

REFERENCES

- Ameer, G.A., Mahmood, T.A., and Langer, R. (2002) A biodegradable composite scaffold for cell transplantation. Journal of Orthopedic Research, 20, 16-19.
- Ames, J.M., and Hofmann, T. (2002) Chemistry and Physiology of Selected Food Colorants. Washington: American Chemical Society.
- Baxter, A., Dillon, M., Taylor, K.D.A., Roberts, G.A.F. (1992) Improved method for i.r. determination of the degree of N-acetylation of chitosan. International Journal of Biological Macromolecular, 14, 166-169.
- Chen, G., Ushida, T., and Tateishi, T. (2001) Development of biodegradable porous scaffolds for tissue engineering. Materials Science and Engineering C 17, 63-69.
- Chen, L., Du, Y., Wu, H., and Xiao, L. (2002) Relationship between molecular structure and moisture-retention ability of carboxymethyl chitin and chitosan. Journal of Applied Polymer Science, 83, 1223-1241.
- Chen, X.G., Wang, Z., Liu, W.S., and Park, H.J. (2002) The effect of carboxymethyl-chitosan on proliferation and collagen secretion of normal and keloid skin fibroblasts. Biomaterials, 23, 4609-4614
- Edwards, J.V., and Vigo, T.L. (2001) Bioactive Fibers and Polymers. Washington: American Chemical Society.
- Guan, Y., Liu, X., Zhang, Y., and Yao, K. (1998) Study of phase behavior on chitosan/viscose rayon blend film. Journal of Applied Polymer Science, 67, 1965-1972.
- Janvikul, W., and Thavornyutikarn, B. (2003) New route to the preparation of carboxymethylchitosans hydrogels. Journal of Applied Polymer Science, 90, 4016-4020.
- Kittur, F.S., Prashanth, K.V.H., Udaya Sankar, K., and Tharanthan, R.N. (2002) Characterization of chitin, chitosan and their carboxymethyl derivatives by differential scanning calorimetry. Carbohydrate Polymers, 49, 185-193.

- Krause, T.J., Zazanis, G., Malatesta, P., and Solina, A. (2001) Prevention of pericardial Adhesions with N-O Carboxymethyl chitosan in the rabbit model. Journal of Investigative Surgery, 14, 93-97.
- Lee, J., Kim, C., Kato, S., and Murakami, R. (2001) Microwave-hydrothermal versus conventional hydrothermal preparation of Ni- and Zn-ferrite powder. Journal of Alloys and Compounds, 325, 276-280.
- Lee, J.Y., Nam, S.H., Im, S.Y., Park, Y.J., Lee, Y.M., Seol, Y.J., Chung, C.P., and Lee, S.J. (2002) Enhanced bone formation by controlled growth factor delivery from chitosan-based biomaterials. Journal of Controlled Release, 78, 187-197.
- Li, Z., Liu, X., Zhuang, X., Guan, Y., and Yao, K. (2002) Manufacture and properties of chitosan/N,O-carboxymethylated chitosan/viscose rayon antibacterial fibers. Journal of Applied Polymer Science, 84, 2049-2059.
- Li, Z., Zhuang X.P., Liu, X.F., Guan L.Y., and Yao, D.K. (2002) Study on antibacterial O-carboxymethylated chitosan/cellulose blend film from LiCl/N,N-dimethylacetamide solution. Polymer, 43, 1541-1547.
- Lim, L.Y., Khor, E., and Ling, C.E. (1999) Effects of dry heat and saturated steam on the physical properties of chitosan. Journal of Biomaterial Research, 48, 111-116.
- Liu, L., Li, Y., Li, Y., Fang, Y. (2004) Rapid N-phthaloylation of chitosan by microwave irradiation. Carbohydrate Polymers, 57, 97-100.
- Liu, X.F., Guan, Y.L., Yang, D.Z., Li, Z., and Yao, K.D. (2001) Antibacterial action of chitosan and carboxymethylated chitosan. Journal of Applied Polymer Science, 79, 1324-1335.
- Lloyd L.L., Kennedy, J.F., Methacanon, P., Paterson, M., and Knill, C.J. (1998) Carbohydrate polymers as wound management aids. Carbohydrate Polymers, 37, 315-322.
- Madhally, S.V. and Matthew, H.W. (1999) Porous chitosan scaffolds for tissue engineering. Biomaterials, 20(1), 1133-1142.
- Miya, M., Iwanmoto, R., Yoshigawa, S., and Mima, S. (1980) I.r. spectroscopic determination of CONH content in highly deacetylated chitosan. International Journal of Biological Macromolecular, 2, 323-324.

- Mori, T., Okumura, M., Matsuura, M., Ueno, K., Tokura, S., Okumoto, Y., Minami, S., and Fujinaga, T. (1997) Effects of chitin and its derivatives on the proliferation and cytokine production of fibroblasts in vitro. Biomaterials, 18, 947-951.
- Muramatsu, K., Masuda, S., Yoshihara, Y., and Fujisawa, A. (2003) In vitro degradation behavior of freeze-dried carboxymethyl-chitin sponges processed by vacuum-heating and gamma irradiation. Polymer Degradation and Stability, 81, 327-332.
- Nishi, N., Nogushi, J., Tokura, S., and Shiota, H. (1979) Studies on chitin. I. Acetylation of chitin. Polymer Journal, 11, 27-32.
- Nishimura, S., Ikeuchi, S.Y., and Tokura, S. (1984) The adsorption of bovine blood proteins onto the surface of O-carboxymethyl-chitin. Carbohydrate Research, 134, 305-312.
- Nishimura, S., Nishi, N., and Tokura, S. (1986) Bioactive chitin derivative activation of mouse-peritoneal macrophages by O-carboxymethyl-chitin. Carbohydrate Research, 146, 251-258.
- Park, S.N., Park, J.C., Kim, H.O., Song, M.J., and Suh, H. (2002) Characterization of porous collagen/hyaluronic acid scaffold modified by 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide cross-linking. Biomaterials, 23, 1205-1212.
- Ragnhild, J., Hjerde, N., Varum, K.M., Grasdén, H., Tokura, S., and Smidsrud, O. (1997) Chemical composition of O-(carboxymethyl)-chitins in relation to lysozyme degradation rates. Carbohydrate Polymers, 34, 131-139.
- Rocha, L.B., Goissis, G., and Rossi, M.A. (2002) Biocompatibility of anionic collagen matrix as scaffold for bone healing. Biomaterials, 23, 449-456.
- Satage, C., Verneuil, B., Branland, P., Granet, R., Krausz, P., Rozier, J., and Petit, C. (2002) Rapid homogeneous esterification of cellulose induced by microwave irradiation. Carbohydrate Polymer, 49, 373-376.
- Shanmugasundaram, N., Ravichandran, P., Neelakanta, R.P., Ramammurthy, N., Pal, S., and Panduranga, R.K. (2001) Collagen-chitosan polymeric scaffolds for the in vitro culture of human epidermoid carcinoma cells. Biomaterials, 1943-1951.

- Stone, C.A., Wright, H., Clarke, T., Powell, R., and Devaraj, V.S. (2000) Healing at skin graft donor sites dressed with chitosan. British Journal of Plastic Surgery, 53, 601-606.
- Sun, T., Xu, P., Liu, Q., Xue, J., and Xie, W. (2003) Graft copolymerization of methacrylic acid onto carboxymethyl chitosan. European Polymer Journal, 39, 189-192.
- Thanou, M., Verhoef, J.C., and Junginger .H.E. (2001) Oral drug absorption enhancement by chitosan and its derivatives. Advanced Drug Delivery Reviews, 52, 117-126.
- Tokura, S., Nishi, N., Takahashi, K, Shirai, A., and Uraki, Y. (1995) Novel drug-delivery system by chitin derivative. Macromolecule Symposia, 99, 201-208.
- Tokura, S., and Tamura, H. (2001) O-carboxymethyl-Chitin concentration in Granulocytes during Bone Repair. Biomacromolecules, 2, 417-421.
- Tokura, S., Nishimura, S., Sakairi, N., and Nishi, N. (1996) Biological activities of biodegradable polysaccharide. Macromolecular symposium, 101, 389-396.
- Tokura, S., Nishimura, S., and Nishi, N. (1983) Studies on chitin IX. Specific binding of calcium ions by carboxymethyl-chitin. Polymer Journal, 15, 597-602.
- Ueno, H., Nakamura, F., Murakami, M., Okumura, M., Kadosawa, T., and Fujinaga, T. (2001) Evaluation effects of chitosan for the extracellular matrix production by fibroblasts and the growth factors production by macrophages. Biomaterials, 22, 2125-2130.
- Vandelli, M.A., Romagnoli, M., Monti, A., Gozzi, M., Guerra, P., Rivasi, F., and Forni, F. (2004) Microwave-treated gelatin microspheres as drug delivery system. Journal of Controlled Release, 96, 67-84.
- Visser, C.E., Voute, A.B.E., Oosting, J., Boon, M.E., and Kok, L.P. (1992) Microwave irradiation and cross-linking of collagen. Biomaterials, 13, 34-37.
- Wang, W., Bo, S., Li, S., and Qin, W. (1991) Determination of the Mark-Houwink equation for chitosans with different degrees of deacetylation. International Journal of Biological Macromolecules, 13, 281-285.

- Xie, W., Xu, P., Wang, W., and Liu, Q. (2002) Preparation of water-soluble chitosan derivatives and their antibacterial activity. Journal of Applied Polymer Science, 85, 1357-1361.
- Zhang, L., Guo, J., Zhou, J., Yang, G., and Du, Y. (2000) Blends membranes from carboxymethylated chitosan/alginate in aqueous solution. Journal of Applied Polymer Science, 77, 610-616.
- Zhao, L., Mitomo, H., Nagasawa, N., Yoshi, F., and Kume, T. (2003) Radiation synthesis and characteristic of the hydrogels based on carboxymethylated chitin derivatives. Carbohydrate Polymers, 51, 169-175.

APPENDICES

APPENDIX A Effects of Microwave Exposure Time and Temperature on the %Weight Loss and Degree of Swelling of CM-chitin and CM-chitosan

Table A1 Effect of microwave exposure time on the %Weight loss of (A) CM-chitin and (B) CM-chitosan films at 100°C

(A)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	100	100	100	100	0
10	100	100	100	100	0
20	100	100	100	100	0
30	61.82	61.81	60.65	61.43	0.672
50	27.25	28.93	31.32	29.17	2.04
70	26.92	30.68	28.37	28.65	1.89

(B)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	100	100	100	100	0
10	100	100	100	100	0
20	52.07	46.37	52.79	50.41	3.51
30	32.10	35.05	34.34	33.83	1.53
50	30.34	29.53	33.33	31.06	2.00
70	25.06	27.62	26.26	26.31	1.28

Table A2 Effect of microwave exposure time on the %Weight loss of (A) CM-chitin and (B) CM-chitosan films at 110°C

(A)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	78.58	82.11	84.06	81.58	2.77
10	37.24	43.84	41.19	40.75	3.32
20	31.39	31.97	29.23	30.86	1.44
30	23.52	21.97	23.53	23.00	0.89
50	21.04	15.72	18.85	18.53	2.67
70	18.59	20.27	20.51	19.79	1.04

(B)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	55.25	53.05	53.94	54.08	1.10
10	37.12	34.06	32.86	34.68	2.19
20	29.62	31.20	31.10	30.64	0.88
30	24.72	25.48	24.34	24.84	0.58
50	21.73	21.69	20.86	21.42	0.49
70	21.02	18.41	21.24	20.22	1.57

Table A3 Effect of microwave exposure time on the %Weight loss of (A) CM-chitin and (B) CM-chitosan films at 120°C.

(A)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	69.66	67.43	72.57	69.88	2.57
10	37.44	37.75	36.59	37.26	0.60
20	21.84	20.42	19.71	20.65	1.08
30	16.19	17.67	18.66	17.50	1.24
50	17.24	15.41	18.02	16.89	1.33
70	16.72	18.76	16.01	17.16	1.42

(B)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	43.52	42.19	51.81	45.84	5.21
10	28.82	29.76	24.93	27.83	2.56
20	18.30	18.67	18.09	18.35	0.29
30	15.22	15.39	14.73	15.11	0.34
50	13.14	18.75	13.26	15.05	3.20
70	17.42	16.46	18.30	17.39	0.92

Table A4 Effect of microwave exposure time on the Degree of swelling of (A) CM-chitin and (B) CM-chitosan films at 100°C

(A)

Time (min)	DS			Average	Standard deviation
5	-	-	-	-	-
10	-	-	-	-	-
20	-	-	-	-	-
30	58.67	63.07	61.50	61.08	2.22
50	10.75	11.50	12.68	11.64	0.97
70	6.32	8.45	7.89	7.55	1.10

(B)

Time (min)	DS			Average	Standard deviation
5	-	-	-	-	-
10	-	-	-	-	-
20	29.86	24.53	28.52	27.63	2.77
30	7.06	7.99	7.71	7.58	0.47
50	5.30	5.35	5.93	5.52	0.35
70	3.62	3.97	3.71	3.76	0.18

Table A5 Effect of microwave exposure time on the Degree of swelling of (A) CM-chitin and (B) CM-chitosan films at 110°C

(A)

Time (min)	DS			Average	Standard deviation
5	109.66	130.77	112.10	117.51	11.54
10	29.67	32.16	31.75	31.19	1.33
20	14.49	15.53	11.55	13.85	2.06
30	4.49	5.19	5.40	5.02	0.47
50	3.81	3.93	3.50	3.74	0.22
70	2.88	3.30	3.54	3.24	0.33

(B)

Time (min)	DS			Average	Standard deviation
5	37.74	33.20	32.56	34.50	2.82
10	11.45	9.96	8.54	9.98	1.45
20	4.70	7.08	6.04	5.94	1.19
30	3.53	3.80	4.00	3.77	0.23
50	3.44	2.60	2.96	3.00	0.42
70	2.67	2.76	2.72	2.71	0.04

Table A6 Effect of microwave exposure time on the Degree of swelling of (A) CM-chitin and (B) CM-chitosan films at 120°C

(A)

Time (min)	DS			Average	Standard deviation
5	114.45	94.90	91.02	100.12	12.55
10	23.72	25.37	24.05	24.38	0.87
20	5.49	4.63	5.32	5.14	0.45
30	4.44	3.64	4.82	4.30	0.60
50	2.71	2.29	2.52	2.50	0.21
70	2.10	2.32	2.23	2.21	0.11

(B)

Time (min)	DS			Average	Standard deviation
5	16.20	14.78	13.70	14.89	1.25
10	5.54	5.82	5.13	5.49	0.34
20	3.30	2.90	3.24	3.14	0.21
30	2.10	2.17	2.05	2.10	0.06
50	1.83	2.14	2.48	2.15	0.32
70	2.01	2.13	2.33	2.15	0.16

Table A7 Effect of microwave exposure temperature on the %Weight loss of (A) CM-chitin and (B) CM-chitosan films at experimental time was 50 min

(A)

Temperature (°C)	%WL			Average	Standard deviation
80	100	100	100	100	0
90	89.09	91.32	84.35	88.25	3.55
100	27.25	28.93	31.32	29.16	2.04
110	21.04	15.72	18.85	18.53	2.67
120	17.24	15.41	18.02	16.89	1.33
130	20.04	17.62	19.58	19.08	1.28
140	19.56	20.07	15.00	18.21	2.79

(B)

Temperature (°C)	%WL			Average	Standard deviation
80	100	100	100	100	0
90	55.60	52.10	57.21	54.97	2.61
100	30.34	29.53	33.33	31.06	2.00
110	21.73	21.69	20.86	21.42	0.49
120	13.14	18.75	13.26	15.05	3.20
130	19.97	18.34	17.47	18.59	1.26
140	18.13	17.39	17.89	17.80	0.37

Table A8 Effect of microwave exposure temperature on the Degree of swelling of (A) CM-chitin and (B) CM-chitosan films at experimental time was 50 min

(A)

Temperature (°C)	DS			Average	Standard deviation
90	123.11	140.90	142.88	135.63	10.88
100	10.75	11.50	12.6861	11.64	0.97
110	3.81	3.93	3.5045	3.74	0.21
120	2.71	2.29	2.5261	2.50	0.21
130	2.09	2.0832	2.0744	2.08	0.01
140	1.70	1.87	1.6490	1.73	0.11

(B)

Temperature (°C)	DS			Average	Standard deviation
90	36.56	31.07	39.33	35.65	4.20
100	5.30	5.35	5.93	5.52	0.35
110	3.44	2.60	2.96	3.00	0.42
120	1.83	2.14	2.48	2.15	0.32
130	2.21	2.19	2.27	2.22	0.04
140	1.85	1.93	1.52	1.76	0.21

APPENDIX B Effects of Autoclave Exposure Time on the %Weight Loss and Degree of Swelling of CM-chitin and CM-chitosan at 120°C

Table B1 Effect of autoclave exposure time on the %Weight loss of (A) CM-chitin and (B) CM-chitosan films at 120°C

(A)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	100	100	100	100	0
10	100	100	100	100	0
20	100	100	100	100	0
30	100	100	100	100	0
50	36.23	31.07	33.42	33.57	2.58
70	22.36	25.15	20.70	22.73	2.24

(B)

Time (min)	%WL			Average	Standard deviation
0	100	100	100	100	0
5	100	100	100	100	0
10	100	100	100	100	0
20	100	100	100	100	0
30	41.96	59.91	50.85	50.90	8.97
50	23.90	30.62	21.73	25.41	4.63
70	21.57	19.96	20.26	20.59	0.85

Table B2 Effect of autoclave exposure time on the Degree of swelling of (A) CM-chitin and (B) CM-chitosan films at 120°C

(A)

Time (min)	DS			Average	Standard deviation
5	-	-	-	-	-
10	-	-	-	-	-
20	-	-	-	-	-
30	-	-	-	-	-
50	22.17	27.98	22.54	24.23	3.25
70	5.07	4.45	4.02	4.51	0.52

(B)

Time (min)	DS			Average	Standard deviation
5	-	-	-	-	-
10	-	-	-	-	-
20	-	-	-	-	-
30	43.97	49.84	39.00	44.27	5.42
50	3.66	5.90	2.96	4.17	1.53
70	2.47	2.85	2.95	2.75	0.25

APPENDIX C Viscosity-Average Molecular Weight of CM-chitin**Table C1** Viscosity-average molecular weight of CM-chitin

Conc. (g/100ml)	Time (sec.)			Average	η_{sp}/C	$\ln(\eta_{rel})/C$
	X_1	X_2	X_3			
0.0000	105.38	105.28	105.12	105.26		
0.00625	111.03	110.50	110.72	110.75	8.0798	7.8824
0.0125	121.75	121.93	122.19	121.95	8.4232	8.0086
0.0250	147.84	147.75	147.28	147.62	8.6853	7.8599
0.0500	210.03	209.69	209.78	209.83	9.4657	7.7499
0.1000	105.38	105.28	105.12	105.26	10.9415	7.3915

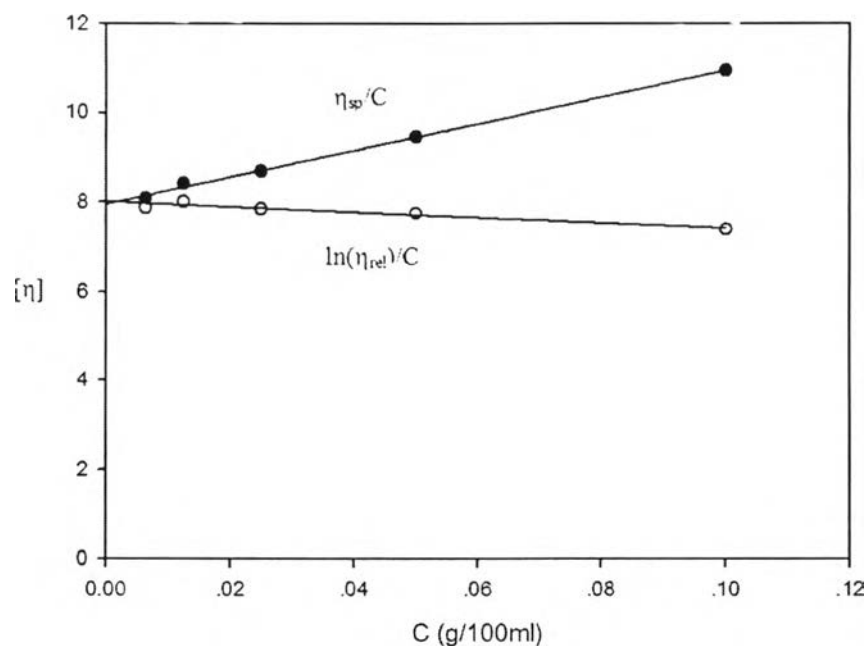


Figure C1 η_{sp}/C and $\ln(\eta_{rel})/C$ against concentration of CM-chitin solution.

The viscosity-average molecular weight of CM-chitin was determined based on Mark-Houwink equation. The K and a values were according to Kaneko *et al.* (1982).

$$[\eta] = 7.92 \times 10^{-5} M^1$$

Where $[\eta]$ = intrinsic viscosity

M = Viscosity-average molecular weight

Interception: $[\eta]$ of CM-chitin was 7.98

From calculation; $M = 100,757$ Da

The viscosity-average molecular weight of CM-chitin obtained from calculation was 100 kDa.

APPENDIX D Cytotoxicity of Microwave-Treated CM-chitin and CM-chitosan

Table D1 Direct cytotoxicity: a number of living cell after polymer films were deposited on confluent cell cultures for (A) 24 h and (B) 48 h

(A)

Types of polymers	Polystyrene	Chitosan	CM-chitosan	Chitin	CM-chitin
Number of cells (% of control)	97.81	88.76	44.85	83.30	97.03
	102.95	84.68	63.41	93.16	111.83
	99.31	86.90	67.81	91.15	91.86
	100	90.99	92.35	104.25	90.20
	99.45	97.77	86.69	100.63	98.64
	100.00	97.45	68.92	102.31	104.22
Average	100	91.09	70.67	95.79	98.96
Standard deviation	1.71	5.46	17.06	8.01	8.04

(B)

Types of polymers	Polystyrene	Chitosan	CM-chitosan	Chitin	CM-chitin
Number of cells (% of control)	98.39	65.33	18.80	67.54	71.89
	103.39	76.21	21.93	78.79	83.86
	98.20	81.68	39.01	83.01	95.03
	100.78	93.19	43.41	97.25	88.78
	97.83	100.90	48.94	101.32	87.80
	101.44	97.25	55.94	94.87	96.27
Average	100	85.76	38.00	87.13	87.27
Standard deviation	2.22	13.72	14.82	12.91	8.85

Table D2 Direct cytotoxicity: Total protein of cells after polymer films were deposited on confluent cell cultures for (A) 24 h and (B) 48 h

(A)

Types of polymers	Polystyrene	Chitosan	CM-chitosan	Chitin	CM-chitin
Total protein of cells (%)	95.86	105.43	102.23	80.14	121.88
	90.34	108.80	101.25	90.35	107.05
	113.79	110.49	105.15	95.07	101.29
Average	100	108.24	102.88	88.52	110.07
Standard deviation	12.25	2.57	2.028	7.63	10.62

(B)

Types of polymers	Polystyrene	Chitosan	CM-chitosan	Chitin	CM-chitin
Total protein of cells (%)	90.48	117.88	93.62	102.10	126.76
	116.24	118.72	137.97	114.55	136.58
	93.27	115.37	105.12	108.74	138.37
Average	100	117.32	112.24	108.46	133.90
Standard deviation	14.13	1.74	23.01	6.23	6.24

Table D3 Indirect cytotoxicity: a number of cells cultured in extraction medium at given concentration of (A) microwave-treated CM-chitin and (B) microwave-treated CM-chitosan

(A)

Concentration (mg/ml)	0.0156	0.03125	0.0625	0.125	0.25	0.5	1	2
Number of cells (% of control)	94.91	89.20	82.42	87.68	95.70	83.50	90.84	87.79
	97.68	85.31	97.34	86.83	90.62	88.24	88.41	85.81
	98.70	80.56	87.85	84.29	94.46	104.8	92.99	87.28
	107.34	82.59	78.81	79.83	98.81	94.63	92.09	76.38
	100	90.33	84.57	91.46	101.9	106.3	92.20	80.62
	90.67	83.05	81.75	90.33	95.19	99.94	86.15	92.76
	98.19	81.58	88.53	80.84	80.84	86.77	87.06	85.14
	83.78	88.41	83.72	74.74	86.32	93.89	92.25	85.31
Average	96.44	85.14	85.64	84.51	92.95	93.89	90.28	85.14
Standard deviation	6.68	3.58	5.48	5.48	6.57	8.53	2.55	5.54

(B)

Concentration (mg/ml)	0.0156	0.03125	0.0625	0.125	0.25	0.5	1	2
Number of cells (% of control)	92.57	90.09	93.63	102.0	90.62	89.50	80.83	75.23
	112.7	95.22	94.51	87.08	89.56	97.81	78.47	69.98
	98.11	99.29	98.05	103.2	106.5	101.0	87.32	69.92
	99.41	93.92	93.04	104.2	101.4	95.99	84.19	70.45
	93.04	113.7	93.57	107.9	96.81	94.81	84.90	75.23
	110.9	98.64	94.69	92.68	90.50	96.69	83.31	69.98
	84.02	92.51	85.96	88.32	89.62	90.97	84.31	69.92
	91.33	94.92	94.98	91.86	87.73	87.26	84.43	70.45
Average	97.81	97.28	93.57	97.16	94.10	94.28	83.49	71.40
Standard deviation	2.59	2.71	4.59	6.78	8.07	3.41	7.31	9.84

APPENDIX E Adhesion of Fibroblasts on Microwave-Treated CM-chitin and CM-chitosan Films

Table E1 Adhesion of fibroblasts on the chitosan, chitin, microwave-treated CM-chitin, and microwave-treated CM-chitosan for (A) 1 h and (B) 2 h

(A)

Types of polymers	Chitosan	CM-chitosan	Chitin	CM-chitin
Number of cells (%)	87.88	17.88	89.85	7.18
	98.02	18.45	152.25	11.83
	114.08	15.49	114.22	7.60
	103.78	40.15	94.69	10.00
	98.48	43.93	117.42	12.00
	97.72	46.21	122.72	15.00
Average	100	30.35	115.19	10.60
Standard deviation	8.61	14.49	22.35	2.96

(B)

Types of polymers	Chitosan	CM-chitosan	Chitin	CM-chitin
Number of cells (%)	148.30	56.19	172.39	26.05
	173.80	28.30	166.61	32.81
	170.98	46.19	167.04	24.64
	181.81	76.51	227.27	24.24
	164.39	78.78	281.81	44.69
	181.81	73.48	286.36	53.78
Average	170.18	59.91	216.91	34.37
Standard deviation	12.62	20.08	56.81	12.26

CURRICULUM VITAE

Name: Mr. Panya Wongpanit

Date of Birth: January 15, 1981

Nationality: Thai

University Education:

1999-2003 Bachelor Degree of Engineering in Petrochemical and Polymeric Materials, Faculty of Engineering and Industrial Technology, Silpakorn University, Nakhonpathom, Thailand.