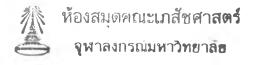
องค์ประกอบทางเคมีและฤทธิ์ทางชีวภาพของเอื้องทอง

นางสาวเกสินี ธนากรเมธา



วิทยานิพนธ์นี้เป็นส่วนหนึ่งของการศึกษาตามหลักสูตรปริญญาเภสัชศาสตรมหาบัณฑิต สาขาวิชาเภสัชเวท ภาควิชาเภสัชเวทและเภสัชพฤกษศาสตร์ คณะเภสัชศาสตร์ จุฬาลงกรณ์มหาวิทยาลัย ปีการศึกษา 2556 ลิขสิทธิ์ของจุฬาลงกรณ์มหาวิทยาลัย



Miss Kasinee Tanagornmeatar

A Thesis Submitted in Partial Fulfillment of the Requirements

for the Degree of Master of Science in Pharmacy Program in Pharmacognosy

Department of Pharmacognosy and Pharmaceutical Botany

Faculty of Pharmaceutical Sciences

Chulalongkorn University

Academic Year 2013

Copyright of Chulalongkorn University

Thesis Title By	CHEMICAL CONSTITUENTS AND BIOACTIVITIES OF DENDROBIUM ELLIPSOPHYLLUM Miss Kasinee Tanagornmeatar
Field of Study	Pharmacognosy
Thesis Advisor	Associate Professor Boonchoo Sritularak, Ph.D.
Thesis Co-Advisor	Professor Kittisak Likhitwitayawuid, Ph.D.
, , , , , , , , , , , , , , , , , , , ,	of Pharmaceutical Sciences, Chulalongkorn of the Requirements for the Master's Degree
(Assistant Professor Rui	Dean of the Faculty of Pharmaceutical Sciences
THESIS COMMITTEE	
S. Amoy	chairman Chairman
(Associate Professor Su	rattana Amnuoypol, Ph.D.)
Boonchoo Srit	ularah Thesis Advisor
(Associate Professor Bo	onchoo Sritularak, Ph.D.)
Cl. liket	Thesis Co-Advisor
(Professor Kittisak Likhit	twitayawuid, Ph.D.)
Witchinda Thank	y charolyath Examiner
(Assistant Professor Wit	chuda Thanakijcharoenpath, Ph.D.)Examiner
	ni Chanvorachote, Ph.D.)
Chaisak Chans	External Examiner

(Chaisak Chansriniyom, Ph.D.)

เกสินี ธนากรเมธา : องค์ประกอบทางเคมีและฤทธิ์ทางชีวภาพของเอื้องทอง. (CHEMICAL CONSTITUENTS AND BIOACTIVITIES OF *DENDROBIUM ELLIPSOPHYLLUM*) อ.ที่ปรึกษาวิทยานิพนธ์หลัก: รศ. ภก. ดร.บุญชู ศรีตุลารักษ์, อ.ที่ ปรึกษาวิทยานิพนธ์ร่วม: ศ. ภก. ดร.กิตติศักดิ์ ลิขิตวิทยาวุฒิ, 176 หน้า.

วัตถุประสงค์การวิจัยครั้งนี้เพื่อศึกษาองค์ประกอบทางเคมีจากสิ่งสกัดในชั้นเมทานอล รวมถึงฤทธิ์ทางชีวภาพของเอื้องทองทั้งต้น ซึ่งยังไม่เคยมีผู้ทำการวิจัยมาก่อน สามารถแยกสาร บริสุทธิ์ที่เคยมีรายงานมาก่อน 10 ชนิด ได้แก่ สารกลุ่มไบเบนซิล คือ moscatilin, 4,4'dihydroxy-3,5-dimethoxybibenzyl และ 4,5,4'-trihydroxy-3,3'-dimethoxybibenzyl, กลุ่มฟลาโวนอยด์ ได้แก่ (2S)-homoeriodictyol, (2S)-eriodictyol, chrysoeriol และ กล่มไดไฮโดรฟีแนนทรีน คือ 4,5-dihydroxy-2,3-dimethoxy-9,10dihydrophenanthrene, กลุ่มโครโมน คือ 5,7-dihydroxy-chromen-4-one และกลุ่มฟีนิล โพรพานอยด์ คือ phloretic acid ซึ่งสารทั้งหมดพิสูจน์โครงสร้างทางเคมีโดยเทคนิคสเปกโตรสโค ปี (UV, IR, MS, NMR) ร่วมกับการเปรียบเทียบข้อมูลที่เคยมีรายงานมาแล้ว จากผลการทดสอบ ฤทธิ์ทางชีวภาพพบว่า 4,5,4'-trihydroxy-3,3'-dimethoxybibenzyl และ luteolin มีฤทธิ์ระดับ ปานกลางยับยั้งเซลล์มะเร็งช่องปาก KB (IC $_{50}$ 61.93 และ 56.22 μ M ตามลำดับ) ซึ่งมีชุดควบคุม ผลบวก คือ ellipticine (IC50 4.99 μ M) และ doxorubicin (IC50 2.19 μ M) รวมทั้งใน เซลล์มะเร็งเต้านม MCF-7 (IC $_{50}$ 135.48 และ 68.01 μ M ตามลำดับ) ซึ่งมีชุดควบคุมผลบวก คือ tamoxifen (IC50 20.46 μ M) และ doxorubicin (IC50 26.29 μ M) นอกจากนี้ 4,4'-dihydroxy-3,5-dimethoxybibenzyl, 4,5,4'-trihydroxy-3,3'-dimethoxybibenzyl, chrysoeriol และ luteolin ยังมีฤทธิ์ต้านการแพร่กระจายของเซลล์มะเร็งปอด H292 เกิดการตายของเซลล์มะเร็ง และกระตุ้นการเกิดอะนอยคิส แบบอะพอพโทซิส โดย 4,5,4'-trihydroxy-3,3'dimethoxybibenzyl (IC $_{50}$ 96.56 μ M) เกิดการตายแบบอะพอพโทซิสมากที่สุดอย่างมีนัยสำคัญ เมื่อเปรียบเทียบกับกลุ่มควบคุม และกระตุ้นการเกิดอะนอยคิสออกฤทธิ์เร็วที่สุดที่ 6 ชั่วโมง ที่ ความเข้มข้น 1 และ 5 µM นอกจากนี้ 4,4'-dihydroxy-3,5-dimethoxybibenzyl ยังมีฤทธิ์ ระดับอ่อนในการต้านไวรัสเริมชนิด HSV-1 และ HSV-2 (IC $_{50}$ 313.61 \pm 40.40 และ 334.56 \pm 52.66 µM ตามลำดับ)

ภาควิชา เภสัชเวทและเภสัชพฤกษศาสตร์ สาขาวิชา เภสัชเวท

ปีการศึกษา 2556

ลายมือชื่อนิสิต <u>เกล็นี ชนากร่เพรา</u> ลายมือชื่อ อ.ที่ปรึกษาวิทยานิพนธ์หลัก ลายมือชื่อ อ.ที่ปรึกษาวิทยานิพนธ์ร่วม # # 5576201233 : MAJOR PHARMACOGNOSY

KEYWORDS: DENDROBIUM ELLIPSOPHYLLUM / CHEMICAL CONSTITUENTS /

BIOACTIVITIES

KASINEE TANAGORNMEATAR: CHEMICAL CONSTITUENTS AND BIOACTIVITIES OF *DENDROBIUM ELLIPSOPHYLLUM*. ADVISOR: ASSOC. PROF. BOONCHOO SRITULARAK, Ph.D., CO-ADVISOR: PROF. KITTISAK LIKHITWITAYAWUID, Ph.D., 176 pp.

The objective of this study was to investigate the chemical constituents and biological activities of Dendrobium ellipsophyllum, a plant with no previous reports. The results led to the isolation of ten known compounds, consisting of bibenzyls (moscatilin, 4,4'-dihydroxy-3,5-dimethoxybibenzyl, three trihydroxy-3,3'-dimethoxybibenzyl), four flavonoids ((2S)-homoeriodictyol, (2S)eriodictyol, chrysoeriol and luteolin), a dihydrophenanthrene, a chromone and a phenylpropanoids (4,5-dihydroxy-2,3-dimethoxy-9,10-dihydrophenanthrene, 5,7dihydroxy-chromen-4-one and phloretic acid, respectively). Their structures were determined by spectroscopic analysis (NMR, MS) and comparision with the previously reported data. The results from bioassays revealed that 4,5,4'trihydroxy-3,3'-dimethoxybibenzyl and luteolin had moderate cytotoxic activity against KB oral cavity cancer cells (IC $_{50}$ 61.93 and 56.22 μ M, respectively), as compared with the positive controls ellipticine (IC₅₀ 4.99 µM) and doxorubicin (IC₅₀ 1.53 μ M). The two compounds had cytotoxicity on MCF-7 breast cancer cells (IC₅₀ 135.48 and 68.01 μ M, respectively), in comparison with tamoxifen (IC₅₀ 20.46 μ M) and doxorubicin (IC₅₀ 26.29 µM). 4,4'-Dihydroxy-3,5-dimethoxybibenzyl, 4,5,4'trihydroxy-3,3'-dimethoxybibenzyl, chrysoeriol and luteolin showed anti-metastatic activity on H292 lung cancer cells, displaying apoptosis induction and anoikis sensitizing activities. 4,5,4'-Trihydroxy-3,3'-dimethoxybibenzyl (IC₅₀ 96.56 μM) possessed highest cytotoxic activity and the fastest action in sensitizing the cells to anoikis at the concentrations of 1 and 5 µM. Significant effects could be detected as early as 6 hours after exposure to the cells. Moreover, 4,4'-dihydroxy-3,5-dimethoxybibenzyl showed weak anti-herpes simplex virus activity against HSV-1 and HSV-2 with IC₅₀ 313.61 \pm 40.40 and 334.56 \pm 52.66 μ M, respectively.

Department: Pharmacognosy and

Pharmaceutical Botany

Field of Study: Pharmacognosy

Academic Year: 2013

Student's Signature Kasinee Tanagornmeatar

Advisor's Signature Romobico Snitularah
Co-Advisor's Signature K. Wellet

ACKNOWLEDGEMENTS

I would like extend to thank my advisor Associate Professor Dr. Boonchoo Sritularak of the Department of Pharmacognosy and Pharmaceutical Botany, Faculty Pharmaceutical Sciences, Chulalongkorn University who gave me knowledge, valuable advice, powerfulness, and resolving the problems, and my co-advisor Professor Dr. Kittisak Likhitwitayawuid of the Department of Pharmacognosy and Pharmaceutical Botany, Faculty Pharmaceutical Sciences, Chulalongkorn University who gave helpful advice and constructive suggestions for this master thesis.

I am grateful to Assistant Professor Dr. Pithi Chanvorachote and Dr. Chatchai Chaotham of Department Pharmacology, Faculty Pharmaceutical Sciences, Chulalongkorn University for research facilities of anti-metastasis assay and kind assistance.

I would like to thank Associate Professor Dr. Vimolmas Lipipun of the Department of Microbiology, Faculty Pharmaceutical Sciences, Chulalongkorn University for the test results of anti-herpes simplex activity.

I am also grateful to my thesis examination committee for useful advice and critical review. I would like to thank for the Graduate School of Chulalongkorn University for granting partial financial support to conduct this investigation.

Finally, I wish to express my deepest gratitude to my teachers in the Department of Pharmacognosy and Pharmaceutical Botany, Chulalongkorn University, and everyone who gave me truthful assistance in the preparation of this thesis.

THALABSTRACT	IV
ENGLISH ABSTRACT	V
ACKNOWLEDGEMENTS	vi
CONTENTS	vii
LIST OF TABLES	xi
LIST OF FIGURES	×ii
LIST OF SCHEMES	xvii
LIST OF ABBREVATIONS	xviii
CHAPTER I INTRODUCTION.	1
CHAPTER II HISTORICAL	10
Chemical constituents of <i>Dendrobium</i> spp	10
CHAPTER III EXPERIMENTAL	75
1. Source of plant materials	75
2. General techniques	75
2.1 Analytical thin-layer chromatography (TLC)	75
2.2 Column Chromatography	75
2.2.1 Vacuum liquid chromatography (VLC)	75
2.2.2 Flash column chromatography (FCC)	76
2.2.3 Medium pressure liquid chromatography (MPLC)	76
2.2.4 Gel filtration chromatography	77
2.3 Spectroscopy	77

P	Page
2.3.1 Mass spectra	77
2.3.2 Ultraviolet (UV) absorption spectra	77
2.3.3 Infrared (IR) spectra	77
2.3.4 Proton and carbon-13 nuclear magnetic resonance (¹ H and ¹³ C-NMR) spectra	78
2.3.5 Optical rotation	78
2.4 Solvents	78
3. Extraction and isolation	78
3.1 Extraction	78
3.2 Separation of methanol extract	78
3.2.1 Isolation of compound DE1 (5,7-Dihydroxy-chromen-4-one)	79
3.2.2 Isolation of compound DE2 (4,5- Dihydroxy-2,3-dimethoxy-9,10-dihydrophenanthrene)	79
3.2.3 Isolation of compound DE3 (Moscatilin)	79
3.2.4 Isolation of compound DE4 (4,4'-Dihydroxy-3,5-dimethoxybibenzyl). 7 and compound DE5 (4,5,4'-Trihydroxy-3,3'-dimethoxybibenzyl). 7	
3.2.5 Isolation of compound DE6 ((2S)-Homoeriodictyol)7	79
3.2.6 Isolation of compound DE7 ((2S)-Eriodictyol)	30
3.2.7 Isolation of compound DE8 (Chrysoeriol) and compound DE9 (Phloretic acid)8	30
3.2.8 Isolation of compound DE10 (Luteolin)	30
4. Physical and spectral data of isolated compounds	34
4.1 Compound DE1 (5,7-Dihydroxy-chromen-4-one)	34
4.2 Compound DE2 (4,5- Dihydroxy-2,3-dimethoxy-9,10-dihydrophenanthrene)	
4.3 Compound DE3 (Moscatilin)	
4.4 Compound DE4 (4,4'-Dihydroxy-3,5-dimethoxybibenzyl)	
4.5 Compound DE5 (4,5,4'-Trihydroxy-3,3'-dimethoxybibenzyl)	

	Page
4.6 Compound DE6 ((2S)-Homoeriodictyol)	86
4.7 Compound DE7 ((2S)-Eriodictyol)	86
4.8 Compound DE8 (Chrysoeriol)	86
4.9 Compound DE9 (Phloretic acid)	87
4.10 Compound DE10 (Luteolin)	87
5. Determination of cytotoxicity	88
6. Determination of anti-metastatic activity	89
6.1 Cells and reagents	89
6.2 Anoikis and cell viability	89
6.3 Apoptosis nuclear staining assay	90
6.4 Statistical analysis	90
7. Determination of anti-herpes simplex virus activity	90
7.1 Viruses and cells	90
7.2 Plaque reduction assay	91
CHAPTER IV RESULTS AND DISCUSSION	92
1. Structure characterization of isolated compounds	92
1.1 Structure determination of compound DE1	92
1.2 Structure determination of compound DE2	95
1.3 Structure determination of compound DE3	97
1.4 Structure determination of compound DE4	99
1.5 Structure determination of compound DE5	. 101
1.6 Structure determination of compound DE6	. 103
1.7 Structure determination of compound DE7	. 105
1.8 Structure determination of compound DE8	. 107
1.9 Structure determination of compound DE9	.109
1.10 Structure determination of compound DE10	.111
2. Cytotoxic activity on KB oral cavity and MCF-7 breast cancer cells	113

	Page
3. Cytotoxicity on H292 lung cancer cells	114
3.1 Apoptosis induction effect of the compounds	115
3.2 Anoikis sensitizing activity	117
4. Anti-Herpes Simplex activity	119
CHAPTER V CONCLUSION	120
REFERENCES	121
APPENDIX	129
VITA	176

LIST OF TABLES

Page
Table 1 Distribution of chemical constituents in the genus <i>Dendrobium</i>
Table 2 NMR Spectral data of compound DE1 (acetone- d_6) and 5,7-dihydroxy-
chromen-4-one (MeOH- d_4)
Table 3 NMR Spectral data of compound DE2 (CDCl ₃)
Table 4 NMR Spectral data of compound DE3 (acetone- d_6) and moscatilin (CDCl $_3$) 98
Table 5 NMR Spectral data of compound DE4 (acetone- d_6) and 4,4 $^\prime$ -dihydroxy-3,5-
dimethoxybibenzyl (CD ₃ OD)100
Table 6 NMR Spectral data of compound DE5 (acetone- d_6) and 4,5,4 $^\prime$ -trihydroxy-3,3 $^\prime$ -
dimethoxybibenzyl (CDCl ₃)102
Table 7 NMR Spectral data of compound DE6 (acetone- d_6) and (25)-homoeriodictyol
[¹ H NMR (acetone-d ₆) and ¹³ C NMR (DMSO-d ₆)]
Table 8 NMR Spectral data of compound DE7 (acetone- d_6) and (2S)-eriodictyol
(DMSO-d ₆)
Table 9 NMR Spectral data of compound DE8 (acetone- d_6) and chrysoeriol108
Table 10 NMR Spectral data of compound DE9 (acetone- d_6) and phloretic acid
(CD ₃ OD)
Table 11 NMR Spectral data of compound DE10 (acetone- d_6) and luteolin (DMSO- d_6)
Table 12 IC ₅₀ Values (µM) for cytotoxicity on KB and MCF-7 cells113
Table 13 ICs Values (uM) for cytotoxicity on H292 cells

LIST OF FIGURES

	Page
Figure 1 Dendrobium ellipsophyllum Tang & Wang	9
Figure 2 Structures of compounds previously isolated from <i>Dendrobium</i> species	5 43
Figure 3 (A) Percentage of cell apoptosis of compounds DE4, DE5, DE8 and DE10	Э
obtained from Hoechst 33342/propidium iodide (PI) assays. Data represent the I	mean
\pm SD (n = 3). * P < 0.05 versus untreated control cells. (B) Morphology of apoptor	otic
nuclei stained with Hoechst 33342 and propidium iodide	116
Figure 4 Anoikis sensitizing activity of compounds DE4 (A), DE5 (B), DE8 (C) and D)E10
(D) as assessed by anoikis assay. The cells were exposed with various concentra	itions
of each compound (0-5 $\mu\text{M})$ and cell viability was determined by XTT assay at t	the
indicated time. Data represent the mean \pm SD (n = 3). * P < 0.05 versus untreated	ed
control cells	118
Figure 5 Mass spectrum of compound DE1	130
Figure 6 IR Spectrum of compound DE1	130
Figure 7 UV Spectrum of compound DE1	130
Figure 8 ¹ H-NMR (300 MHz) Spectrum of compound DE1 (acetone-d ₆)	131
Figure 9 C-NMR (75 MHz) Spectrum of compound DE1 (acetone- d_6)	131
Figure 10 COSY Spectrum of compound DE1 (acetone- d_6)	132
Figure 11 DEPT 135 Spectrum of compound DE1 (acetone- d_6)	132
Figure 12 Mass spectrum of compound DE2	133
Figure 13 IR Spectrum of compound DE2	133
Figure 14 UV spectrum of compound DE2	133
Figure 15 H-NMR (300 MHz) Spectrum of compound DE2 (CDCL)	134

Figure	16 C-NMR (75 MHz) Spectrum of compound DE2 (CDCl ₃)	. 134
Figure	17 HMBC Spectrum of compound DE2 (CDCl ₃)	. 135
Figure	18 HSQC Spectrum of compound DE2 (CDCl ₃)	. 136
Figure	19 NOESY Spectrum of compound DE2 (CDCl ₃)	. 137
Figure	20 Mass spectrum of compound DE3	. 138
Figure	21 IR Spectrum of compound DE3	. 138
Figure	22 UV Spectrum of compound DE3	. 139
Figure	23 H-NMR (500 MHz) Spectrum of compound DE3 (acetone- <i>d</i> ₆)	. 139
Figure	24 H-NMR (500 MHz) Spectrum of compound DE3 (acetone- $d_{\acute{a}}$)	. 140
Figure	25 H-NMR (500 MHz) Spectrum of compound DE3 (acetone- d_6)	. 140
Figure	26 C-NMR (125 MHz) Spectrum of compound DE3 (acetone-d ₆)	. 141
Figure	27 C-NMR (125 MHz) Spectrum of compound DE3 (acetone- d_6)	. 141
Figure .	28 C-NMR (125 MHz) Spectrum of compound DE3 (acetone- d_6)	.142
Figure	29 Mass spectrum of compound DE4	. 143
Figure	30 IR Spectrum of compound DE4	. 143
Figure	31 UV Spectrum of compound DE4	. 144
Figure	32 H-NMR (500 MHz) Spectrum of compound DE4 (acetone- d_6)	. 144
Figure	33 H-NMR (500 MHz) Spectrum of compound DE4 (acetone- d_6)	. 145
Figure	34 H-NMR (500 MHz) Spectrum of compound DE4 (acetone- d_6)	. 145
Figure	35 C-NMR (125 MHz) Spectrum of compound DE4 (acetone- d_6)	. 146

Figure	36	HMBC Spectrum of compound DE4 (acetone-d ₆)	146
Figure	37	HMBC Spectrum of compound DE4 (acetone- d_6)	147
Figure	38	HMBC Spectrum of compound DE4 (acetone-d ₆)	147
Figure	39	HMBC Spectrum of compound DE4 (acetone- d_6)	148
Figure	40	Mass spectrum of compound DE5	149
Figure	41	IR Spectrum of compound DE5	149
Figure	42	UV Spectrum of compound DE5	150
Figure	43	H-NMR (500 MHz) Spectrum of compound DE5 (acetone- d_6)	150
Figure	44	1 H-NMR (500 MHz) Spectrum of compound DE5 (acetone- d_6)	151
Figure	45	1 H-NMR (500 MHz) Spectrum of compound DE5 (acetone- d_6)	151
Figure	46	C-NMR (125 MHz) Spectrum of compound DE5 (acetone- d_6)	152
Figure	47	C-NMR (125 MHz) Spectrum of compound DE5 (acetone- d_6)	152
Figure	48	C-NMR (125 MHz) Spectrum of compound DE5 (acetone- d_6)	153
Figure	49	NOESY Spectrum of compound DE5 (acetone- d_6)	153
Figure	50	Mass spectrum of compound DE6	154
Figure	51	IR Spectrum of compound DE6	154
Figure	52	UV Spectrum of compound DE6	155
Figure	53	H-NMR (500 MHz) Spectrum of compound DE6 (acetone- d_6)	155
Figure	54	t H-NMR (500 MHz) Spectrum of compound DE6 (acetone- d_6)	156
Figure	55	t H-NMR (500 MHz) Spectrum of compound DE6 (acetone- d_6)	156

Figure	56	H-NMR (500 MHz) Spectrum of compound DE6 (acetone-d ₆)	157
Figure	57	H-NMR (500 MHz) Spectrum of compound DE6 (acetone-d ₆)	157
Figure	58	C-NMR (125 MHz) Spectrum of compound DE6 (acetone- d_6)	158
Figure	59	NOESY Spectrum of compound DE6 (acetone- $d_{_{6}}$)	158
Figure	60	Mass spectrum of compound DE7	159
Figure	61	IR Spectrum of compound DE7	159
Figure	62	UV Spectrum of compound DE7	159
Figure	63	H-NMR (500 MHz) Spectrum of compound DE7 (acetone- d_6)	160
Figure	64	13 C-NMR (125 MHz) Spectrum of compound DE7 (acetone- d_{6})	160
Figure	65	DEPT 135 Spectrum of compound DE7 (acetone- d_6)	161
Figure	66	HSQC Spectrum of compound DE7 (acetone- d_6)	161
Figure	67	NOESY Spectrum of compound DE7 (acetone- d_6)	162
Figure	68	HMBC Spectrum of compound DE7 (acetone- d_6)	162
Figure	69	HMBC Spectrum of compound DE7 (acetone- d_6)	163
Figure	70	Mass spectrum of compound DE8	164
Figure	71	IR Spectrum of compound DE8	164
Figure	72	UV Spectrum of compound DE8	165
Figure	73	H-NMR (500 MHz) Spectrum of compound DE8 (acetone- d_6)	165
Figure	74	H-NMR (500 MHz) Spectrum of compound DE8 (acetone-d ₆)	166
Figure	75	C-NMR (125 MHz) Spectrum of compound DE8 (acetone- d_6)	166
Figure	76	DEPT135 Spectrum of compound DE8 (acetone-d ₆)	167

Figure	77	NOESY Spectrum of compound DE8 (acetone-d ₆)	167
Figure	78	NOESY Spectrum of compound DE8 (acetone- d_6)	168
Figure	79	Mass spectrum of compound DE9	169
Figure	80	IR Spectrum of compound DE9	169
Figure	81	UV Spectrum of compound DE9	169
Figure	82	H-NMR (300 MHz) Spectrum of compound DE9 (acetone- d_6)	170
Figure	83	13 C-NMR (75 MHz) Spectrum of compound DE9 (acetone- $\frac{d}{6}$)	170
Figure	84	DEPT 135 Spectrum of compound DE9 (acetone- d_6)	171
Figure	85	Mass spectrum of compound DE10	172
Figure	86	IR Spectrum of compound DE10	172
Figure	87	UV Spectrum of compound DE10	173
Figure	88	H-NMR (500 MHz) Spectrum of compound DE10 (acetone- d_6)	173
Figure	89	H-NMR (500 MHz) Spectrum of compound DE10 (acetone- d_{ϵ})	174
Figure	90	C-NMR (125 MHz) Spectrum of compound DE10 (acetone- d_6)	174
Figure	91	C-NMR (125 MHz) Spectrum of compound DE10 (acetone-d _g)	175
Figure	92	DEPT135 Spectrum of compound DE10 (acetone-d ₆)	175

LIST OF SCHEMES

Page
Sheme 1 Separation of the MeOH extract of <i>Dendrobium ellipsophyllum</i> 81

LIST OF ABBREVATIONS

Acetone- d_6 = Deuterated acetone

 α = Alpha Beta

br s = Broad singlet (for NMR spectra)

°C = Degree Celsius

CC = Column chromatography

CDCl₃ = Deuterated chloroform

CD₃OD = Deuterated methanol

cm = Centimeter

¹³C NMR = Carbon-13 Nuclear Magnetic Resonance

d = Doublet (for NMR spectra)

dd = Doublet of doublets (for NMR spectra)

 δ = Chemical shift

2D-NMR = Two dimensional Nuclear Magnetic Resonance

DMSO-d₆ = Deuterated dimethylsulfoxide

ESIMS = Electrospray Ionization Mass Spectrometry

EtOAc = Ethyl acetate

FCC = Flash Column Chromatography

g = Gram

GF = Gel Filtration Chromatography

Glc = Glucose

Hr = Hour

¹H-NMR = Proton Nuclear Magnetic Resonance

HR-ESI-MS = High Resolution Electrospray Ionization Mass Spectrometry

HSV-1 = Herpes Simplex Virus type 1 HSV-2 = Herpes Simplex Virus type 2

Hz = Hertz

IC₅₀ = Concentration exhibiting 50% inhibition

IR = Infrared spectrum

J = Coupling constant

Kg = Kilogram

 λ_{max} = Wavelength at maximal absorption

ε = Molar absorptivity

 $[M+H]^{\dagger}$ = Pseudomolecular ion

 $[M+Na]^{\dagger}$ = Sodium-adduct molecular ion

m = Multiplet (for NMR spectra)

MeOH = Methanol

mg = Milligram

mL = Milliliter

 μ g = Microgram

μg/mL = Microgram per milliliter

μL = Microliter

 μ M = Micromolar

mm = Millimeter

MPLC = Medium Pressure Liquid Column Chromatography

MS = Mass spectrum

m/z = Mass to charge ratio

nm = Nanometer

NMR = Nuclear Magnetic Resonance

NOESY = Nuclear Overhauser Effect Spectroscopy

ppm = Part per million

Rha = Rhamnose

s = Singlet (for NMR spectra) t = Triplet (for NMR spectra)

TLC = Thin Layer Chromatography

UV-VIS = Ultraviolet and Visible spectrophotometry

VLC = Vacuum Liquid Column Chromatography