without antibiotics. (GIBCO;Grand Island, N.Y. USA.)

Sodium di hydrogen phosphate ( $\text{NaH}_2\text{PO}_4$ ). (May&Baker Dudenham, UK.)

Sodium bicarbonate ( $\text{NaHCO}_3$ ), AR grade. (BDH, Poole, UK.)

Sodium citrate dihydrate ( $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$ ). (May&Baker, Dagenham, UK.)

Sodium chloride ( $\text{NaCl}$ ). (E.Merck, Darmstadt, W.Germany)

Sodium nitrite ( $\text{NaNO}_2$ ) (BDH, Poole, UK )

Sodium hydroxide ( $\text{NaOH}$ ). (E.merck, Damstadt, W.Germany)

Streptomycin sulfate (Dumex, Bangkok, Thailand)

Tritiated thymidine (methyl- $^3\text{H}$ ), sterile aqueous solution, 75 GBq/ mM. (Amersham, Amersham, UK.)

Trypan blue (BDH, Poole, UK.)

Toluene ( $\text{C}_7\text{H}_8$ ), Scintillation grade. (E.merck, Damstadt, W.Germany)

Tetrasodium EDTA [ $\text{C}_{10}\text{H}_{16}\text{N}_2\text{O}_8$ ]. (E.merck, Darmstadt, W.Germany)

Wright stain (E.merck, Darmstadt, W.Germany)



## B. Antiserum and serum.

Anti-human immunoglobulin polyvalent (IgG,A,M) fluorescein labelled from sheep. (Wellcome reagent, Beckenham, UK.)

Horse serum (Flow lab., Ayrshire, Scotland)

OKT4 & OKT8 monoclonal antibodies. (Ortho diagnostic, N.J., USA.)

Rabbit anti-mouse immunoglobulin FITC conjugated (DAKO Igs., Glostrup, Denmark)

## C. Glassware.

Disposable polystyrene tube conical base, 110 X 17 mm., and round base, 100 X 15 mm., with stopper. (Nunc, Roskilde, Denmark)

Erlenmeyer flask with screw cap lid, capacity 125 ml. (Kimble, Kimax, Ohio, USA.)

Glass tube with screw cap lid, size 16 X 125 mm. (Kimble, Kimex, Ohio, USA.)

Plastic tissue culture dishes, diameter 60 X 15 mm. with lid. (Nunc, Roskilde, Denmark)

Serological pipet (Pyrex, Corning, N.Y., USA.)

Sterile flat bottom 96 wells micro-plate with lid. (Nunc, Roskilde, Denmark)

D. Instrument.

Automatic pipet. (Gilson, Lyon, France)

Automatic microcell harvester, model CH 103 (Dynatech, Sasex, UK.)

$\beta$ -counter, model LS 100 C (Beckman, CA., USA.)

CO incubator (Forma Scientific, Ohio, USA.)

<sup>2</sup>Fluorescence microscope, model BH (Olympus, Tokyo, Japan)

Gel puncher 3 mm. diameter (Gelman, Michigan, USA.)

Halminton syringe, microliter (Halminton, Bonaduz, Switzerland)

Light microscope, model BH (Olympus, Tokyo, Japan)

PH meter, model 10 (Corning, N.Y., USA.)

Refrigerated centrifuge, Model Certra 7-R (IEC, Boston, Ma., USA.)

Vacuum pump (Doerr, W.I., USA.)

Waterbath ( Precision Scientific, Chicago, USA.)



## APPENDIX II

### REAGENTS AND PREPARATIONS

1. Reagents for white blood cell count and differential cell count and method for blood smear stained.

1.1 White blood cell diluent

Glacial acetic acid	3	ml.
Distilled water (DW)	100	ml.

1.2 Wright's stain solution

Wright's stain	0.1	gm.
Absolute methanol (acetone free)	60	ml.

Keep in the dark at room temperature.

1.3 Phosphate buffer solution for Wright's stain

KH PO <sub>2</sub> 4	5.13	gm.
Na PO <sub>2</sub> 4	4.12	gm.
Add DW to	1	liter

Adjust pH to 6.7

The blood smear was flood with Wright 's stain solution for 3 mins and added phosphate buffer saline solution for 3 mins. Then, the blood smear was washed with ranning tap water and let it air dry.



## 2. Reagent for mononuclear cell preparation.

### 2.1 Ficoll-Hypaque solution

#### 2.1.1 9% Ficoll

Ficoll                    9    gm.

DW                        100   ml.

Sterile by autoclave

#### 2.1.2 33.9% Hypaque

50% Hypaque                33.9   ml.

DW                        16.1   ml.

9% Ficoll was mixed with 33.9% Hypaque in the ratio 2.4:1 respectively. This solution should have specific gravity about 1.077

### 2.2 Hank's balance salt solution (HBSS).

One case of HBSS without  $\text{NaHCO}_3$  was added with  $\text{NaHCO}_3$  0.35 gm. and DW 1 liter. After adjusted pH to 7.4 with 1 M. NaOH or 1 M. HCl, HBSS was sterilized by filtration with 0.22  $\mu\text{m}$ . millipore membrane.

### 2.3 RPMI 1640

One case of RPMI 1640 was added with  $\text{NaHCO}_3$  2 gm. and DW to 1000 ml. After adjusted pH to 7.4 with 1 M. NaOH or 1 M. HCl, it was sterilized by filtration with 0.22  $\mu\text{m}$ . membrane filter.

## 2.4 Penicillin 10,000 Units/ml.

## 2.4.1 Stock penicillin 100,000 Units/ml.

Penicillin G 1,000,000 Units per  
ampule was reconstituted with sterile DW 10 ml. and mixed.

## 2.4.2 Working penicillin 10,000 Units/ml.

Stock penicillin 100,000 Units/ml.

0.1 ml.

RPMI 1640

0.9 ml.

## 2.5 Streptomycin 10,000 Ug/ml.

## 2.5.1 stock streptomycin 10,000 Ug/ml.

Streptomycin 1 mg. was reconstituted  
with sterile DW 10 ml. and mixed.

## 2.5.2 Working streptomycin 10,000 Ug/ml.

Stock streptomycin 100,000 Ug/ml.

0.1 ml.

RPMI 1640

0.9 ml.

## 2.6 1 M. HEPES

HEPES 23.82 gm.

DW 100 ml.

Sterile by autoclave

## 2.7 Tissue culture medium (TCM).

RPMI 1640	9.2 ml.
Heat inactivated pool human AB serum	5 ml.
Penicillin G 10,000 Units/ml.	1 ml.
Sterptomycin 10,000 Ug/ml.	1 ml.
1 M. HEPES	1 ml.
sterile by filtration	

## 3. Reagent for spontaneous E-rosette formation

## 3.1 Modified Alsever's solution

Glucose	24.6 gm.
Sodium citrate	9.6 gm.
NaCl	5.04 gm.
DW	1200 ml.

Adjusted pH to 6.1 with citric acid and sterile by filtration.

## 3.2 Sheep red blood cell (SRBC) collection

Peripheral blood of sheep from jugular venepuncture were resuspended in sterilized modified Alsever solution in the ratio 4:1 respectively and mixed, store at 4 C.



### 3.3 Phosphate buffer saline (PBS) pH 7.4

#### 3.3.1 Solution A

NaH PO <sub>2</sub> .H O	27.6 gm.
2 4 2	

#### 3.3.2 Solution B

Na HPO <sub>2</sub> .12 H O	71.63 gm.
2 4 2	
DW	1000 ml.

#### 3.3.3 PBS pH 7.4

Solution A	16.5 ml.
Solution B	33.5 ml.
NaCl	8.5 ml.
DW to	1,000 ml.

Adjusted pH to 7.4 and sterile by autoclave.

### 3.4 1% SRBC.

SRBC suspension were washed 3 times with PBS pH 7.4 by centrifuged at 300 G for 5 mins. Pack SRBC 0.1 ml. were resuspended in 9.9 ml. of HBSS and mixed. 1% SRBC was freshly prepared before used.

## 4. Reagents for T-cell subset determination.

### 4.1 OKT monoclonal antibodies.

Lyophilized form of OKT4 or OKT8 monoclonal antibodies (Ortho diagnostic, N.J., USA.) were reconstituted with 1 ml. of sterile distilled water. A portion of 100 U1. were aliquated and kept at -70 °C .



After thawing, these antibodies for use were stored at 4 C.

4.2 Rabbit anti-mouse immunoglobulin FITC conjugated.

Rabbit anti mouse immunoglobulin fluorescein labelled (DAKO, Glostrup, Denmark) were diluted to 1:20 with sterile PBS pH 7.4 and kept in the dark at 4 C.

4.3 1% paraformaldehyde solution

4.3.1 Stock 10% paraformaldehyde

Paraformaldehyde 1 gm.

PBS pH 7.4 100 ml.

4.3.2 working 1% paraformaldehyde

10 % paraformaldehyde 1 ml.

PBS pH 7.4 10 ml.

4.4 mounting media

PBS pH 7.4 was mixed equal volume with glycerine.

5. Reagent for surface membrane immunoglobulin-FITC conjugated.

Anti-human immunoglobulin FITC conjugated (poly valent specific for Ig G,A,M) was diluted to 1:20 with sterile PBS pH 7.4 and stored in the dark at 4 C.

## 6. Reagents for nonspecific esterase enzyme staining.

## 6.1 Fixative solution.

Na HPO	20	mg.
2  4		
KH PO	100	mg.
2  4		
DW	30	ml.
acetone	45	ml.
30% formaldehyde	25	ml.

Mixed and adjusted to pH 6.6 . Keep in refrigerator.

## 6.2 Pararosaniline solution.

Pararosaniline HCl	1	gm.
2 N HCl	25	ml.

Store in refrigerator.

## 6.3 4% sodium nitrite solution.

NaNO	100	mg.
2		
DW	2.5	ml.

freshly prepared.

## 6.4 M/15 sorensen 's phosphate buffer (pH 6.3)

Na HPO	2.128	gm.
2  4		
KH PO	6.984	gm.
2  4		
DW	1000	ml.

6.5  $\alpha$ -naptyl butyrate solution.

$\alpha$ -naptyl butyrate	1	gm.
dimethyl formamide	50	ml.

Mixed reagents in glass bottle. Keep solution in freezer and protect from light.

6.6 0.5% methyl green.

Methyl green	500	mg.
DW	100	ml.



Store in refrigerator and filter before used.

7. Reagents for lymphocyte transformation test.

7.1 Tissue culture medium (TCM).

RPMI 1640	9.2	ml.
Heat inactivated pool human AB serum	5	ml.
Penicillin G 10,000 Units/ml.	1	ml.
Sterptomycin 10,000 Ug/ml.	1	ml.
1 M. HEPES	1	ml.

sterile by filtration

7.2 Purified protein derivative (PPD).

PPD 1 mg./ml. was placed in sterile dialysis membrane tubing and dialyzed in sterile PBS pH 7.4 over night at 4 C with 3 times changed of PBS. This preservative free PPD was aliquated in sterile vial and stored in the dark at -70 C.

### 7.3 Phytohemagglutinin (PHA).

Lyophilized form of PHA were reconstitute with sterile DW 10 ml. The stock PHA solution was titrated for dose response curve before used and stored at -70° C.

### 7.4 Tritiated thymidine solution (<sup>3</sup>HTdR).

(methyl-<sup>3</sup>H) thymidine, sterile aqueous solution 1 mci/ml. was diluted with 4.9 ml. of TCM to make the final concentration to 20 Uci/ml. 25  $\mu$ l. of 20 Uci/ml. thymidine were dropped in a well of cell culture to give the final concentration of thymidine to 0.5 Uci/ml.

### 7.5 Scintillation fluid.

PPO	5 gm.
POPOP	0.1 gm.
Toluene	1 liter

## 8. Reagents for leukocyte migration inhibition test.

### 8.1 6% dextran in NSS.

Dextran T500	6 gm.
NaCl	0.85 gm.
DW	100 ml.

Mixed to dissolve and sterilized by autoclave.

8.2 Penicillin 10,000 Units/ml. and streptomycin 10,000 Ug/ml.

Stock Penicillin 100,000 Units/ml.	0.1 ml.
Stock streptomycin 100,000 Ug/ml.	0.1 ml.
Sterile DW	0.8 ml.

8.3 TC 199 medium.

TC 199, 10x	1 ml.
Heat inactivated horse serum	1 ml.
Penicillin/streptomycin solution	0.1 ml.
1 M HEPES	0.1 ml.
Sterile DW	7.8 ml.

Adjusted to pH7.4 with 10% NaHCO<sub>3</sub>. Steriled by filtration.

8.4 TC 199 agarose medium.

The TC 199 agarose medium contained 0.8% agarose, 1 X of TC 199, 10% horse serum, Penicillin G 100 Units/ml., streptomycin 100 Ug/ml. and 10 mM HEPES.

8.4.1 1.6% stock agarose medium.

Agarose (type III)	1.6 gm.
DW	100 ml.

Melt agarose by heat on boiling bath and used 5 ml. of melted agarose allowed to cool at 50 C water bath.





#### 8.4.2 2 X TC 199 medium.

TC 199, 10 X            1 ml.

Heat inactivated horse serum 1 ml.

Penicillin/sterptomycin    0.1 ml.

1 M HEPES                0.1 ml.

Sterile DW                2.8 ml.

Adjusted to pH 7.4 with 10% NaHCO<sub>3</sub> and warm at 50 °C in water bath. Then, 5 ml. of 1.6% melted agarose were mixed equal volume with warm 2 X TC 199 medium at 50 °C and 5 ml. of melted TC 199 agarose medium were placed in 60 X 15 mm. sterile plastic dish, allowed to cool. The agarose medium was cut holes with 3 mm. gel puncher and made the holes by vacuum pump. The agarose plate was stored in the moist chamber.

#### 8.5 Fixing reagent.

##### 8.5.1 Saturated picric acid solution.

Picric acid            20 gm.

DW                    800 ml.

##### 8.5.2 Fixing reagent.

Saturated picric acid solution 100 ml.

Glacial acetic acid            20 ml.

## 8.6 Lysing solution.

NH Cl            8.26 gm.  
    <sub>4</sub>  
KHCO            1 gm.  
    <sub>3</sub>  
EDTA, tetra sodium 0.037 gm.  
DW to            1000 ml.

## 9. Reagent for trypan blue exclusion.

0.5% trypan blue dye : trypan blue 0.5 gm.  
dissolved in NSS 100 ml. and filtrated before used.

Cell suspensions 10 ul. were mixed with 80 Ul. of NSS and 10 Ul. of 0.5% trypan blue. These cell suspensions were spraded on hemocytometer and counted under light microscope within 5 mins. after cell mixing. The viable cells were reflected the light and no color, but the dead cells were stained blue color.



## CURRICULUM VITAE

Mr. Wattana Panmoung was born on May 28, 1958 in Bangkok, Thailand. He graduated with B.Sc. in Medical Technology (2nd. class honors) from Chulalongkorn University in 1979. His academic position is Faculty member of Immunology Unit, Department of Microbiology, Faculty of Medicine, Chulalongkorn University. On August 21-27, 1983, he participated in 5th International Congress of Immunology in Kyoto, Japan with his presentation "Suppressor T-cells in Thai patients with systemic lupus erythematosus". In addition, he was trained in the production and characterization of monoclonal antibodies from the Institute for General and Experimental Pathology, University of Innsbruck, Innsbruck, Austria, between October 1985 - June 1986. During 7 years of his occupational experience, he have had publications:

1. Phanuphak, P., Ratanavongsiri, J., Panmoung, W., Trongkawad, P., Aphaiwong, O., Chawanasai, A., Ooneglap, T., " Socioeconomy and pattern of drug abuse in heroin and opium addicts," Chula. Med. J., 26(4), 253-265, 1982.
2. Phanuphak, P., Tirawatpong, S., Hanvanich, M., Panmoung, W., Vejjajiva, S., Mollar-or, P., Sitprija, V., Intaraprasert, R., Phantumkosol, D., " Autoantibodies in falciparum malaria: a sequential study in 183 Thai patients," Clin. Exp. Immunol., 53, 627-633, 1983.

3. Ratanavongsiri, J., Panmoung, W., Phanuphak, P., " Immunologic studies of heroin addicts," Asain Pac. J. Aller. Immunol., 2(2), 195-199, 1985.
4. Dhamabutra, N., Panmoung, W., " Resent progress: Laboratory diagnosis of Clamidial infections," Chula. Med. J., 29(6), 1985.
5. Phanuphak, P., Locharererkul, C., Panmoung, W., Wilde, H., " A report of three cases of AIDS in Thailand," Asian Pac. J. Aller. Immunol., 3,195-199, 1985.
6. Panmoung, W., Dhamabutra, N., " The production and application of monoclonal antibodies," Chula. Med. J., 30(12), 1179-1191, 1986.

