

REFERENCES

1. Alpide-Tendencia, E. V.; Dureza, L. A. Isolation of *Vibrio* spp. From *Penaeus monodon* (Fabricius) with Red Disease Syndrome *Aquaculture* 154(1997): 107-114.
2. Bassler, B. L.; Greenberg, E. P.; Stevens, A. M. Cross-Species Induction of Luminescence in Quorum Sensing Bacterium *Vibrio harveyi* *Journal of Bacteriology* 1997: 4043-4045.
3. Karunasagar, I.; Otta, S. K.; Karunasagar, Indrani Biofilm Formation by *Vibrio harveyi* on Surfaces *Aquaculture* 140(1996): 241-245.
4. Phianphak W.; Rengpipat S.; Rukpratanporn S.; Longyant S.; Chaivisuthangkura P.; Sithigorngul W.; Sithigorngul P. Production of Monoclonal Antibodies for Detection of *Vibrio harveyi* *Diseases of Aquatic Organisms* 63(2-3)(2005): 161-168.
5. Robertson, P. A .W.; Xu, H. S.; Austin, B. An Enzyme-Linked Immunosorbent Assay (ELISA) for the Detection of *Vibrio harveyi* in Penaeid Shrimp and Water *Journal of Microbiological Methods* 34(1998): 31-39.
6. Thomulka, K. W.; Lange, J. H. Use of the Bioluminescent Bacterium *Vibrio harveyi* to Detect Biohazardous Chemicals in Soil and Water Extractions With and Without Acid *Exotoxicology and Environmental Safety* 32(1995): 201-204.
7. Vinod, M. G.; Shiju, M. M.; Umesha, K. R.; Rajeeva, B. C.; Krohne, G.; Karunasagar, Indrani; Karunasagar, Iddya. Isolation of *Vibrio harveyi* Bacteriophage with a Potential for Biocontrol of Luminous Vibriosis in Hatchery Environments *Aquaculture* 2005 (article in press).
8. www.health.enotes.com/medicine-encyclopedia/vibriosis.htm

9. Lee, K. -K.; Chen, Y. -L.; Liu, P. -C. Hemostasis of Tiger Prawn *Penaeus monodon* Affected by *Vibrio harveyi*, Extracellular Products and a Toxic Cysteine Protease *Blood Cells, Molecules, and Diseases* 25(13)(1999): 180-192.
10. www.absoluteastronomy.com/reference/bioluminescence.htm
11. Herring, P. J. Bioluminescence in Action NY, Academic Press: 1978.
12. Ulman, A. An Introduction to Ultrathin Organic Films: From Langmuir-Blodgett to Self-Assembly San Diego, Academic Press: 1991.
13. Kima, H. J.; Kwak, S.; Kim, Y. S.; Seo, B. I.; Kim, E. R.; Lee, H. Adsorption Kinetics of Alkanethiols Studied by Quartz Crystal Microbalance *Thin Solid Films* 327-329(1998): 191-194.
14. Shen, G.; Wang, H; Tan, S; Li, J.; Shen, Guoli; Yu, R. Detection of Antisperm Antibody in Human Serum using a Piezoelectric Immunosensor based on Mixed Self-Assembled Monolayers *Analytica Chimica Acta* 540(2005): 279-284.
15. Fung, Y. S.; Wong, Y. Y. Self-Assembled Monolayers as the Coating in a Quartz Piezoelectric Crystal Immunosensor to Detect *Salmonella* in Aqueous Solution *Analytical Chemistry* 73(21)(2001): 5302-5309.
16. Su, X. -L.; Li, Y. A Self-Assembled Monolayer-based Piezoelectric Immunosensor for Rapid Detection of *Escherichia coli* O157:H7 *Biosensor and Bioelectronics* 19(2004): 563-574.
17. Jr, R. C.; Lee, T. R. Thiol-based Self-Assembled Monolayers: Formation and Organization *Encyclopedia of Materials: Science and Technology* 2001: 9332-9344.
18. Park, I. S.; Kim, D. K.; Adanyi, N.; Varadi, M.; Kim, A. Development of a Direct-Binding Chloramphenical Sensor based on Thiol or Sulfide Mediated

- Self-Assembled Antibody Monolayer *Biosensor and Bioelectronics* 19(2004): 667-674.
19. Shen, G.; Wang, H.; Tan, S.; Li, J.; Yu, R. Improved Method for the Preparation of Carboxylic Acid and Amine Terminated Self-Assembled Monolayers of Alkanethiolates *Langmuir* 21(7)(2005): 2633-2636.
 20. Du, W. Electrostatic Self-Assembly of Biocompatible Thin Films Thesis submitted to the Faculty of Virginia Polytechnic Institute and State University 2000.
 21. Colorado, R.; Villazana, R. J.; Lee, T. R. Self-Assembled Monolayer on Gold Generated from Aliphatic Dithiocarboxylic Acids *Langmuir* 14(1998): 6337-6340.
 22. Smith, R. K.; Lewis, P. A.; Weiss, P. S. Patterning Self-Assembled Monolayers *Progress in Surface Science* 75(2004): 1-68.
 23. Chechik, V.; Crooks, R. M.; Stirling, C. J. M. Reaction and Reactivity in Self-Assembled Monolayer *Advanced Materials* 12(16)(2000): 1161-1171.
 24. Ulman, A.; Eilers, J. E.; Tillman, N. Packing and Molecular Orientation of Alkanethiol Monolayers on Gold Surfaces *Langmuir* 5(1989): 1147-1152.
 25. Walczak, M. M.; Chung, C.; Stole, S. M.; Widrig, C. A.; Porter, M. D. Structure and Interfacial Properties of Spontaneously Adsorbed n-Alkanethiolate Monolayers on Evaporated Silver Surfaces *Journal of American Chemical Society* 113(1991): 2370-2378.
 26. Bain, C. D.; Troughton, E. B.; Tao, Y. T.; Evall, J.; Whitesides, G. M.; Nuzzo, R. G. Formation of Monolayer Films by the Spontaneous Assembly of Organic Thiols from Solution onto Gold *Journal of American Chemical Society* 111(1989): 321-335.
 27. Nuzzo, R. G.; Dubois, L. H.; Allara, D. L. Fundamental Studies of Microscopic Wetting on Organic Surface. I. Formation and Structural Characterization of a

- Self-Consistent Series of Polyfunctional Organic Monolayer *Journal of American Chemical Society* 112(1990): 558-569.
28. Laibinis, P. E.; Whitesides, G. M.; Allara, D. L.; Tao, Y. T.; Parikh, A. N.; Nuzzo, R. G. Comparison of the Structures and Wetting Properties of Self-Assembled Monolayers of n-Alkanethiols on the Coinage Metal Surface, Cu, Ag, Au *Journal of American Chemical Society* 113(1991): 7152-7167.
 29. Sellers, H; Ulman, A.; Shnidman, Y.; Eilers, J. E. Structure and Binding of Alkanethiolates on Gold and Silver Surfaces: Implications of Self-Assembled Monolayers *Journal of American Chemical Society* 115(1993): 9389-9401.
 30. Ulman, A. Formation and Structure of Self-Assembled Monolayers *Chemical Reviews* 96(1996): 1533-1554.
 31. www.biology.arizona.edu/immunology/tutorials/antibody/structure.html
 32. www.pathology.wustl.edu/htm/facilitieshybridaddisad.htm
 33. Sears, D.W. Antibody-Antigen Interactions 1997, Available from : www.whfreeman.com/immunology/CH06/kuby06.htm
 34. Lesk, A. M. Introductin to Protein Science : Architecture, Function and Genomics New York, Oxford University Press: 2004.
 35. Siuzdak, G. Mass Spectrometry for Biotechnology Sandiego, Academic Press: 1996.

APPENDIX

Self-assembly Monolayer (SAM) Formation of Carboxyl-terminated Alkanethiol onto Gold Surface.

Table A-1 Water contact angle and frequency shift due to SAM formation (Δf_s) of 10 mM MPA as a function of time.

Time (min)	Water Contact angle (degree)	Δf_s (Hz)
Gold	103.6±3.9	-
8	43.4±2.1	4
16	40.6±2.5	12
24	35.2±1.9	19
48	35.4±2.1	20

Table A-2 Frequency shift due to SAM formation of each alkanethiol (Δf_s) as a function of concentration.

Concentration (mM)	Δf_s (Hz)		
	MUA	MPA	DTDPA
5	27	15	8
10	22	19	14
15	25	15	10

Table A-3 Water contact angle of SAM of alkanethiol as a function of thiol concentration.

Concentration (mM)	Water contact angle (degree)		
	MUA	MPA	DTDPA
5	57.0±2.9	59.8±1.1	62.6±2.7
10	54.6±2.4	36.6±1.8	48.0±2.0
15	53.4±1.5	41.6±2.2	54.2±0.8

Activation of Carboxyl Groups of SAM-modified Substrates

Table A-4 Water contact angle and frequency shift due to the activation (Δf_a) of the MPA-modified substrate as a function of immersion time using 15:45 mM of NHS/EDCI.

Time (min)	Water contact angle (degree)	Δf_a (Hz)
0	36.0±1.8	-
15	50.6±1.7	22.2
30	54.6±1.9	25.7
45	54.0±3.1	18.7
60	52.0±3.1	16.0
90	51.8±1.5	20.0
120	45.2±2.0	20.0
240	44.0±0.7	19.0

Table A-5 Water contact angle due to the activation (Δf_a) of the MPA-modified substrate as a function of NHS/EDCI concentration using 30 min immersion time.

Concentration NHS(mM)/EDCI(mM)	Water contact angle (degree)
control	36.6±1.8
10/30	52.2±1.1
15/45	56.2±2.6
30/90	54.0±2.2

Table A-6 Frequency shift due to the activation (Δf_a) of the MPA-modified substrate as a function of NHS/EDCI concentration using 30 min immersion time.

Concentration NHS(mM)/EDCI(mM)	Δf_a (Hz)
control	-2
10/30	21
15/45	27
30/90	25

Immobilization of Monoclonal Antibody (MAb) against *Vibrio harveyi*

Table A-7 Frequency shift due to the MAb immobilization (Δf_i) on the NHS-modified substrate as a function of immobilization time using 0.1 mg/mL MAb.

Immobilization Time (h)	Δf_i (Hz)
8	28.3±3.2
15	38.3±9.2
24	40.0±3.6

Table A-8 Frequency shift due to the MAb immobilization (Δf_i) as a function of MAb concentration using the immobilization time of 15h.

Concentration (mg/mL)	Δf_i (Hz)
control	-2.0
0.05	20.0
0.1	22.0
0.5	27.0
1	27.5

Table A-9 Frequency shift due to *V. harveyi* binding (Δf_b) with the MAb-immobilized substrate after the treatment with blocking reagents. The concentration of *V. harveyi* used was 10^5 CFU/mL.

Blocking reagent	Δf_b (Hz)
None	4.0
1% BSA	21.0
6%ethanolamine	10.5

Table A-10 Frequency shift due to *V. harveyi* binding (Δf_b) with the MAb-immobilized substrate as a function of MAb concentration used in the immobilization step after the treatment with 1%BSA.

Time (min)	Δf_b (Hz)
0.05	11.0
0.1	21.0
0.5	9.0
1	6.0

Table A-11 Frequency shifts due to MAb immobilization (Δf_i) and *V. harveyi* binding (Δf_b) of the MPA-CE mixed SAM as a function of dilution ratio (%MPA).

% MPA	Δf_i (Hz)	Δf_b (Hz)
100%	34	21
80%	41	23
60%	39	34
40%	44	33
20%	47.8	39

Table A-12 Frequency shifts due to MAb immobilization (Δf_i) and *V. harveyi* binding (Δf_b) of the MPA-ME mixed SAM as a function of dilution ratio (%MPA).

% MPA	Δf_i (Hz)	Δf_b (Hz)
100%	34	21
80%	51	40
60%	24	17
40%	18	14
20%	16	19

Table A-13 Frequency shift due to *V. harveyi* binding (Δf_b) of the MAb-modified substrate prepared from MPA monolayer as a function of *V. harveyi* concentration.

Log concentration (CFU/mL)	Δf_b (Hz)
control	2.0
2	-12±7.0
3	7±6.7
4	17±4.0
5	21±7.2
6	27±9.5
7	30±4.4
8	29±14

Table A-14 Frequency shift due to bacteria binding (Δf_b) of the MAb-immobilized substrate prepared from MPA monolayer.

Bacteria	Δf_b (Hz)
control	2.0
<i>V. harveyi</i>	21.0 ± 7.2
<i>V. vulnificus</i>	5.6 ± 3.8
<i>V. paraheamolyticus</i>	-4.0 ± 3.2

Table A-15 Frequency shift due to *V. harveyi* binding (Δf_b) of the MAb-immobilized substrate prepared from MPA monolayer after 1 cycle of regeneration in 0.1 M glycine/HCl buffer solution (pH = 2.3) as a function of regeneration time.

Time (min)	Δf_b (Hz)
15	4.0
30	9.0
45	8.0
60	16.0
120	-21.0

VITAE

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