

REFERENCES

1. Y. Cao, T. I. Croll, J. G. Lees, B. E. Tuch, and J. J. Cooper-White. Scaffolds, Stem Cells, and Tissue Engineering: A Potent Combination. Aust. J. Chem 58 (2005) : 691-703.
- 2 B.L. Seal, T.C. Otero, A. Panitch. Review:Polymeric biomaterials for tissue and organ regeneration. Materials Science and Engineering 34 (2001) : 147-230.
3. G. L. Bowlin. A new spin on scaffolds. Materials today May (2004) : 64.
4. W. J. Li, C. T. Laurencin, E. J. Caterson, R. S. Tuan, F. K. Ko. Electrospun nanofibrous structure: A novel scaffold for tissue engineering. J Biomed Mater Res 60 (2002) : 613-621.
5. K. S. Rho, L. Jeong, G. Lee, B. M. Seo, Y. J. Park, S. D. Hong, S. Roh, J. J.Cho, W. H. Park, B. M. Min. Electrospinning of collagen nanofibers: Effects on the behavior of normal human keratinocytes and early-stage wound healing. Biomaterials 27 (2006) : 1452–1461.
6. P. X. Ma. Scaffolds for tissue fabrication. Materials today May (2004) : 30-40.
7. R.B. Martin. Bone as a Ceramic Composite Material. Materials Science 293 (1999) : 5-16.
8. P. Wuttichroenmongkol, N. Sanchavanakit, P. Pavasant, and P. Supaphol. Novel Bone Scaffolds of Electrospun Polycaprolactone Fibers Filled with Nanoparticles. J. Nanoscience and Nanotechnology 6 (2006) : 1-9.
9. H. Terai, D. Hannouche, E. Ochoa, Y. Yamano, J. P. Vacanti. In vitro engineering of bone using a rotational oxygen-permeable. Materials Science and Engineering 20 (2002) : 3 –8.

10. Z. M. Huanga, Y. Z. Zhang, M. Kotaki, S. Ramakrishna. A review on polymer nanofibers by electrospinning and their applications in nanocomposites. Composites Science and Technology 63 (2003) : 2223–2253.
11. J.M. Deitzel, J. Kleinmeyer, D. Harris, N.C. Beck Tan. The effect of processing variables on the morphology of electrospun nanofibers and textiles. Polymer 42 (2001) : 261–272.
12. Y.M. Shin, M.M. Hohman, M.P. Brenner, G.C. Rutledge. Experimental characterization of electrospinning : the electrically forced jet and instabilities. Polymer 42 (2001) : 9955-9967.
13. K. Ohgo, C. Zhao, M. Kobayashi, T. Asakura. Preparation of non-woven nanofibers of Bombyx mori silk, Samia cynthia ricini silk and recombinant hybrid silk with electrospinning method. Polymer 44 (2003) : 841–846.
14. K. Ohkawa, D. Cha, H. Kim, A. Nishida, H. Yamamoto. Electrospinning of Chitosan, Macromolecule rapid communication. 25 (2004) : 1600-1605.
15. H. Yoshimoto, Y.M. Shin, H. Terai, J.P. Vacanti. A biodegradable nanofiber scaffold by electrospinning and its potential for bone tissue engineering. Biomaterials 24 (2003) 2077–2082.
16. A. Sonthisombat and P. T. Speakman. Silk:Queen of Fibres-The Concise Story. (2004) : 1-28.
17. Y.Q Zhang. Applications of natural silk protein sericin in biomaterials. Biotechnology Advances 20 (2002) : 91-100.
18. M. Tsukada, G. Freddi, and N. Kasai. J. Polymer Science 32 (1994) : 1175-1182.

19. G. H. Altman, F. Diaz, C. Jakuba, T. Calabro, R. L. Horan, J. Chen, H. Lu, J. Richmond, D. L. Kaplan. Silk-based biomaterials. *Biomaterials* 24 (2003) : 401–416.
20. H. J. Jin, S. V. Fridrikh, G. C. Rutledge, and D. L. Kaplan. Electrospinning Bomvyx mori with Poly (ethylene oxide). *Biomacromolecules* 3 (2002) : 1233-1239.
21. H. J. Jin, J. Chen, V. Karageorgiou, G. H. Altman, D. L. Kaplan. Human bone marrow stromal cell responses on electrospun silk fibroin. *Biomaterials* 25 (2004) : 1039-1047.
22. B. M. Min, G. Lee, S. H. Kim, Y. S. Nam, T. S. Lee, W. H. Park. Electrospinning of silk fibroin nanofibers and its effect on the adhesion and spreading of normal human keratinocytes and fibroblasts in vitro. *Biomaterials* 25 (2004) : 1289–1297.
23. L. S. Rummler, et al. The Anatomy and Biochemistry of Myelin and Myelination. *Orthopaedics* 14 (2004) : 146-152.
24. C. Miller, H. Shanks, A. Witt, G. Rutkowski, S. Mallapragada. Oriented Schwann cell growth on micropatterned biodegradable polymer substrates. *Biomaterials* 22 (2001) : 1263-1269.
25. Y. Yuan, P. Zhang, Y. Yang, X. Wang, X. Gu. The interaction of Schwann cells with chitosan membranes and fibers in vitro. *Biomaterials* 25 (2004) 4273–4278.
26. C. A. Sundback, J. Y. Shyu, Y. Wang, W. C. Faquin, R. S. Langer, J. P. Vacanti, T. A. Hadlock. Biocompatibility analysis of poly(glycerol sebacate) as a nerve guide material. *Biomaterials* (2005) : 1-11.
27. H. Fong, I. Chun, D.H. Reneker. Bead nanofibers formed during electrospinning. *Polymer* 40 (1999) 4585-4592.

28. I. C. Um , H. Y. Kweon , Y. H. Park , S. Hudson. Structural characteristics and properties of the regenerated silk fibroin prepared from formic acid.
International Journal of Biological Macromolecules 29 (2001) : 91-97.
29. J. Ayutsede, M. Gandhi, S. Sukigara, M. Micklus, H. E. Chen, F. Ko.
Regeneration of Bombyx mori silk by electrospinning Part 3:
characterization of electrospun nonwoven mat.
Polymer 46 (2005) : 1625–1634.

APPENDICES

APPENDIX A

Table A1 The absorbance data of indirect cytotoxicity test from MTT assays

Replicates	Absorbance		
	Controls	Films	Fibrous mats
1	0.817	0.868	0.790
	0.813	0.775	0.844
	0.825	0.850	0.880
	0.825	0.792	0.811
Average	0.820	0.821	0.831
Standard deviation	0.006	0.045	0.039
2	0.772	0.656	0.766
	0.782	0.625	0.731
	0.789	0.748	0.756
	0.766	0.746	0.745
Average	0.777	0.694	0.750
Standard deviation	0.010	0.063	0.015
3	0.853	0.897	0.935
	0.859	0.910	0.906
	0.843	0.903	0.952
	0.848	0.887	0.929
Average	0.851	0.899	0.931
Standard deviation	0.007	0.010	0.019
Average	0.816	0.805	0.837
Standard deviation	0.008	0.039	0.024

Table A2 The absorbance data of direct cytotoxicity test from MTT assays

Replicates	Absorbance		
	Controls	Films	Fibrous mats
1	0.784	0.620	0.714
	0.795	0.643	0.721
	0.800	0.648	0.719
	0.786	0.651	0.717
Average	0.791	0.641	0.718
Standard deviation	0.008	0.014	0.003
2	0.735	0.557	0.609
	0.747	0.571	0.579
	0.748	0.579	0.581
	0.725	0.577	0.603
Average	0.739	0.571	0.593
Standard deviation	0.011	0.010	0.015
3	0.683	0.478	0.566
	0.717	0.457	0.543
	0.688	0.447	0.561
	0.720	0.466	0.532
Average	0.702	0.462	0.551
Standard deviation	0.019	0.013	0.016
Average	0.744	0.558	0.620
Standard deviation	0.013	0.012	0.011

Table A3 The absorbance data of Schwann cell attachment on the controls at the different times in culture from MTT assays

Replicates	Absorbance for the controls				
	Times in culture (h)				
	1	2	4	8	16
1	0.258	0.327	0.324	0.395	0.572
	0.270	0.259	0.385	0.413	0.619
	0.273	0.281	0.367	0.366	0.549
	0.291	0.326	0.321	0.329	0.493
Average	0.273	0.287	0.349	0.376	0.558
Standard deviation	0.014	0.029	0.032	0.037	0.052
2	0.272	0.281	0.401	0.408	0.563
	0.270	0.247	0.316	0.375	0.551
	0.253	0.276	0.341	0.377	0.565
	0.273	0.321	0.348	0.380	0.569
Average	0.267	0.281	0.352	0.385	0.562
Standard deviation	0.010	0.030	0.036	0.016	0.008
3	0.264	0.264	0.347	0.381	0.612
	0.252	0.261	0.391	0.365	0.547
	0.256	0.299	0.365	0.417	0.626
	0.288	0.308	0.334	0.368	0.592
Average	0.265	0.283	0.359	0.383	0.594
Standard deviation	0.016	0.024	0.025	0.024	0.034
Average	0.268	0.284	0.353	0.381	0.572
Standard deviation	0.013	0.028	0.031	0.025	0.032

Table A4 The absorbance data of Schwann cell attachment on the film scaffolds at the different times in culture from MTT assays

Replicates	Absorbance for the film scaffolds				
	Times in culture (h)				
	1	2	4	8	16
1	0.263	0.323	0.3	0.489	0.572
	0.290	0.293	0.284	0.387	0.619
	0.291	0.267	0.293	0.435	0.549
	0.265	0.281	0.32	0.480	0.493
Average	0.277	0.291	0.299	0.448	0.558
Standard deviation	0.015	0.024	0.015	0.047	0.052
2	0.261	0.278	0.291	0.408	0.563
	0.257	0.305	0.288	0.44	0.551
	0.301	0.309	0.267	0.482	0.565
	0.268	0.252	0.333	0.452	0.569
Average	0.272	0.286	0.295	0.446	0.562
Standard deviation	0.020	0.027	0.028	0.031	0.008
3	0.293	0.323	0.349	0.489	0.612
	0.243	0.315	0.269	0.399	0.547
	0.245	0.255	0.276	0.461	0.626
	0.323	0.267	0.305	0.46	0.592
Average	0.276	0.290	0.300	0.451	0.594
Standard deviation	0.039	0.034	0.036	0.038	0.034
Average	0.275	0.289	0.298	0.331	0.665
Standard deviation	0.025	0.028	0.026	0.014	0.035

Table A5 The absorbance data of Schwann cell attachment on the fibrous mat scaffolds at the different times in culture from MTT assays

Replicates	Absorbance for the fibrous mat scaffolds				
	Times in culture (h)				
	1	2	4	8	16
1	0.311	0.319	0.337	0.489	0.684
	0.251	0.307	0.343	0.387	0.612
	0.265	0.293	0.359	0.435	0.599
	0.277	0.258	0.32	0.480	0.72
Average	0.276	0.294	0.340	0.448	0.654
Standard deviation	0.026	0.026	0.016	0.047	0.058
2	0.248	0.333	0.356	0.408	0.591
	0.299	0.253	0.317	0.44	0.634
	0.243	0.323	0.345	0.482	0.677
	0.305	0.249	0.325	0.452	0.722
Average	0.274	0.290	0.336	0.446	0.656
Standard deviation	0.033	0.045	0.018	0.031	0.056
3	0.284	0.325	0.351	0.489	0.734
	0.315	0.303	0.341	0.399	0.653
	0.27	0.291	0.319	0.461	0.691
	0.233	0.251	0.345	0.460	0.659
Average	0.276	0.293	0.339	0.451	0.684
Standard deviation	0.034	0.031	0.014	0.038	0.037
Average	0.275	0.292	0.338	0.448	0.665
Standard deviation	0.031	0.034	0.016	0.038	0.050

Table A6 The absorbance data of Schwann cell proliferation on the controls at the different times in culture from MTT assays

Replicates	Absorbance for the controls		
	Times in culture [day(s)]		
	1	2	3
1	0.259	0.371	0.371
	0.268	0.373	0.393
	0.270	0.383	0.397
	0.274	0.394	0.390
Average	0.268	0.380	0.388
Standard deviation	0.006	0.011	0.012
2	0.275	0.4	0.407
	0.277	0.401	0.421
	0.284	0.410	0.412
	0.288	0.414	0.416
Average	0.281	0.406	0.414
Standard deviation	0.006	0.007	0.006
3	0.300	0.417	0.441
	0.301	0.418	0.442
	0.309	0.420	0.426
	0.320	0.451	0.440
Average	0.308	0.427	0.437
Standard deviation	0.009	0.016	0.008
Average	0.285	0.404	0.413
Standard deviation	0.007	0.011	0.008

Table A7 The absorbance data of Schwann cell proliferation on the film scaffolds at the different times in culture from MTT assays

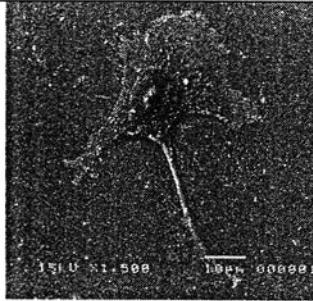
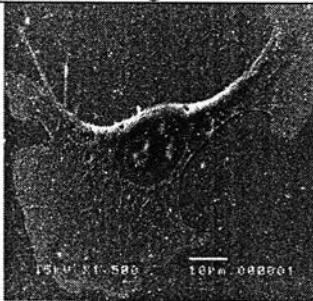
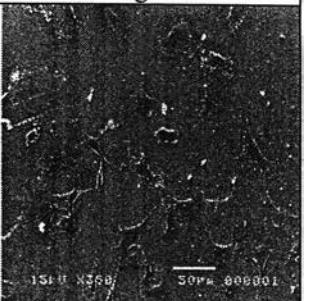
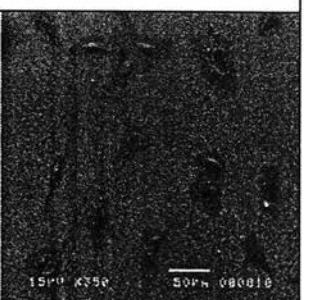
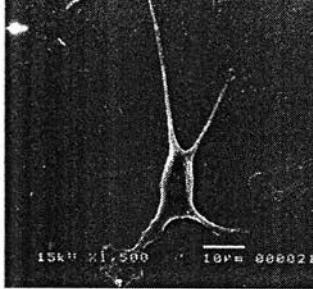
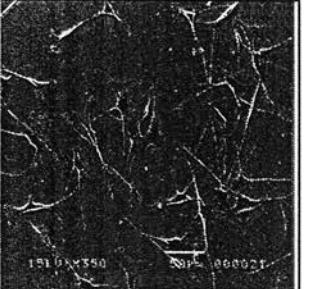
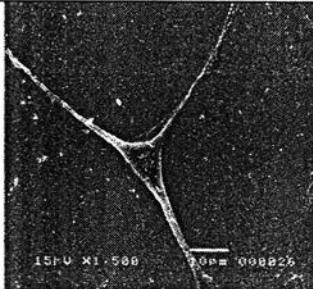
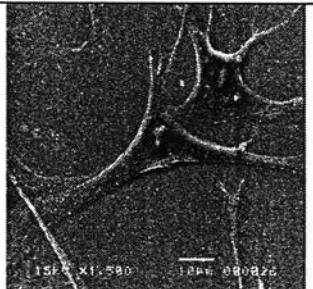
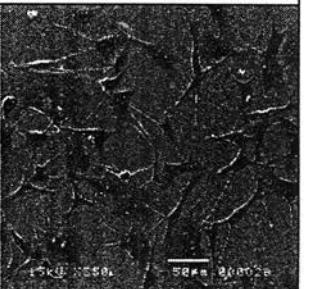
Replicates	Absorbance for the film scaffolds		
	Times in culture [day(s)]		
	1	2	3
1	0.267	0.246	0.266
	0.263	0.265	0.268
	0.255	0.270	0.276
	0.253	0.280	0.287
	Average	0.260	0.265
	Standard deviation	0.007	0.014
2	0.270	0.302	0.323
	0.276	0.310	0.317
	0.280	0.312	0.355
	0.296	0.315	0.345
	Average	0.281	0.310
	Standard deviation	0.011	0.006
3	0.305	0.331	0.388
	0.315	0.340	0.394
	0.295	0.357	0.416
	0.302	0.367	0.403
	Average	0.304	0.349
	Standard deviation	0.008	0.016
	Average	0.281	0.308
	Standard deviation	0.009	0.012
			0.337
			0.013

Table A8 The absorbance data of Schwann cell proliferation on the fibrous mat scaffolds at the different times in culture from MTT assays

Replicates	Absorbance for the fibrous mat scaffolds		
	Times in culture [day(s)]		
	1	2	3
1	0.242	0.295	0.367
	0.240	0.296	0.358
	0.249	0.298	0.348
	0.238	0.306	0.367
Average	0.242	0.299	0.360
Standard deviation	0.005	0.005	0.009
2	0.267	0.336	0.358
	0.260	0.307	0.357
	0.268	0.313	0.367
	0.274	0.326	0.385
Average	0.267	0.321	0.367
Standard deviation	0.006	0.013	0.013
3	0.303	0.348	0.417
	0.299	0.354	0.412
	0.309	0.374	0.424
	0.296	0.384	0.448
Average	0.302	0.365	0.425
Standard deviation	0.006	0.017	0.016
Average			
Standard deviation	0.270	0.328	0.384
	0.005	0.012	0.013

APPENDIX B

Table B1 The SEM images of Schwann cells cultured on the control at the different times in culture.

Times in culture	Controls		
	Fig. 1	Fig. 2	Fig. 3
1 h.			
Scale bar =	10 µm	10 µm	50 µm
2 h.			
Scale bar =	10 µm	10 µm	50 µm
4 h.			
Scale bar =	10 µm	10 µm	50 µm
8 h.			
Scale bar =	10 µm	10 µm	50 µm

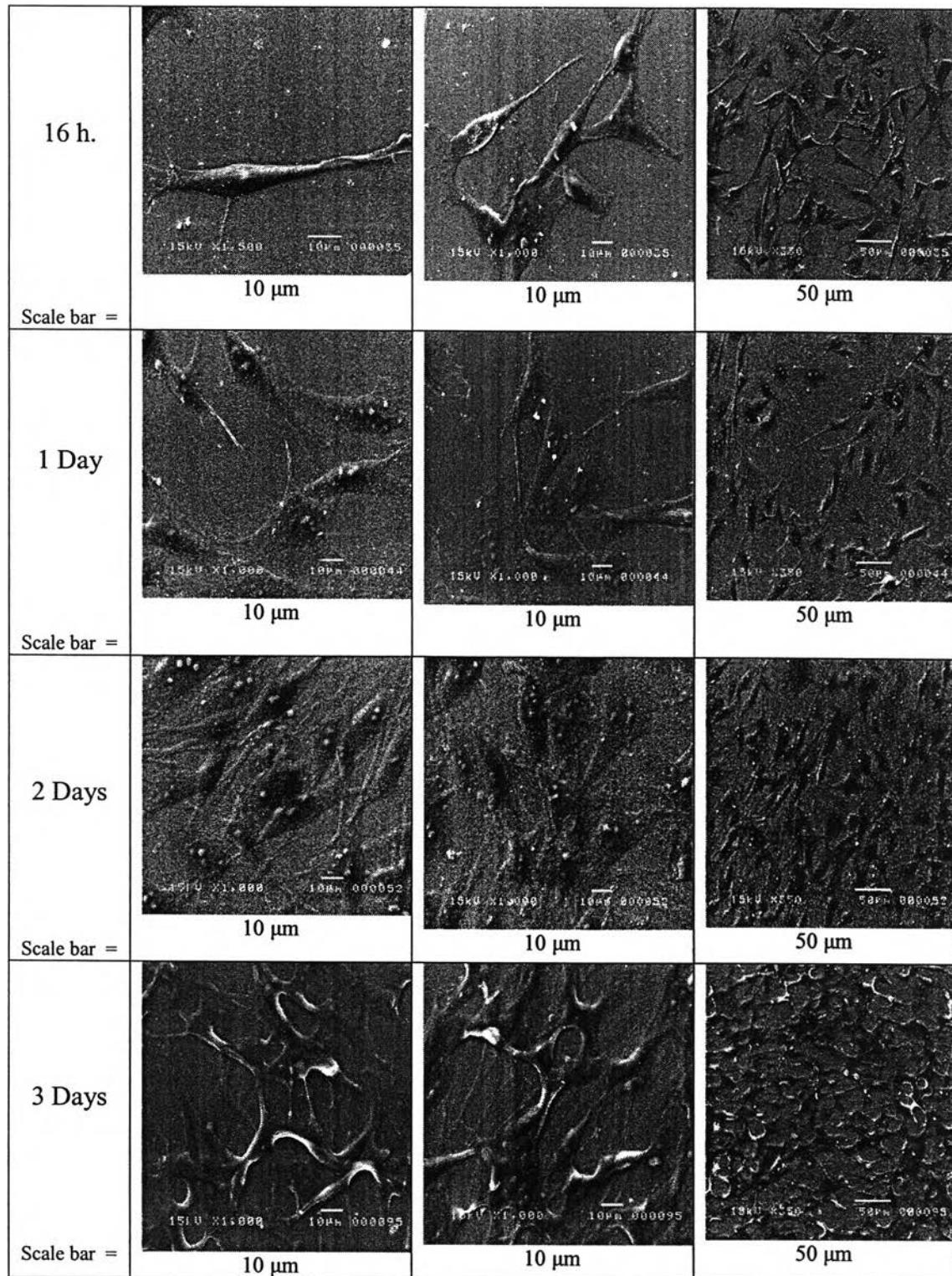
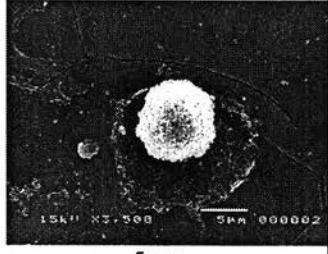
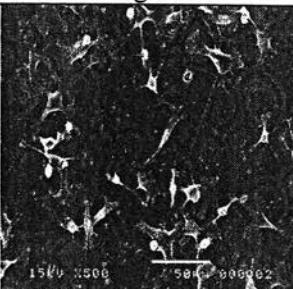
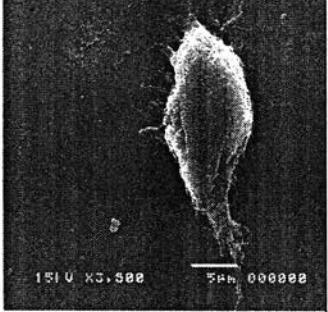
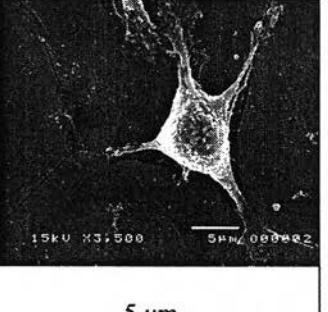
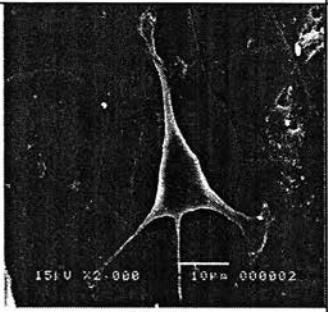
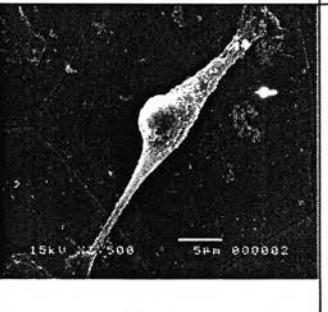
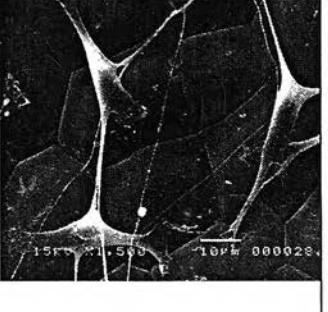
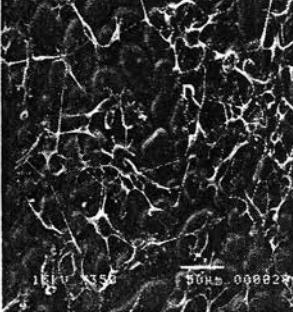


Table B2 The SEM images of Schwann cells cultured on the film scaffolds at the different times in culture.

Times in culture	Film scaffolds		
	Fig. 1	Fig. 2	Fig. 3
1 h. Scale bar =			
2 h. Scale bar =			
4 h. Scale bar =			
8 h. Scale bar =			

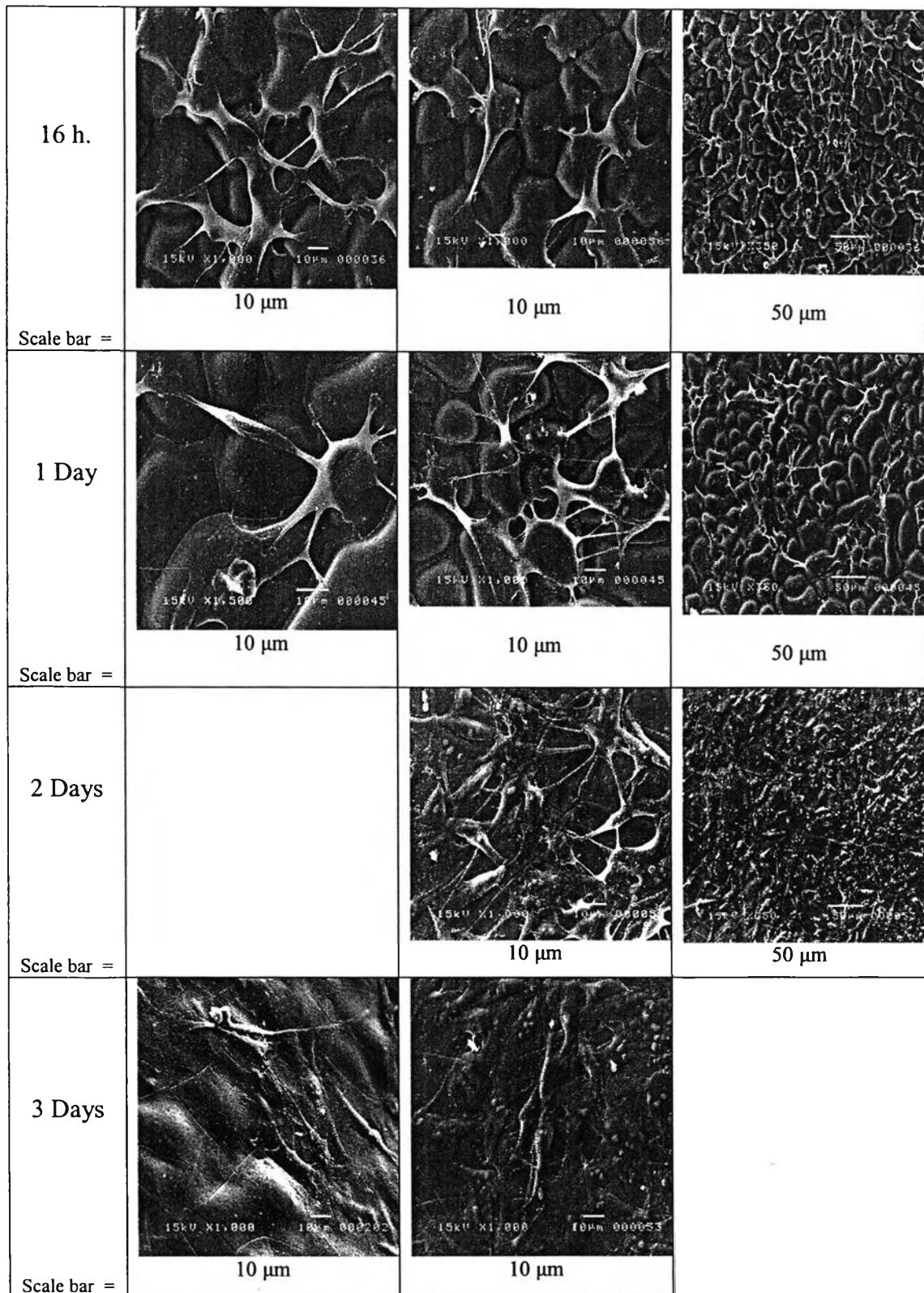
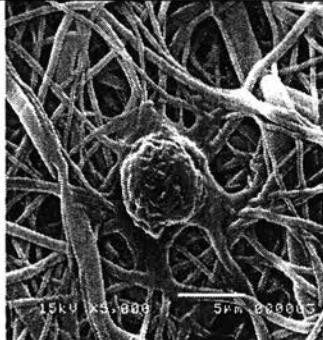
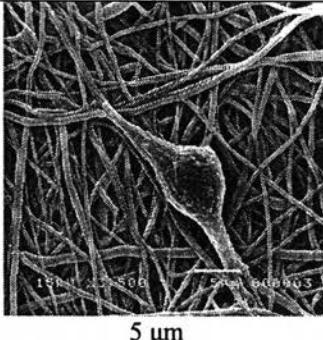
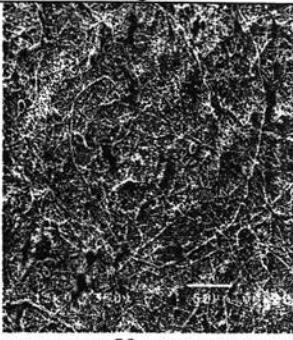
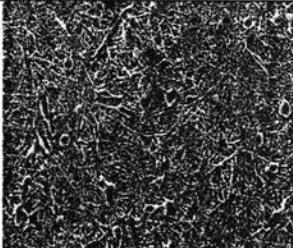
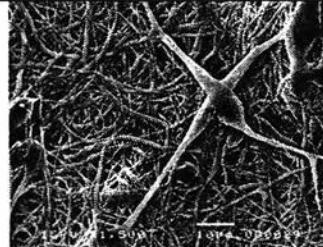
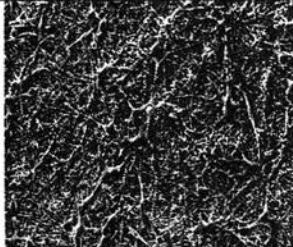
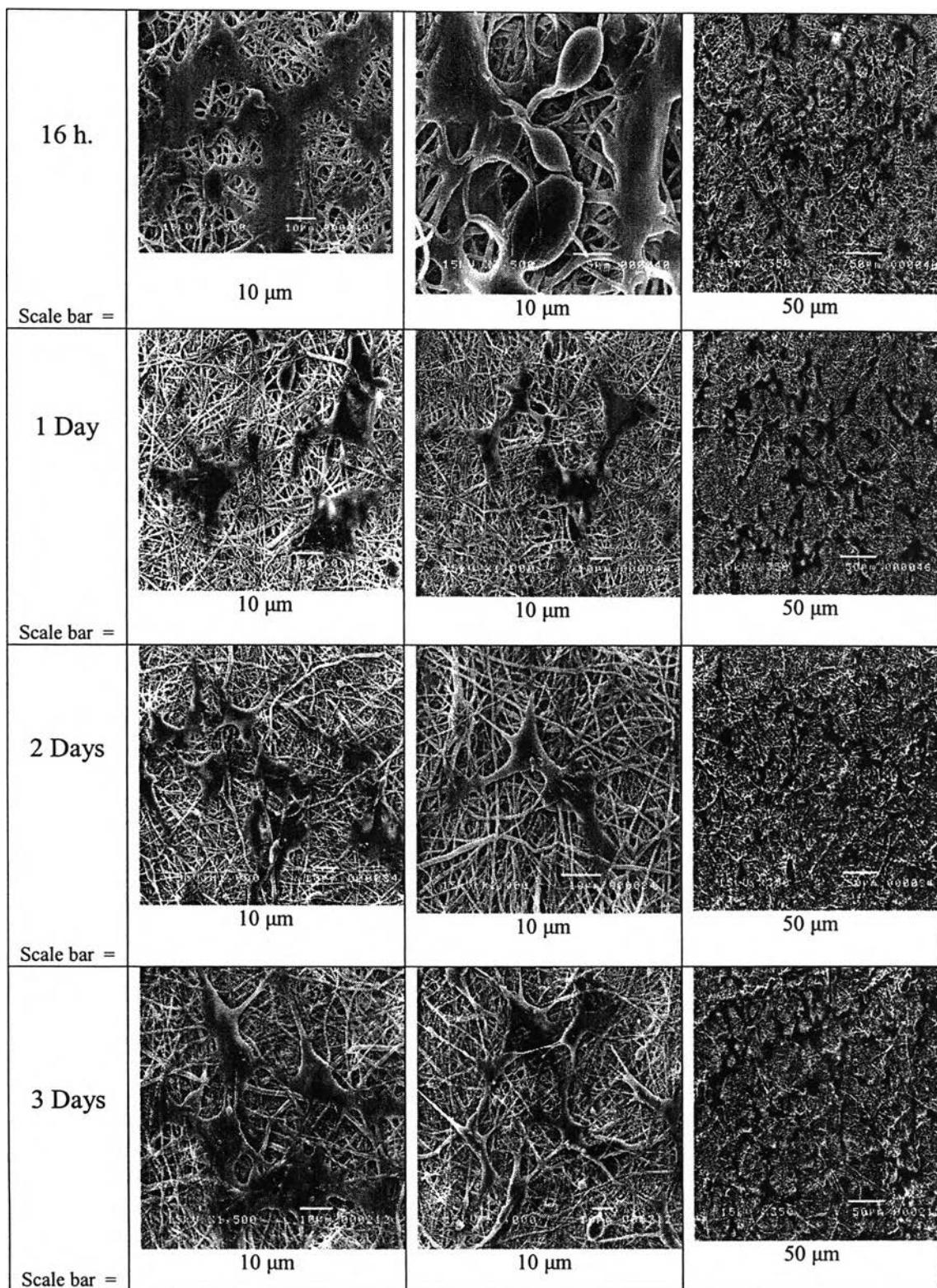


Table B3 The SEM images of Schwann cells cultured on the fibrous mat scaffolds at the different times in culture.

Times in culture	Fibrous mat scaffolds		
	Fig. 1	Fig. 2	Fig. 3
1 h. Scale bar =			
2 h. Scale bar =			
4 h. Scale bar =			
8 h. Scale bar =			



VITA

Tatiya Laksana-ngam was born on 19 May 1982 at Roi-ed province. She is a daughter of Mr. Supat Laksana-ngam and Ms. Sunit Laksana-ngam. She has one younger brother named Mr. Gowaras Laksana-ngam. In 2000, she graduated from Princess Chulabhorn's College Mukdaharn high school, then she kept on her study on Bachelor' s degree of Science, the major of Chemistry, at Mahidol University. In 2004, after she graduated from Mahidol University, she still wanted to learn something new and she was very interested in polymer science. Therefore, she decided to study on Master's degree of Science, in the field of Applied Polymer Science and Textile Technology, the major of Materials Science, at Chulalongkorn University and graduated in 2006.

