

## Chapter 4

### Results and Discussions

#### 4.1 Morphometric analysis

All specimens were measured for 5 parameters and weight as well as were identified for their age level and sex. The data were shown in Table 1. Results comparing the two subspecies were obtained using analysis of variance

The wing length, tarsometatarsus length, base of beak to nape length, third digit length showed no difference between two groups. But the beak length showed a significant different between two groups (Beak length mean= 1.410, F value=10.06 at  $p<0.05$ ). See Appendix III.

The beak length compared between two subspecies showed the difference, but all specimens are caged birds in captivity, so the result might have some bias due to their changed feeding behavior from their wild habitat, or differences in husbandry between the two sample groups.

Morphometric analysis using more characters should have been done for a larger number of animal both in the wild and captive for differentiation of their morphology, and might have produced more significant taxonomic differences.



#### 4.2 Protocol used for DNA extraction.

2 protocols for mtDNA extraction were used; first the Chelex extraction and second, the classical phenol-chloroform extraction. Ethidium bromide stained agarose gel (Figure 4.1) shows the DNA extracted from Chelex compare to phenol-chloroform extracted product. Chelex extracted product can not be observed on the gel while concentrated Phenol-Chloroform extracted product gave a clear signal of DNA quantity obtained from the extraction.

The comparative result was also observed by the first PCR product on 2% agarose gel electrophoresis. Using the Chelex extract to add PCR mixture as template DNA gave no signal from PCR. Total of 20 samples has been tried and double extraction trials were done.

PCR signals were successfully obtained using phenol-chloroform extracts as an alternative template in PCR mixture. One microliter of Bovine Serum Albumin (BSA) needs to be added as an inhibitors terminator (Cooper, 1994) to all PCR mixtures, otherwise no signals can be obtained.

Cooper (1994) mentioned that the Heme and cytochrome from blood are the major PCR inhibitors. Meckvichai (1997) succeeded in using Chelex extracts for cytochrome b gene amplification but it could not be used for D-loop amplification in this study.

A concentrated purified phenol-chloroform extract is recommended to use as a template DNA for chicken D-loop amplification if sample used are bloodstains.

### 4.3 DNA amplification from D-loop region

Optimization of PCR condition, using the universal cytochrome b primer for amplification of the some samples of Chelex extracted product and the positive bands were obtained. The PCR mixture and cycles was described in chapter 2.

Using the Chelex extracted solution as template DNA for the PCR reaction gave all negative results when using D-loop primer for amplification.

The alternative phenol-chloroform extracted products were used as an improved substitution. These samples gave positive signals of the D-loop amplified product (Figure 4.2 a and b) but Bovine Serum Albumin (BSA, Sigma) was needed for every PCR reaction tube as inhibitor terminators.

PCR primers was a chicken specifically designed and could amplify the entire 1254 bp fragment, the whole D-loop region.

M 1 2



Figure4.1 Ethidium bromide stained agarose gel (1.5%) show the DNA extracted from Chelex compare to phenol-chloroform extracted product. M= $\phi$ X174 DNA marker, Lane 1= Chelex extracted product can not be observed on the gel, Lane 2= Phenol-Choroform extracted product (concentrated) gave a clear signal of DNA quantity obtained.

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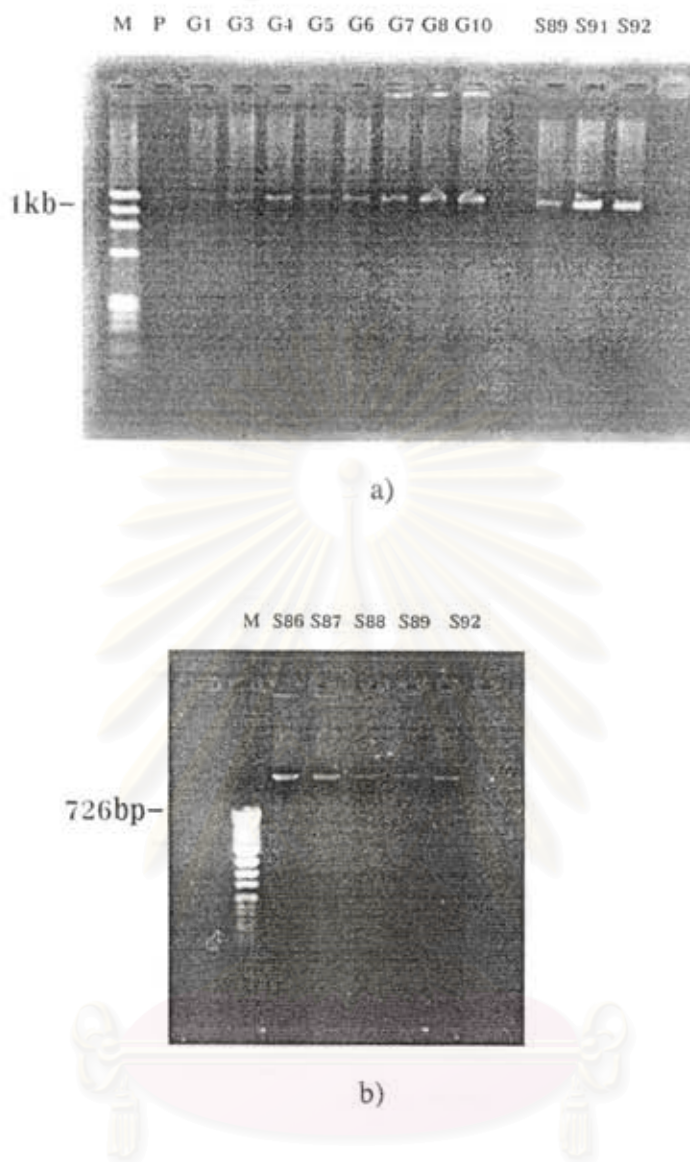


Figure 4.2 Ethidium bromide stained gel showing the positive PCR signal of amplified Chicken D-loop gene product.

- a) M=ØX174 Hae III DNA marker, G1, 3, 4,5, 6, 7, 8 and 10= *Gallus gallus gallus* samples, S89, 91, 92= *G. g. spadiceus* samples, P = positive control.
- b) M=ØX174 Hinf I DNA marker, S86, 87, 88, 89 and 92= *G. g. spadiceus* samples.

A G C T



Figure 4.3 The exposed X-ray film from  $^{32}\text{P}$ -labelled chicken D-loop DNA sequence.

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#### 4.4 Second PCR generating ss-DNA

For manual DNA sequencing using chain-terminating inhibitor (Sanger, 1977), it is better to generate the single-stranded DNA from the first PCR product. Gyllensten (1988) described the protocol for asymmetric PCR that it could generate the single-stranded DNA for the sequencing. The result is not shown here.

#### 4.5 DNA sequence analysis

225 bp sequence from 16 samples, 6 *Gallus gallus spadiceus* and 10 *G. g. gallus*, were analyzed. The obtained sequences were mitochondrial D-loop L-chains (base position L41-L323 compare to *Gallus domesticus* by Desjardins and Morais, 1990 with 3 bases missing). The nucleotide composition of the D-loop sequence were reported by Desjardins and Morais (1990) as; A=26.7, G=13.3, C=26.3 and T=33.7.

There are 5 sites of variation in *G. g. gallus* and 15 sites in *G. g. spadiceus* which means the genetic variation in the observed area of *G. g. spadiceus* is three times higher than that of *G. g. gallus*.

Within subspecies *G. g. gallus*, 5 transitions (TS) were found. There is no transvertional (TV) nucleotide substitution in this subspecies. The genetic distance within this subspecies= 0.0000-0.0225. Sequence divergence varies from 0 to 2.25%. Fumihito et al (1994) reported the variation of D-loop region of 2 *G. g. gallus* from Thailand that there are 1.25 % sequence divergence.

Within subspecies *G. g. spadiceus*, 10 transitions and 5 transversions were found. The TS: TV ratio is 2:1. The sequence number S84 and S85 showed 100% homology along 225 bp alignment so they were treated into the same taxa when using for the parsimonious analysis. The genetic distance within this subspecies=



0.0000-0.0654. Sequence divergence varies from 0 to 6.54% while Fumihito et al (1994) reported 4.25 % from one sample of *G. g. spadiceus* from Thailand.

There are 9 sites variable between two subspecies. The TS: TV ratio is 8:1. The genetic distances between two subspecies= 0.0134-0.0800. Genetic distances between groups' seem to be larger than that of within each group.

Using the published Japanese quail (*Coturnix coturnix japonica*) as an outgroup reference, 20 variation site were found after aligning the sequences by CLUSTAL V in PHYLIP program version 3.57c (Felsenstein, 1993). The obtained sequences showed the CCC base triplets, which are shown underlined in Figure 4.4, that are similar to the sequences published by Fumihito et al. (1994) but were missed from domestic chicken (*Gallus domesticus*) D-loop that was published by Desjardins and Morais (1990).

All sequences were analyzed for their phylogenetic relationship using the genetic distance data (Kimura's 2 parameters) shown in Table 3.2. The tree was constructed with neighbor-joining method using PHYLIP program version 3.572c and was shown in Figure 4.6. The samples were divided in two groups. One with all the *G. g. spadiceus* (S84, 85, 86, 87, 88 AND 92) and *G. g. gallus* number G9, and another with 9 *G. g. gallus* samples (G 1, 2, 3, 4, 5, 6, 7, 8 and 10)

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## CLUSTAL V multiple sequence alignment

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JAPQU      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
1994      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
S84      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
S85      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
S86      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGGGCATACATTT
S87      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGGGCATACATTT
S88      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGGGCATACATTT
S92      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGGGCATACATTT
G1      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G2      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G3      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G4      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G5      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G6      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G7      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G8      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G9      CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
G10     CCCCTTTCCCCCCAGGGGGGGTATACTATGCATAATCGTGCATACATTT
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JAPQU      ATATTCCACATATACTATGGTACCGGTAATATATATTATATACGFACTAA
1994      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
S84      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
S85      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
S86      ATATACCACATATATTATGGCACC GGTAATATATACTATATATGFACTAA
S87      ATATACCACATATATTATGGCACC GGTAATATATACTATATATGFACTAA
S88      ATATACCACATATATTATGGCACC GGTAATATATACTATATATGFACTAA
S92      ATATATCCCATATATTATGGCACC GATAATATATACTATATTGCCACTAA
G1      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G2      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G3      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G4      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G5      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G6      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G7      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G8      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G9      ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
G10     ATATACCACATATATTATGGTACCGGTAATATATACTATATATGFACTAA
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JAPQU	-----AACCA-TAC--G TTCACC
1994	AGCTCCAAACCACTACCAAGTCACC
S84	AGCTCCAAACCACTACCAAGTCACC
S85	AGCTCCAAACCACTACCAAGTCACC
S86	AGCTCCAAACCACTACCAAGTCACC
S87	AGCTCCAAACCACTACCAAGTCACC
S88	AGCTCCAAACCACTACCAAGTCACC
S92	AGCTCCAAACCACTACCAAGTCACC
G1	AGCTCCAAACCACTACCAAGTCACC
G2	AGCTCCAAACCACTACCAAGCCACC
G3	AGCTCTAAACCACTACCAAGCCACC
G4	AGCTCTAAACCACTACCAAGCCACC
G5	AGCTCTAAACCACTACCAAGCCACC
G6	AGCTCTAAACCACTACCAAGCCACC
G7	AGCTCTAAACCACTACCAAGCCACC
G8	AGCTCCAAACCACTACCAAGTCACC
G9	AGCTCCAAACCACTACCAAGTCACC
G10	AGCTCTAAACCACTACCAAGCCACC
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Figure 4.4 L-chain sequences of 225 bases of the mitochondrial control region from *G. g. gallus* and *G. g. spadiceus* using the published Japanese quail (*JAPQU*, *Coturnix coturnix japonica*) as an outgroup. 1994 is Thai Red junglefowl published sequence by Fumihito et al., 1994. CCC base triplets underlined in the first section were bases missed in two previous publications by Desjardins and Morais 1990 but similar to the sequence by Fumihito et al., 1994. The stars under the blocks mark the homology of nucleotides.

From the DNA sequence analysis, it is shown that *Gallus gallus spadiceus* samples have higher divergence in their nucleotide sequence than that of *G. g. gallus*. Using the assumption that mutation occur through time of evolution, it is possible that the *G. g. gallus* might have evolved from *G. g. spadiceus*.

The wider range of distribution and more number of population and individuals may cause the higher genetic variation in *G. g. spadiceus* even small number of samples were collected.

From tree drawn by neighbor-joining method, sample number G9 that fall into the same group with other subspecies might have had an evidence of genetic hybridization with *G. g. spadiceus* but need confirmation by nuclear DNA profile.

The most parsimonious tree showed the distinctive two groups with the unclear polytomies, which are S84, S85, G1, G2, G8 and G9. These samples might have common sequence characters in the observed 225 bp even their morphology are different. The sequence number S84 and S85 that showed 100 % homology might have common ancestor.

Regarding the conservation genetics, the animal number G8 and G9 may not be appropriate to use as a parent stock for the *gallus* lineage production since their DNA profile were closed to their *spadiceus* counterparts. All other sample would be a good stock and should have been preserved.

Table 2. Estimated genetic distances among 16 samples, published sequence (1994) and outgroup (Japq, *Coturnix coturnix japonica*) obtained from 225 bp of D-loop sequence.

Japq	1994	S84	S85	S86	S87	S88	S92	G1	G2	G3	G4	G5	G6	G7	G8	G9	G10
Japq	-																
1994	0.0844	-															
S84	0.0910	0.0181	-														
S85	0.0910	0.0181	0.0000	-													
S86	0.1160	0.0414	0.0226	0.0226	-												
S87	0.1094	0.0366	0.0180	0.0180	0.0045	-											
S88	0.1160	0.0604	0.0411	0.0411	0.0272	0.0226	-										
S92	0.1556	0.0805	0.0605	0.0605	0.0462	0.0414	0.0654	-									
G1	0.0910	0.0135	0.0045	0.0045	0.0272	0.0226	0.0458	0.0654	-								
G2	0.0970	0.0181	0.0089	0.0089	0.0318	0.0271	0.0505	0.0702	0.0045	-							
G3	0.0970	0.0226	0.0134	0.0134	0.0365	0.0317	0.0552	0.0751	0.0089	0.0045	-						
G4	0.0970	0.0365	0.0180	0.0180	0.0411	0.0364	0.0600	0.0800	0.0225	0.0180	0.0134	-					
G5	0.0970	0.0365	0.0180	0.0180	0.0411	0.0364	0.0600	0.0800	0.0225	0.0180	0.0134	0.0000	-				
G6	0.0970	0.0365	0.0180	0.0180	0.0411	0.0364	0.0600	0.0800	0.0225	0.0180	0.0134	0.0000	0.0000	-			
G7	0.0970	0.0365	0.0180	0.0180	0.0411	0.0364	0.0600	0.0800	0.0225	0.0180	0.0134	0.0000	0.0000	0.0000	-		
G8	0.0910	0.0135	0.0045	0.0045	0.0272	0.0226	0.0458	0.0654	0.0000	0.0045	0.0089	0.0225	0.0225	0.0225	0.0225	-	
G9	0.0910	0.0272	0.0089	0.0089	0.0318	0.0271	0.0505	0.0702	0.0134	0.0180	0.0225	0.0180	0.0180	0.0180	0.0180	0.0134	-
G10	0.0970	0.0318	0.0134	0.0134	0.0365	0.0317	0.0552	0.0751	0.0180	0.0134	0.0089	0.0045	0.0045	0.0045	0.0045	0.0180	0.0225







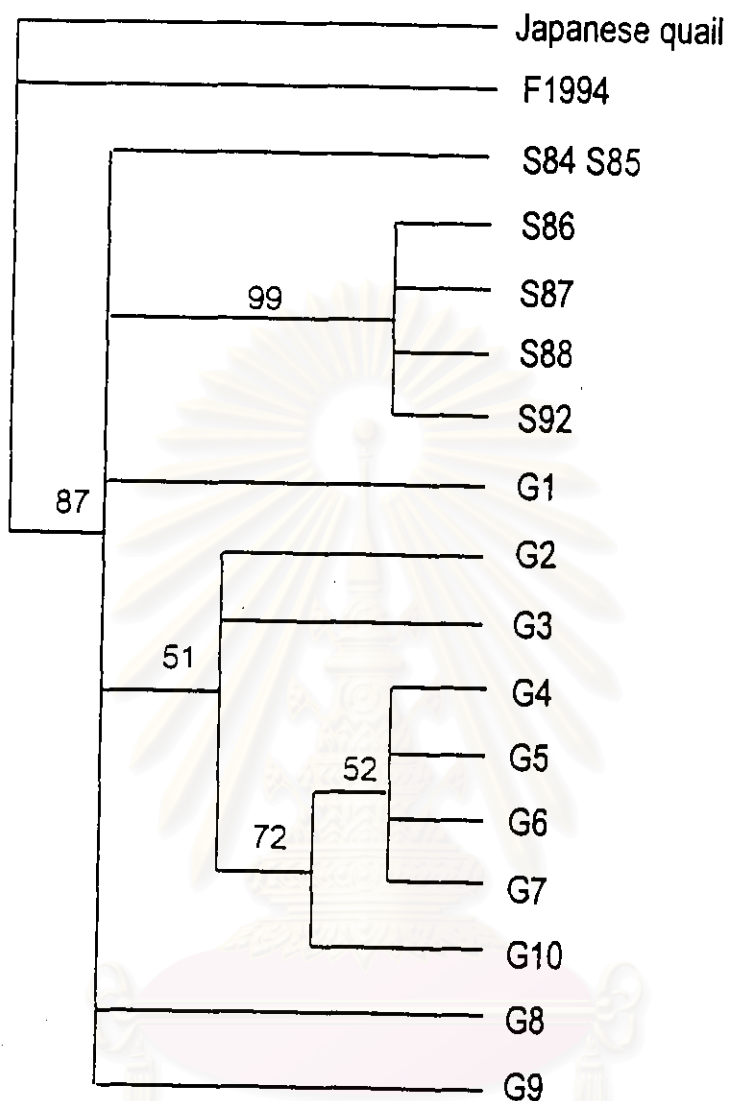


Figure 4.7 The most parsimonious tree drawn from PAUP version 3.0 shows the separation of two group with some polytomies which are not support by 50% bootstrapping criteria.