ELECTROSPUN FIBER MATS AND HYDROGELS CONTAINING HERBAL SUBSTANCES FOR BIOMEDICAL APPLICATIONS

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ABSTRACT

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Keywords: Electrospinning / Poly L-(lactic acid)/ Polyacrylonitrile (PAN)/

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adenophorum / Wound dressing

New biomaterial efffective for use as topical/transdermal patches or wound dressings containing herbal substances (gallic acid, caffeic acid, and Eupatorium adenophorum essential oil) were developed. In order to improve the antioxidant properties, gallic acid, a natural phenolic antioxidant, was incorporated in PLLA electrospun fiber mats. The release characteristic of gallic acid from these materials were investigated by the total immersion method. Incorparation of gallic acid in the PLLA electrospun fibers improved the antioxidant properties. Caffeic acid (CA) was chemically immobilized onto the individual fiber surface of electrospun PLLA fiber mats to enhance the hydrophilicity and impart the antioxidant activity to the fibrous membrane. Indirect cytotoxicity evaluation, with murine dermal fibroblasts (L929) and human dermal fibroblasts (HDFa) revealed that the neat and the modified PLLA fibrous matrices in the level that were not harmful to the cells. Moreover, the wound dressing application was explored by the studies of gelatin hydrogels containing E. adenophorum essential oil emulsion which could be fabricated into casting-films and improved its water resistance properties by crosslinking with glutaralgehyde. It showed the antibacterial activities against Gram positive and Gram negative bacteria. In addition, the mangosteen extract-loaded polyacrylonitrile fiber mats were fabricated for filter application as a surgical mask. This study demonstrated a convenient procedure and the potential to develop antimicrobial and antituberculosis properties of electrospun fibrous membranes containing Garcinia mangostana (Mangosteen extract).

บทคัดย่อ

ปิยฉัฏร ช่วยสีนวล : วัสคุเส้นใยจากเทคนิคอิเลคโตรสปันและวัสคุไฮโครเจลที่ มีสารออกฤทธิ์ทางชีวภาพซึ่งสกัดจากสมุนไพรไทยและการประยุกต์ใช้ทางการแพทย์ (Electrospun fiber mats and hydrogels containing herbal substances for biomedical applications) อ. ที่ปรึกษา: ศาสตราจารย์ คร. พิชญ์ ศุภผล 159 หน้า

ในปัจจุบันสารสกัดจากสมุนไพรไทยกำลังได้รับความสนใจเป็นอย่างมากเนื่อง ค้วยคุณสมบัติที่คีของสารออกฤทธิ์ต่างๆที่มีอยู่ในสารสกัด เช่น สมบัติในการช่วยให้การ หายของแผลเร็วขึ้น การศึกษานี้จึงต้องการศึกษากระบวนการขึ้นรูปผลิตภัณฑ์จากพอลิเม อร์ชนิคต่างๆ ซึ่งมีคุณสมบัติของการออกฤทธิ์ของสารสกัดจากสมุน ใพรไทยและการ นำไปประยุกต์ใช้ สำหรับการประยุกต์ใช้เป็นวัสดุปิดแผล พอลิแลคติกแอซิคซึ่งมี ส่วนประกอบของแกลลิคแอซิคถูกขึ้นรูปด้วยกระบวนการอิเลคโตรสปันและศึกษา คุณสมบัติในการปลคปล่อยสารออกฤทธิ์พบว่าเส้นใยอิเลคโตรสปันที่ใต้มีคุณสมบัติใน การต่อด้านอนุมูลอิสระเมื่อทคสอบด้วย DPPH assay คุณสมบัติพื้นผิวของเส้นใยโพลิ เลกติดแอซิดยังได้ถูกปรับปรุงเพื่อให้มีคุณสมบัติในการคูดซับน้ำดีขึ้นและเพิ่มคุณสมบัติ ในการต่อต้านอนุมูลอิสระโดยการใช้เทคนิค Grafting ด้วยคาเฟอิกแอซิคพบว่าเส้น ใยอิเลคโตรสปันคังกล่าวมีคุณสมบัติที่ดีในการช่วยให้เซลล์ไฟโบรพลาสที่ได้จากผิวหนัง (human dermal fibroblast) ผิวผนังมีความสามารถในการเกาะบนพื้นผิวเส้นใยอิเลคโตรส ปันได้เป็นอย่างดี นอกจากนี้กระบวนการขึ้นรูปเจลาตินไฮโดรเจลยังได้ถูกศึกษา คุณสมบัติในการใช้เป็นวัสคุปิดแผล จากการศึกษาเจลาตินไฮโครเจลซึ่งปรับปรุง คุณสมบัติการป้องกันน้ำด้วยวิธีการ Crosslinking ด้วยกลูตารัลดิไฮด์และมีส่วนผสม ของสารสกัดจากสมุนไพรสาบหมาพบว่าวัสคุไฮโครเจลที่ได้มีคุณสมบัติในการออกฤทธิ์ ยับยั้งเชื้อแบคทีเรียชนิคแกรมบวกและแกรมลบชนิคต่างๆ ได้เป็นอย่างดีซึ่งวัสดุดังกล่าว ทั้งหมคสามารถนำไปใช้เป็นวัสคูเพื่อนำไปใช้ทางการแพทย์ที่มีประสิทธิภาพ สำหรับ คุณสมบัติในการออกฤทธิ์ยับยั้งเชื้อแบคทีเรียที่ทำให้เกิดโรค เช่น เชื้อวัณโรค โพลิอะครี โรในไตรค์ซึ่งถูกขึ้นรูปด้วยกระบวนการอิเลคโตรสปันและมีส่วนผสมของสารสกัดจาก เปลือกมังคุดนั้นพบว่ามีคุณสมบัติในการออกฤทธิ์ต่อด้านเชื้อวัณ โรค ได้ดี โดยวัสคุ ้คังกล่าวสามารถถูกขึ้นรูปในรูปแบบของแผ่นกรองอากาศซึ่งสามารถนำไปประยุกศ์ใช้ เป็นหน้ากากอนามัยเมื่อจำเป็นต้องสัมผัสกับเชื้อวัณ โรคได้ดี

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IAGI	P	A	G	F
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spun PAN fiber mats.

ABBREVIATIONS

A. anitratus Acinetobacter anitratus

A. baumannii Acinetobacter baumannii

A. calcoaceticus Acinetobacter calcoaceticus

A. Iwoffii Acinetobacter Iwoffii

AA Antioxidant activity

Abs Absorbance
AC Asiaticoside

a-ePLLA Aminolyzed PLLA fiber mats

Al Aluminum

ANOVA One-Way Analysis of Variance

ATCC American Type Culture Collection

B. cepacia Burkholderia cepacia

B. cereus Bacillus cereus

B. subtillis Bacillus subtillis

C. albican Candida albican

CA Caffeic acid

CA Centella asiatica

CA-g-ePLLA CA-grafted PLLA fiber mats

CLSI Clinical and Laboratory Standards Institute

-CONH- Amide linkage

-COO- Ester linkage

-COOH Carboxylic acid

DCM Dichloromethane

DMAc Dimethylacetamide

DMEM Dulbecco's modified Eagle's medium

DMF Dimethyl formamide

DMSO Dimethyl sulfoxide

DPPH 1,1-diphenyl-2-picrylhydrazyl

E. adenophorum Eupatorium adenophorum

E. coli Escherichia coli

E. faecalis Enterococcus faecalis

EDC 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide

El Electron ionization

ePLLA Electrospun PLLA fiber mats

e-spinning Electrospinning

e-spun Electrospun

FBS Fetal bovine serum

G. mangostana Garcinia mangostana

GC-MS Gas chromatography-mass spectrometry

GSH Glutathione

GSSG Oxidized glutathione

GTA Glutaraldehyde

h Hour

HDFa Human dermal fibroblast cell

HFIP 1,1,3,3-hexafluoro-2-propanol

HMD 1,6-hexamethylenediamine

IBU Ibuprofen

IND Indomethacin

K. oxytoca Klebsiella oxytoca

K. pneumonia Klebsiella pneumonia

L.monocytogenes Listeria monocytogenes

L929 Murine dermal fibroblasts

MDR-TB Multidrug-resistant tuberculosis

MH Mueller-Hinton

MIC Minimum Inhibitory concentration

MTT 3-(4,5-Dimethyl-2-thiazolyl)-2, 5-

diphenyltetrazolium bromide

Na₂HPO₄·7H₂O Disodiumhydrogenphosphateheptahydrate

Na₂S₂O₅ Sodium metabisulfite

NaCl Sodium chloride

NAP Naproxen

NF Nanofiltration

NH₂ Amino groups

NHS *N*-hydroxysuccinimide

o/w Oil in water

P. fluorescens Pseudomonas fluorescens

P. mirabilis Proterus mirabilis

P. aeruginasa Pseudomonas aeruginasa

PAN Polyacrylonitrile

PBS Phosphate buffer saline

PDLA Poly(D,L-lactic acid)

PEVA Poly(ethylene-co-vinyl acetate)

PLLA Poly(L-lactic acid)
RS Reactive species

S. pyogenes Strephylococcus pyogenes
S. agalactiae Strephylococcus agalactiae

S. aureus Staphylococcus aureus

S. boydii Shigella boydii

S. dysenteriae Shigella dysenteriae

S. enteritidis Salmononelia enteritidis

S. epidermidis Staphylococcus epidermidis

S. flexneri Shigella flexneri

S. marcescens Serratia marcescens

S. sonnei Shigella sonnei

S. typhi Salmononella typhi

SEM Scanning electron microscopy

SFM Serum-free medium

SUL Sulindac

TCPS Tissue-culture polystyrene plate

TFA Trifluoroacetic acid

US-FDA US Food and Drug Administration

UV-vis UV-visible spectrophotometer

V. cholerae Vibrio cholerae

Vit E vitamin E

WVTR Water vapor transmission rate

XPS X-ray photoelectron spectroscopy

LIST OF SYMBOLS

A	Area of bottle mouth
$A_{\text{control}} \\$	Absorbance values of the testing solution without the
	presence of the as-loaded or the as-released gallic acid
A_{sample}	Absorbance values of the testing solution with the presence of
	the as-loaded or the as-released gallic acid
k	Rate of the release of gallic acid that incorporates physical
	characteristics of the matrix/gallic acid system
Mt	Cumulative amount of gallic acid released at an arbitrary time t
$M\alpha$	Cumulative amount of the substance released at an infinite time
N	Exponent characterizing the mechanism with which the release
	kinetics
W	Weight of each specimen after submersion in each respective
	medium
W_d	Weight of dry hydrogels after immersed in phosphate buffer
	(PBS) solution
W_g	Weight of dry hydrogel after extraction
W_i	Initial weight of the sample in its dry state
W_o	Initial weight of dry hydrogel
W_s	Weight of swollen hydrogel
W_t	Weight of bottle after placed in oven
W_w	Weight of each fiber mat specimen
$ ho_{\scriptscriptstyle{S}}$	Bulk densities of the fiber mats
$ ho_{\!\scriptscriptstyle{W}}$	Density of water