

REFERENCE

1. Bakir, F., Damluji, S. F., Amin-Zaki, L., Murtadha, M., Khalidi, A., Al-Rawi, Y., Tikriti, S., Dhahir, H. I., Clarkson, T.W., Smith, J. C., and Doherty, R. A. Methylmercury poisoning in Iraq. Science 181(1973): 230-241.
2. Gage, J. C. The trace determination of phenyl- and methylmercury salts in biological material. Analyst 86(1961): 457-458.
3. Westoo, G. Determination of methylmercury compounds in foodstuffs. I. methylmercury compounds in fish, identification and determination. Acta. Chem. Scand. 20(1966): 2131-2137.
4. Beauchemin, D., Siu, K. W. M., and Berman, S. S. Determination of organomercury in biological reference materials by inductively coupled plasma mass spectrometry using flow injection analysis. Anal. Chem. 60(1988): 2587-2590.
5. Woller, A., Garraud, H., Martin, F., Donard, O. X., and Fodor, P. Determination of total mercury in sediments by microwave-assisted digestion-flow injection-inductively coupled plasma mass spectrometry. J. Anal. At. Spectrom. 12(1997): 53-56.
6. Brahma, N. K., Corns, W. T., Stockwell, P. B., Ebdon, L., and Evans, E. H. At line determination of mercury in process streams using atomic fluorescence spectrometry. J. Anal. At. Spectrom. 12(1997): 611-616.
7. Jian, W. and McLeod, C. W. Simple oxidative pretreatment for determination of organomercury by cold vapor atomic fluorescence spectrometry. Mikrochim. Acta. 109(1992): 117-120.
8. Goto, M., Shibakawa, T., Arita, T., and Ishii, D. Continuous monitoring of total and inorganic mercury in wastewater and other waters. Anal. Chim. Acta. 140(1982): 179-185.

9. Hanna, C. P., Tyson, J. F., and McIntosh, S. Determination of total mercury in waters and urine by flow injection atomic absorption spectrometry procedures involving on-and off-line oxidation of organomercury species. Anal. Chem. 65(1993): 653-656.
10. Atallah, R. H. and Kalman, D. A. Selective determination of inorganic mercury and methylmercury in tissues by continuous flow and cold vapor atomic absorption spectrometry. J. Anal. Toxicol. 17(1993): 87-92.
11. Vargas, M. C. and Romero, R. A. Mercury determination by cold vapor atomic absorption spectrometry in several biological indicators from lake maracaibo, Venezuela. Analyst 117(1992): 645-647.
12. Rezende, M. C. R., Canpos, R. C., and Curtius, A. J. Speciation of mercury in fish samples by solvent extraction, methylmercury reduction directly in the organic medium and cold vapor atomic absorption spectrometry. J. Anal. At. Spectrom. 8(1993): 247-251.
13. Magos, L. Selective atomic absorption determination of inorganic mercury and methylmercury in undigested biological samples. Analyst 96(1971): 847-853.
14. Capelli, R., Fezia, C., Franchi, A., and Zanocchi, G. Extraction of methylmercury from fish and its determination by atomic-absorption spectroscopy. Analyst 104 (1979): 1197-1200.
15. Minagawa, K., Takizawa, Y., and Kifune, I. Determination of very low levels of inorganic and organic mercury in natural waters by cold-vapor atomic absorption spectrometry after preconcentration on a chelating resin. Anal. Chem. Acta. 115(1980): 103-110.
16. Oda, C. E. and Ingle, J. D. Jr. Continuous flow cold vapor atomic absorption determination of mercury. Anal. Chem. 53(1981): 2030-2033.
17. Fernandez, B. A., Campa, M. R. F., and Sanz-medel, A. Improvement in mercury cold vapor atomic techniques by resorting to organized assemblies and on-line membrane drying of vapor. J. Anal. At. Spectrom. 8(1993): 1097-1102.
18. Westoo, G. Determination of methylmercury compounds in foodstuffs. II. determination of methylmercury in fish, egg, meat, and liver. Acta Chem. Scand. 21(1967): 2131-2137.

19. Westoo, G. Determination of methylmercury salts in various kinds of biological material. Acta Chem.Scand. 22(1968): 2131-2137.
20. Filho, N. L. D., Gushikem, Y., and Polito, W. 2-Mercaptobenzothiazole clay as matrix for sorption and preconcentration of some heavy metals from aqueous solution. Anal. Chim. Acta. 306(1995): 167-176.
21. Liang, L. and Bloom, N. S. Determination of total mercury by single-stage gold amalgamation with cold vapor atomic spectrometric detection. J. Anal. At. Spectrom. 8(1993): 591-594.
22. Frech, W., Baxter, D. C., Dyvik, G., and Dybdahl, B. On the determination of total mercury in natural gases using the amalgamation technique and cold vapor atomic absorption spectrometry. J. Anal. At. Spectrom. 10(1995): 769-775.
23. Dumarey, R., Dams, R., and Hoste, J. Comparison of the collection and desorption efficiency of activated charcoal, silver, and gold for the determination of vapor-phase atmospheric mercury. Anal. Chem. 57(1985): 2638-2643.
24. Ellis, L. A. and Roberts, D. J. Novel method to determine mercury in sediment using a gold coated dual 'bent' tube atom trap. J. Anal. At. Spectrom. 11(1996): 1063-1066.
25. Hanna, C. P., Haigh, P. E., Tyson, J. F., and McIntosh, S. Examination of separation efficiencies of mercury vapor for different gas-liquid separators in flow injection cold vapour atomic absorption spectrometry with amalgam preconcentration. J. Anal. At. Spectrom. 8(1993): 585-590.
26. Konishi, T. and Takahashi, H. Direct determination of inorganic mercury in biological materials after alkali digestion and amalgamation. Analyst 108(1983): 827-834.
27. Talmi, Y. The rapid sub-picogram determination of volatile organomercury compounds by gas chromatography with a microwave emission spectrometric detector system. Anal. Chim. Acta. 74(1975): 107-117.
28. Cappon, J. and Smith, J. C. Gas-chromatographic determination of inorganic mercury and organomercurials in biological materials. Anal. Chem. 49(1977): 365-369.
29. Goolvard, L. and Smith, H. Determination of methylmercury in human blood. Analyst 105(1980): 726-729.

30. Talmi, Y. and Norvell, V.E. A rapid method for the determination of methylmercury chloride in water samples by gas chromatography with a microwave emission spectrometric detector. Anal. Chim. Acta. 85(1976): 203- 208.
31. Rubi, E., Lorenzo, R. A., Casais, M. C., and Cela, R. Quality control in the routine analysis of methylmercury in biological and environmental materials using gas chromatography with electron capture detection. Appl. Organomet. Chem. 8 (1994): 677-686.
32. Chiba, K., Yoshida, K., Tanabe, K., Haraguchi, H., and Fuwa, K. Determination of alkylmercury in seawater at the nanogram per liter level by gas chromatography/atmospheric pressure helium microwave-induced plasma emission spectrometry. Anal. Chem. 55(1983): 450-453.
33. Emteborg, H., Baxter, D. C., and Frech, W. Speciation of mercury in human whole blood by capillary gas chromatography with a microwave-induced plasma emission detector system following complexometric extraction and butylation. Analyst 117(1992): 567-663.
34. Bettmer, J., Bradter, M., Buscher, W., Erber, D., Rieping, D., and Cammann, K. GC-MIP-PED as an element-specific system for the determination of organomercury compounds. Appl. Organomet. Chem. 9(1995): 541-545.
35. Snell, J. P., Frech, W., and Thomassen, Y. Performance improvements in the determination of mercury species in natural gas condensate using an on-line amalgamation trap or solid-phase micro-extraction with capillary gas chromatography-microwave-induced plasma atomic emission spectrometry. Analyst 121(1996): 1055-1060.
36. Ceulemans, M. and Adams, F. C. Integrated sample preparation and speciation analysis for the simultaneous determination of methylated species of tin, lead, and mercury in water by purge-and-trap injection-capillary gas chromatography-atomic emission spectrometry. J. Anal. At. Spectrom. 11(1996): 201-206.

37. Lansens, P., Meuleman, C., Laino, C. C., and Baeyens, W. Comparative study of microwave-induced plasma atomic emission spectrometry and atomic fluorescence spectrometry as gas chromatographic detectors for the determination of methylmercury in biological samples. J. Anal. At. Spectrom. 7(1993): 45-51.
38. Kato, T., Uehiro, T., Yasuhara, A., and Morita, M. Determination of methylmercury species by capillary column gas chromatography with axially viewed inductively coupled plasma atomic emission spectrometric detection. J. Anal. At. Spectrom. 7(1992): 15-18.
39. Rapsomanikis, S. and Craig, P. J. Speciation of mercury and methylmercury compounds in aqueous samples by chromatography-atomic absorption spectrometry after ethylation with sodium tetraethylborate. Anal. Chim. Acta. 248(1991): 563-567.
40. Fischer, R., Rapsomanikis, S., and Andreae, M. O. Determination of methylmercury in fish samples using GC/AA and sodium tetraethylborate derivatization. Anal. Chem. 65(1993): 763-766.
41. Liang, L., Horvat, M., and Bloom, N. S. An improved speciation method for mercury by GC/CVAFS after aqueous phase ethylation and room temperature precollection. Talanta 41(1994): 371-379.
42. Alli, A., Jafe, R., and Jones, R. Analysis of organomercury compounds in sediments by capillary GC with atomic fluorescence detection. J. High Res. Chromatogr. 17(1994): 745-748.
43. Bowles, K. C. and Apte, S. C. Determination of methylmercury in natural water samples by steam distillation and gas chromatography-atomic fluorescence spectrometry. Anal. Chem. 70(1998): 395-399.
44. Moens, L., Smaele, T. D., Dams, R., Broeck, P. V. D., and Sandra, P. Sensitive, simultaneous determination of organomercury, -lead, and -tin compounds with headspace solid phase microextraction capillary gas chromatography combined with inductively coupled plasma mass spectrometry. Anal. Chem. 69(1997): 1604-1611.

45. Cai, Y. and Bayona, J. M. Determination of methylmercury in fish and river water samples using in situ sodium tetraethylborate derivatization following by solid-phase microextraction and gas chromatography-mass spectrometry. J.Chromatogr. A 696(1995): 113-122.
46. Lansens, P., Meuleman, C., and Baeyens, W. Evaluation of gas chromatographic columns for the determination of methylmercury in aqueous head space extracts from biological samples. J. Chromatogr. 586(1991): 329-340.
47. Lorenzo, R. A., Casais, C., Carro, A. M., and Cela, R. Evaluation of capillary columns used in the routine determination of methylmercury in biological and environmental materials. J. Chromatogr. 605(1992): 69-80.
48. Petersen, J. H. and Drabaek, I. Evaluation of CP Sil 8 film thickness for the capillary GC determination of methylmercury. Mikrochim. Acta 109(1992): 125-129.
49. Rapsomanikis, S. Derivatization by ethylation with sodium tetraethylborate for the speciation of metals and organometallics in environmental samples. Analyst 119 (1994): 1429-1439.
50. Holak, W. Determination of methylmercury in fish by high-performance liquid chromatography. Analyst 107(1982): 1457-1461.
51. Fujita, M. and Takabatake, E. Continuous flow reducing vessel in determination of mercuric compounds by liquid chromatography/cold vapor atomic absorption spectrometry. Anal. Chem. 55(1983): 454-457.
52. Krull, I. S., Bushee, D. S., Schleicher, R. G., and Smith, S. B. Jr. Determination of inorganic and organomercury compounds by high-performance liquid chromatography-inductively coupled plasma emission spectrometry with cold vapor generation. Analyst 111(1986): 345-349.
53. Langseth, W. Determination of organic and inorganic mercury compounds by reversed-phase high performance liquid chromatography after extraction of the compounds as their dithizonates. Anal. Chim. Acta 185(1986): 249-258.

54. Evans, O. and Mckee, G. D. Optimisation of high-performance liquid chromatographic separations with reductive amperometric electrochemical detection: speciation of inorganic and organomercury. Analyst 112(1987): 983-988.
55. Bushee, D. S. Speciation of mercury using liquid chromatography with detection by inductively coupled plasma mass spectrometry. Analyst 113(1988): 1167-1170.
56. Evans, O. and Mckee, G. D. Determination of mercury(II) and organomercury compounds by reversed-phase liquid chromatography with reductive electrochemical detection. Analyst 113(1988): 243-246.
57. Wu, J. C. G. Interfacing HPLC and cold-vapor AA with on-line preconcentration for mercury speciation. Spect. Lett. 24(1991): 681-697.
58. Shum, S. C. K., Pang, H., and Houk, R. S. Speciation of mercury and lead compounds by microbore column liquid chromatography-inductively coupled plasma mass spectrometry with direct injection nebulization. Anal. Chem. 64 (1992): 2444-2450.
59. Hempel, M., Hintelmann, H., and Wilken, R-D. Determination of organic mercury species in soils by high-performance liquid chromatography with ultraviolet detection. Analyst 117(1992): 669-672.
60. Sarzanini, C., Sacchero, G., Aceto, M., Abollino, O., and Mentasi, E. Simultaneous determination of methyl-, ethyl-, phenyl- and inorganic mercury by cold vapor atomic absorption spectrometry with on-line chromatographic separation. J. Chromatogr. 626(1992): 151-157.
61. Huang, C-W. and Jiang, S-J. Speciation of mercury by reversed-phase liquid chromatography with inductively coupled plasma mass spectrometric detection. J. Anal. At. Spectrom. 8(1993): 681-686.
62. Wang, Y-C. and Whang, C-W. High-performance liquid chromatography of inorganic mercury and organomercury with 2-mercaptobenzothiazole. J. Chromatogr. 628(1993): 133-137.
63. Hintelmann, H. and Wilken, R-D. The analysis of organic mercury compounds using liquid chromatography with on-line atomic fluorescence spectrometric detection. Appl. Organomet. Chem. 7(1993): 173-180.

64. Medina, I., Rudi, E., Mejuto, C., Casais, C., and Cela, R. Speciation of methylmercury in marine samples by HPLC with UV detection. Analisis 21 (1993): 215-218.
65. Bettmer, J., Cammann, K., and Robecke, M. Determination of organic ionic lead and mercury species with high-performance liquid chromatography using sulphur reagents. J. Chromatogr. A 654(1993): 177-182.
66. Falter, R. and Scholer, H. F. Interfacing high-performance liquid chromatography and cold-vapor atomic absorption spectrometry with on-line UV irradiation for the determination of organic mercury compounds. J. Chromatogr. A 675(1994) : 253-256.
67. Ho, Y-S. and Uden, P. C. Determination of inorganic Hg(II) and organic mercury compounds by ion-pair high-performance liquid chromatography. J. Chromatogr. A 688(1994): 107-116.
68. Aizpun, B., Fernandez, M. L., Blanco, E., and Sanz-medel, A. Speciation of inorganic mercury(II) and methylmercury by vesicle-mediated high-performance liquid chromatography coupled to cold vapor atomic absorption spectrometry. J. Anal. At. Spectrom. 9(1994): 1279-1284.
69. Fabbri, D. and Trombini, C. A novel derivatisation procedure for inorganic mercury(II) for HPLC analysis. Chromatographia 39(1994): 246-248.
70. Sarzanini, C., Sacchero, G., Aceto, M., Abollino, O., and Mentasi, E. Ion chromatographic separation and on-line cold vapor atomic absorption spectrometric determination of methylmercury, ethylmercury and inorganic mercury. Anal. Chim. Acta 284(1994): 661-667.
71. Schickling, C. and Broekaert, J. A. Determination of mercury species in gas condensates by on-line coupled high-performance liquid chromatography and cold-vapor atomic absorption spectrometry. Appl. Organomet. Chem. 9(1995): 29-36.
72. Wan, C-C., Chen, C-S., and Jiang, S-J. Determination of mercury compounds in water Samples by liquid chromatography-inductively coupled plasma mass spectrometry with an in situ nebulizer/vapor generator. J. Anal. At. Spectrom. 12(1997): 683-687.

73. Bidlingmeyer, B. A., Deming, S. N., Price, W. P., Jr., Sachok, B., and Petrussek, M. Retention mechanism for reversed-phase ion-pair liquid chromatography. J.Chromatogr. 186(1979): 419-434.
74. Melander, W. R., Kalghatgi, K., and Horvath, C. Formation of ion pairs under conditions employed in reversed-phase chromatography. J. Chromatogr. 201(1980): 201-209.
75. Foley, J. P. and May, W. E. Optimization of secondary chemical equilibria in liquid chromatography: theory and verification. Anal. Chem. 59(1987): 102-109.
76. Timerbaev, A. R., Tsoi, I. G., and Petrukhin, O. M. Mathematical simulation of complex chromatographic systems: a simulation model of reversed-phase liquid chromatography of metal chelates. Anal. Chim. Acta. 269(1992): 229-238.
77. Timerbaev, A. R. and Bonn, G. K. Complexation ion chromatography—an overview of developments and trends in trace metal analysis. J. Chromatogr. 640(1993): 195-206.
78. Chen, J-G., Weber, S. G., Glavina, L. L., and Cantwell, F. F. Electrical double-layer models of ion-modified (ion-pair) reversed-phase liquid chromatography. J. Chromatogr. A. 656 (1993): 549-576.
79. Janos, P. Complex-forming equilibria in ion chromatography. J. Chromatogr. A. 699 (1995): 1-10.
80. Krull, I. S. Trace metal analysis and speciation.: Journal of chromatography library—volume 47, The Netherlands : Elsevier Science Publishers B. V., 1991
81. Fergusson, J. E. The heavy elements: chemistry, environmental impact and health effects, 2nd ed., Great Britain : Pergamon, 1991.
82. Vernet, J. P. Trace metals in the environment 1. Heavy metals in the environment, 2nd ed., The Netherlands : Elsevier Science Publisher B. V., 1994
83. Craig, P. J. Organometallic compounds in the environment. Harlow: Longmans, 1986.
84. Hutchison, T. C. and Meema, K. M. Lead, mercury, cadmium and arsenic in the environment. John Wiley & Sons Ltd., 1987
85. Newman, M. C. and McIntosh, A. W. Metal ecotoxicology concepts & applications. The United States of America : Lewis Publishers, Inc., 1991.

86. Robert, C. W., Melvin, J. A., and William, H. B. CRC handbook of chemistry and physics. 65th ed., Florida : CRC Press, Inc., 1984.
87. AOAC International. AOAC® Peer-verified methods program: manual on policies and procedures. The United States of America.



สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

APPENDIX

Table A-1: The chromatographic conditions of the study of effect of mobile phase pH adjusted with hydrochloric acid and sodium hydroxide solution

Parameter	Conditions
Analytical Column	Hypersil column, 250 x 4.6 mm. I.D., 5 μ m
Mobile Phase	The mixture methanol-water (30:70% v/v) containing 0.0050M TBABr and 0.0050 % v/v 2-mercaptoethanol adjusting mobile phase pH with 0.2M HCl and 0.2M NaOH.
Flow Rate	1.00 mL/min
Injection Volume	10 μ L
Detector	Photodiode Array Detector
Data Acquisition	Maximum plot (200–400 nm)

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

Table A-2: The results of the effect of mobile phase pH adjusted with hydrochloric acid and sodium hydroxide solution on the chromatographic parameters for the separation of mercury compounds. (Triplicate analyses)

pH	compound	t _R (min)	W (min)	k'	α*	R _s **
2.0	solvent	2.924±0.012	-	-	1.209	1.327
	Hg ²⁺	5.419±0.055	0.367±0.088	0.853		
	MeHg ⁺	5.938±0.068	0.416±0.007	1.031		
	PhHg ⁺	35.125±0.345	0.658±0.035	11.022		
3.0	solvent	2.921±0.004	-	-	1.118	1.081
	Hg ²⁺	5.226±0.022	0.472±0.051	0.789		
	MeHg ⁺	5.682±0.013	0.372±0.094	0.945		
	PhHg ⁺	31.092±0.154	0.833±0.165	9.644		
4.0	solvent	2.919±0.010	-	-	1.210	1.172
	Hg ²⁺	5.102±0.054	0.400±0.088	0.747		
	MeHg ⁺	5.558±0.045	0.378±0.111	0.904		
	PhHg ⁺	29.97±0.616	0.825±0.035	8.968		
5.0	solvent	2.929±0.004	-	-	1.170	0.940
	Hg ²⁺	5.251±0.035	0.417±0.045	0.793		
	MeHg ⁺	5.646±0.029	0.422±0.038	0.928		
	PhHg ⁺	30.841±0.271	0.711±0.150	9.529		
6.0	solvent	2.921±0.008	-	-	1.146	0.940
	Hg ²⁺	5.277±0.016	0.383±0.029	0.806		
	MeHg ⁺	5.621±0.019	0.350±0.028	0.924		
	PhHg ⁺	29.877±0.969	0.784±0.071	9.228		
7.0	solvent	3.072±0.258	-	-	1.117	0.672
	Hg ²⁺	5.233±0.075	0.317±0.058	0.703		
	MeHg ⁺	5.483±0.064	0.428±0.009	0.785		
	PhHg ⁺	27.355±0.024	1.074±0.129	7.905		
8.1	solvent	2.927±0.045	-	-	1.178	0.964
	Hg ²⁺	5.314±0.052	0.467±0.100	0.815		
	MeHg ⁺	5.737±0.034	0.411±0.010	0.960		
	PhHg ⁺	31.831±0.512	0.783±0.099	9.875		

* selectivity factor between inorganic and methylmercury

** peak resolution between inorganic and methylmercury

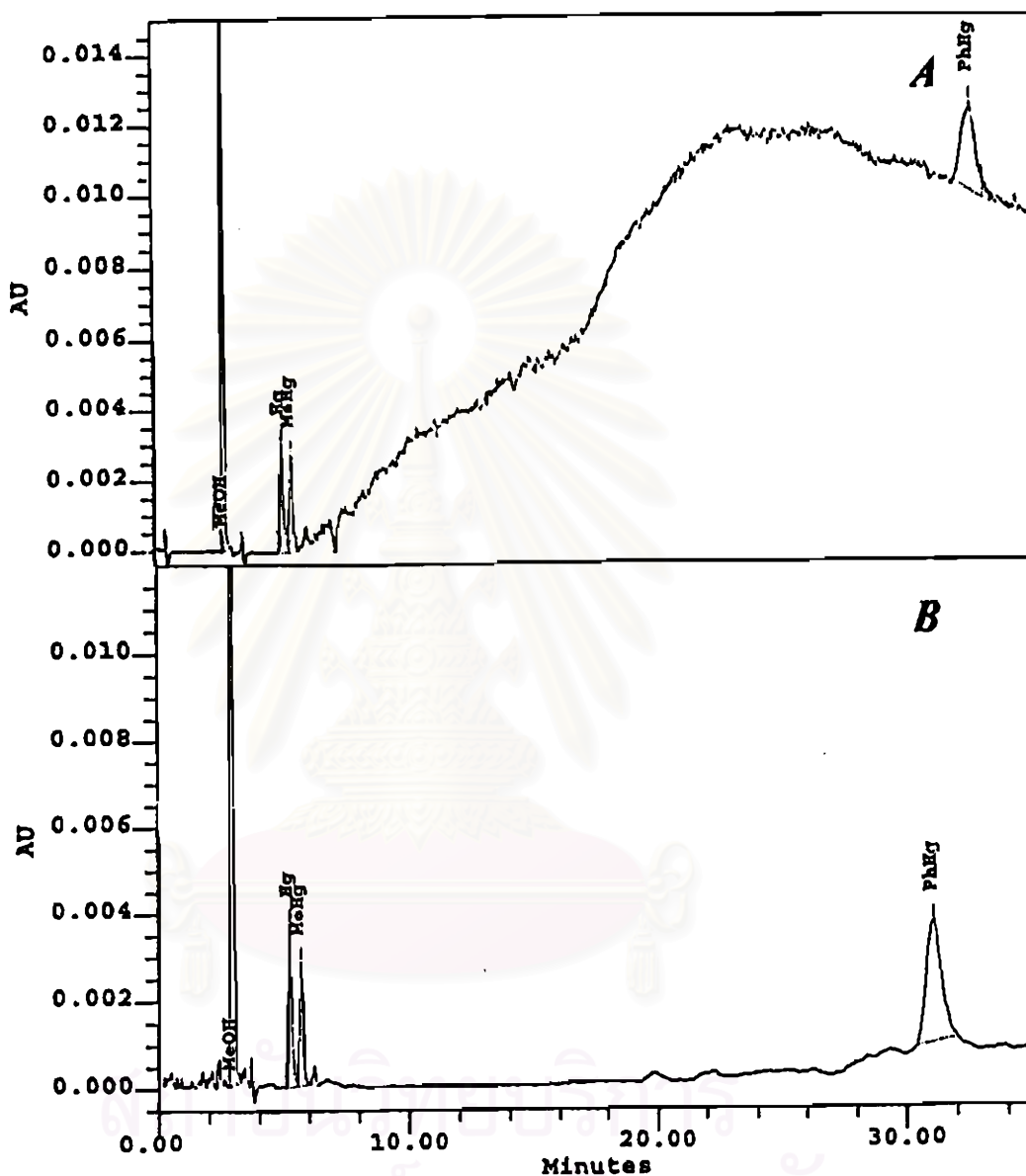


Figure A-1: The chromatograms of mercury compounds at variety of mobile phase pH adjusted with hydrochloric acid and sodium hydroxide solution (chromatographic conditions are shown in Table A-1).

- | | | | |
|-----------|-----------|-----------|-----------|
| A) pH 2.0 | B) pH 3.0 | C) pH 4.0 | D) pH 5.0 |
| E) pH 6.0 | F) pH 7.0 | G) pH 8.1 | |

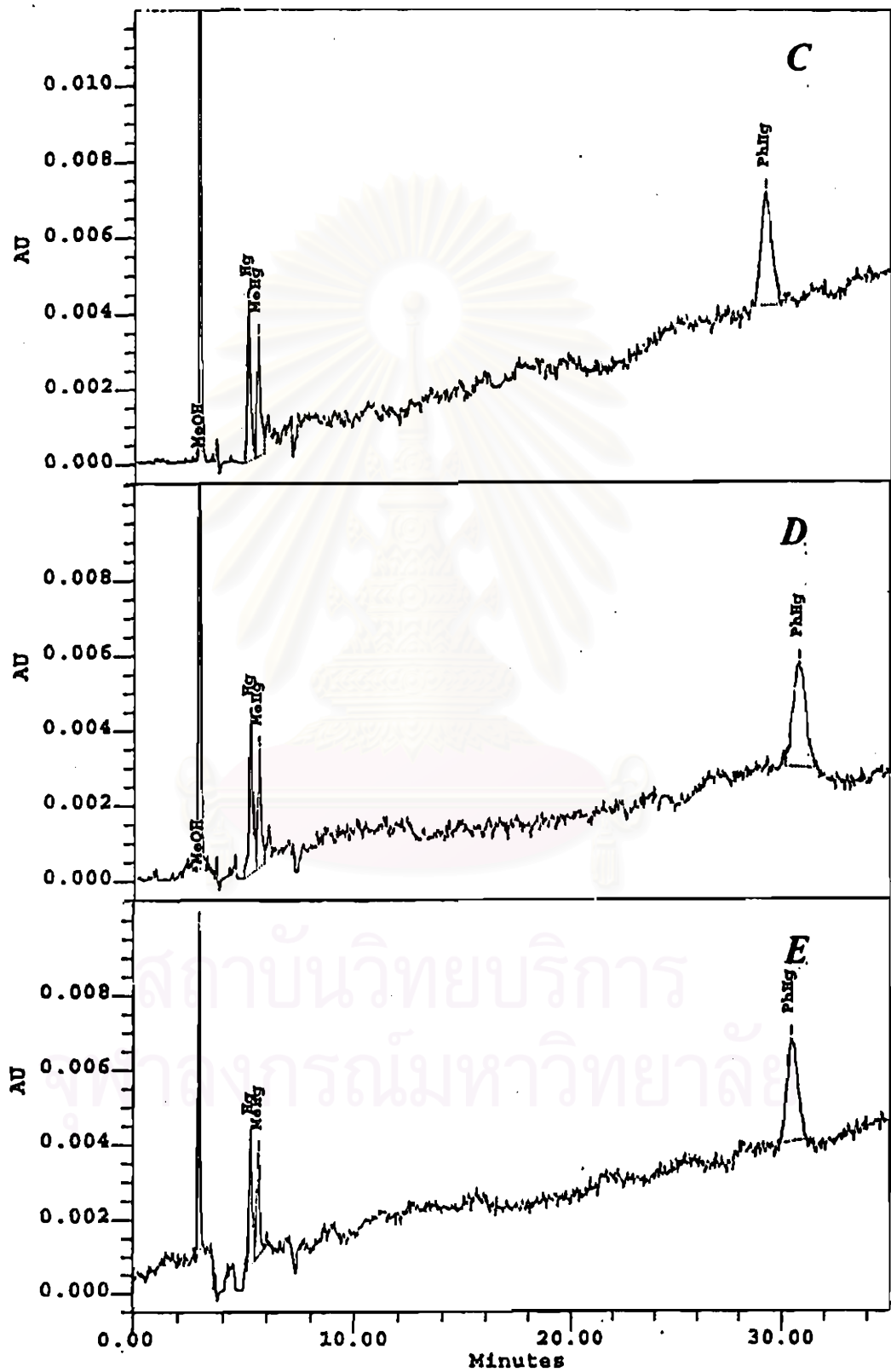


Figure A-1 (continue)

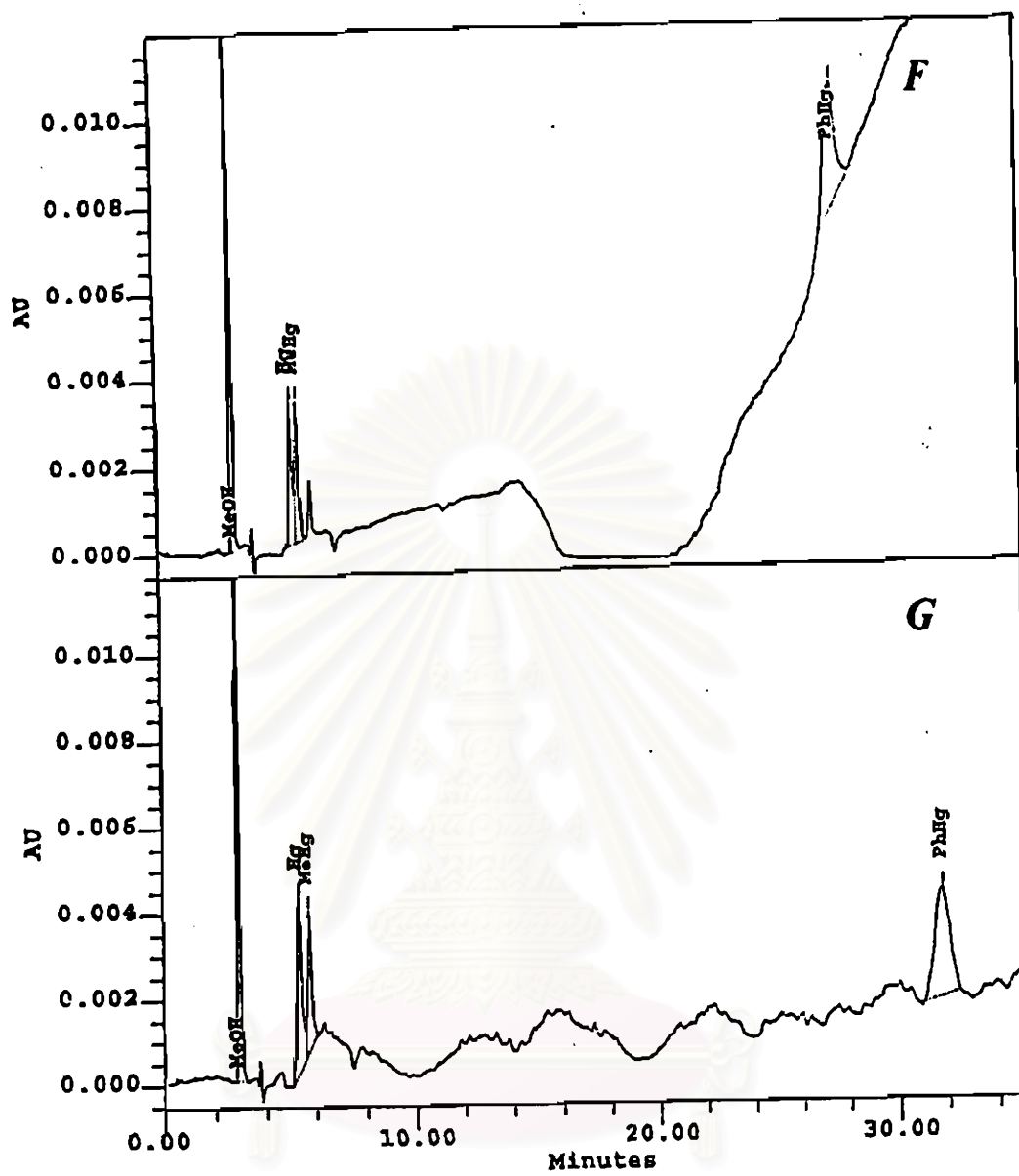


Figure A-1 (continue)

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

Table A-3: The chromatographic conditions of the study of effect of mobile phase pH adjusted with acetic acid-acetate buffer solution.

Parameter	Conditions
Analytical Column	Hypersil column, 250 x 4.6 mm. I.D., 5 μ m
Mobile Phase	The mixture methanol-water (30:70% v/v) buffered with 1.00×10^{-3} M AcONa-AcOH containing 0.0050M TBABr and 0.0050 % v/v 2-mercaptoethanol.
Flow Rate	1.00 mL/min
Injection Volume	10 μ L
Detector	Photodiode Array Detector
Data Acquisition	Maximum plot (200-400 nm)

Table A-4: The results of the effect of mobile phase pH adjusted with acetic acid-acetate buffer solution on the chromatographic parameters for the separation of mercury compounds. (Triplicate analyses)

pH	compound	t_r (min)	W (min)	k'	α^*	R_s^{**}
3.00	solvent	2.922 \pm 0.007	-	-	1.202	1.254
	Hg ²⁺	5.221 \pm 0.019	0.395 \pm 0.092	0.742		
	MeHg ⁺	5.670 \pm 0.009	0.322 \pm 0.092	0.892		
	PhHg ⁺	31.228 \pm 0.237	1.750 \pm 0.424	9.440		
3.50	solvent	2.932 \pm 0.005	-	-	1.177	0.836
	Hg ²⁺	5.448 \pm 0.025	0.578 \pm 0.130	0.858		
	MeHg ⁺	5.893 \pm 0.006	0.487 \pm 0.178	1.010		
	PhHg ⁺	34.120 \pm 0.666	0.850 \pm 0.117	10.637		
4.00	solvent	2.925 \pm 0.004	-	-	1.191	1.182
	Hg ²⁺	5.363 \pm 0.012	0.424 \pm 0.012	0.834		
	MeHg ⁺	5.830 \pm 0.026	0.367 \pm 0.076	0.993		
	PhHg ⁺	32.436 \pm 1.596	0.675 \pm 0.082	10.058		
5.00	solvent	2.924 \pm 0.002	-	-	1.178	1.060
	Hg ²⁺	5.391 \pm 0.030	0.372 \pm 0.054	0.844		
	MeHg ⁺	5.830 \pm 0.012	0.456 \pm 0.079	0.994		
	PhHg ⁺	33.702 \pm 0.380	0.866 \pm 0.189	10.526		

* selectivity factor between inorganic and methylmercury

** peak resolution between inorganic and methylmercury

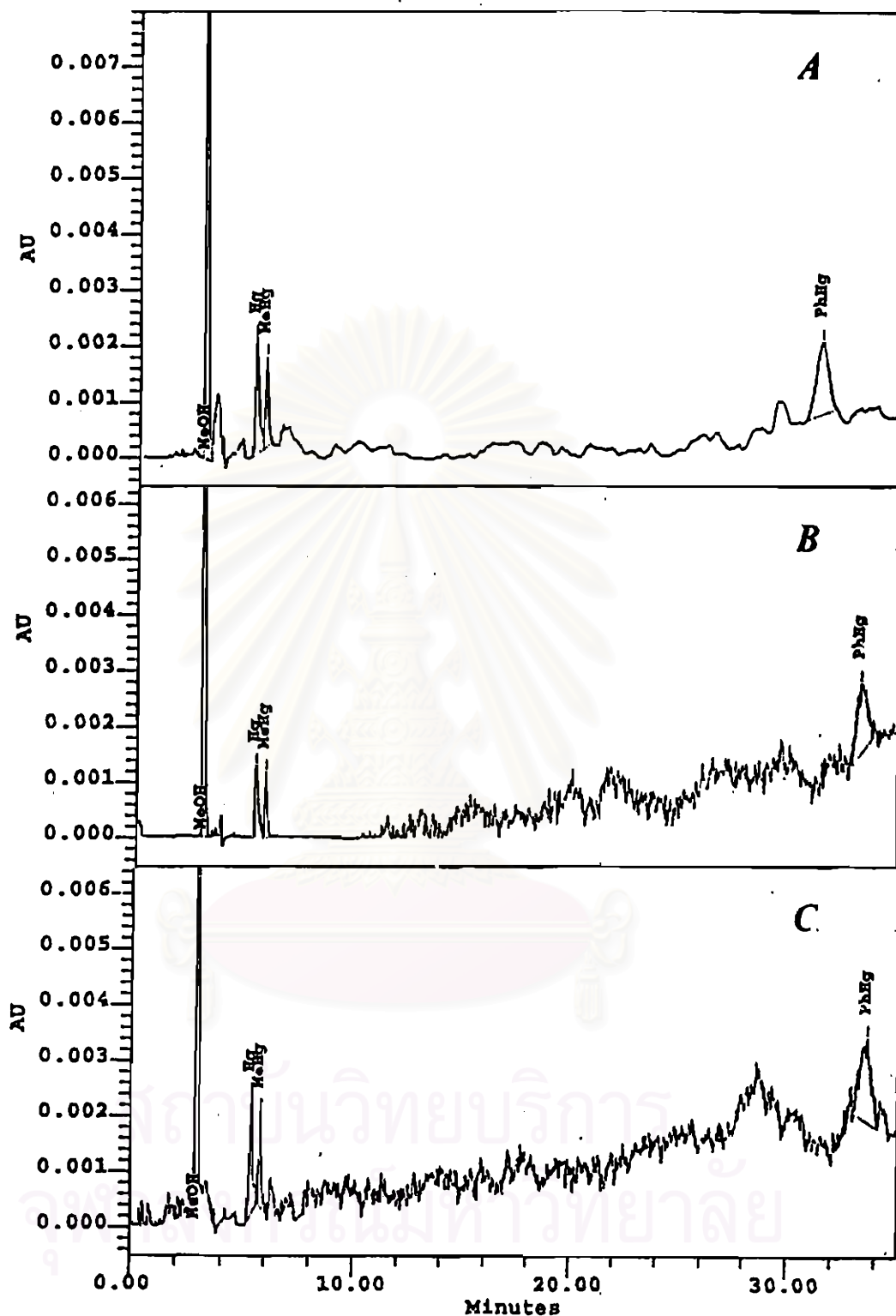


Figure A-2: The chromatograms of mercury compounds at variety of mobile phase pH with acetate-acetic acid buffered solution (chromatographic conditions are shown in Table A-3)

A) pH 3.00

B) pH 4.00

C) pH 5.00

Table A-5: The chromatographic conditions of the study of effect of 2-mercaptoethanol concentration

Parameter	Conditions
Analytical Column	Hypersil column, 250 x 4.6 mm. I.D., 5 μ m
Mobile Phase	The mixture methanol-water (30:70% v/v) buffered with 1.00×10^{-3} M AcONa-AcOH pH 3.00 containing 0.0050M TBABr and 2-mercaptoethanol.
Flow Rate	1.00 mL/min
Injection Volume	10 μ L
Detector	Photodiode Array Detector
Data Acquisition	Maximum plot (200-400 nm)

Table A-6: The results of the effect of 2-mercaptoethanol concentration on the chromatographic for the separation of mercury compounds. (Triplicate analyses)

%mercapto ethanol	compound	t_R (min)	W (min)	k'	α^*	R_s^{**}
0.0010%	solvent	2.908±0.007	-	-	1.180	0.986
	Hg ²⁺	5.321±0.012	0.462±0.016	0.830		
	MeHg ⁺	5.754±0.003	0.417±0.034	0.979		
	PhHg ⁺	32.408±0.103	1.826±0.228	10.144		
0.0020%	solvent	2.909±0.008	-	-	1.165	1.007
	Hg ²⁺	5.393±0.008	0.417±0.017	0.854		
	MeHg ⁺	5.804±0.014	0.400±0.017	0.995		
	PhHg ⁺	33.704±0.202	1.744±0.289	10.586		
0.0040%	solvent	2.909±0.003	-	-	1.192	1.032
	Hg ²⁺	5.215±0.091	0.483±0.076	0.793		
	MeHg ⁺	5.659±0.066	0.378±0.019	0.945		
	PhHg ⁺	30.924±0.023	1.108±0.106	9.630		
0.0050%	solvent	2.913±0.002	-	-	1.188	1.048
	Hg ²⁺	5.258±0.008	0.456±0.034	0.805		
	MeHg ⁺	5.697±0.002	0.383±0.034	0.956		
	PhHg ⁺	31.658±0.129	1.472±0.298	9.868		
0.0060%	solvent	2.907±0.007	-	-	1.187	1.009
	Hg ²⁺	5.201±0.099	0.450±0.047	0.789		
	MeHg ⁺	5.630±0.670	0.400±0.117	0.937		
	PhHg ⁺	30.404±0.126	1.300±0.322	9.459		
0.0075%	solvent	2.908±0.014	-	-	1.170	1.114
	Hg ²⁺	5.319±0.121	0.350±0.104	0.829		
	MeHg ⁺	5.730±0.068	0.590±0.025	0.970		
	PhHg ⁺	31.985±2.718	1.175±0.154	9.998		

* selectivity factor between inorganic and methylmercury

** peak resolution between inorganic and methylmercury

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

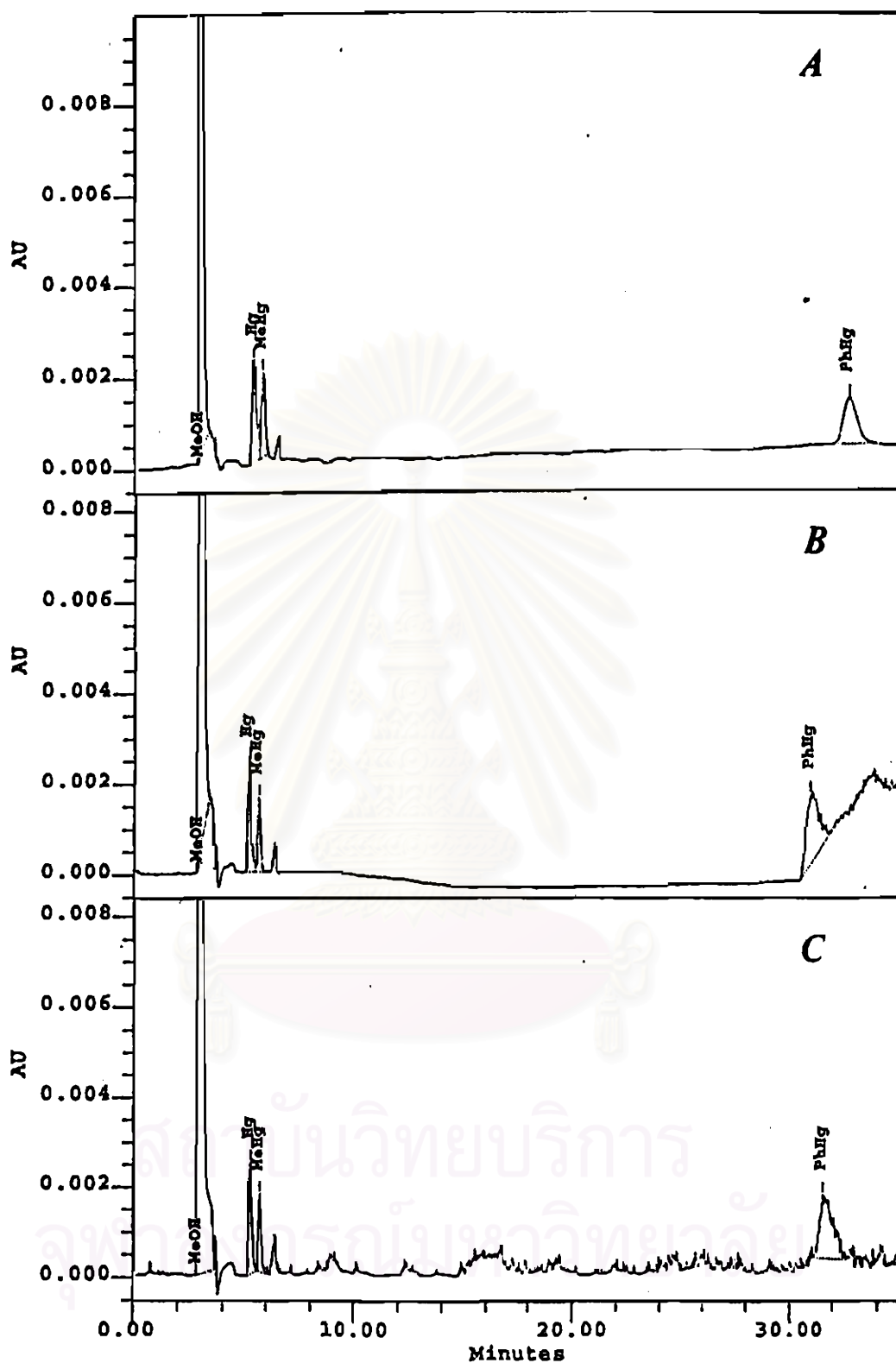


Figure A-3: The chromatograms of mercury compounds at variety of 2-mercaptoethanol concentration (chromatographic conditions are shown in Table A-5)

A) 0.0010% B) 0.0040% C) 0.0050% D) 0.0060%

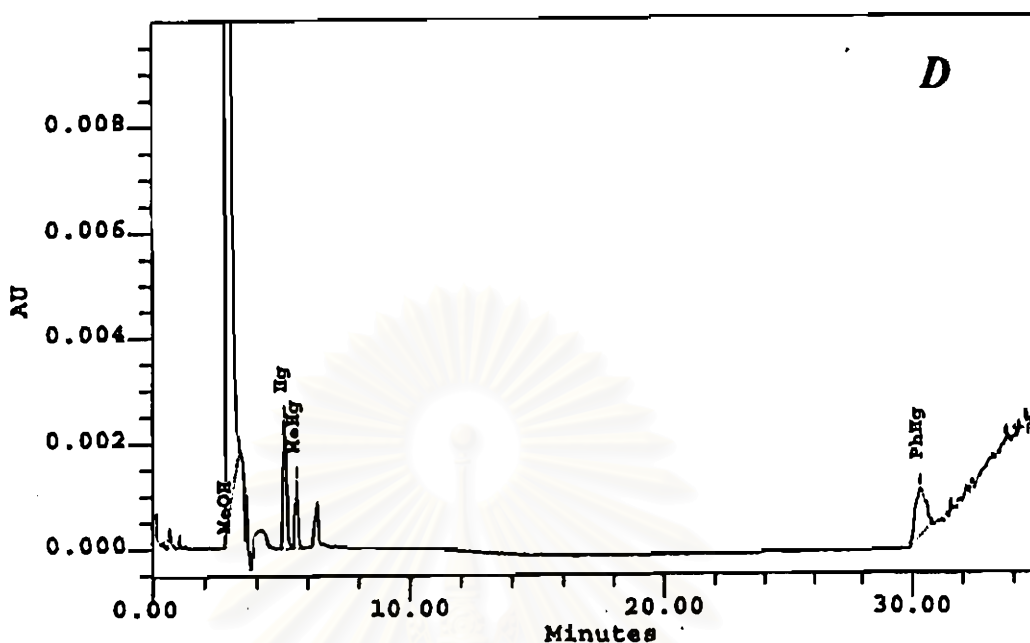


Figure A-3 (continue)

Table A-7: The chromatographic conditions of the study of effect of tetrabutylammonium bromide concentration

Parameter	Conditions
Analytical Column	Hypersil column, 250 x 4.6 mm. I.D., 5 μ m
Mobile Phase	The mixture methanol-water (30:70% v/v) buffered with 1.00×10^{-3} M AcONa-AcOH pH 3.00 containing 0.0050 % v/v 2-mercaptoethanol and TBABr.
Flow Rate	1.00 mL/min
Injection Volume	10 μ L
Detector	Photodiode Array Detector
Data Acquisition	Maximum plot (200-400 nm)

Table A-8 : The results of the effect of tetrabutylammonium bromide concentration on the chromatographic parameters for the separation of mercury compounds. (Triplicate analyses)

TBABr concentration	compound	t_R (min)	W (min)	k'	α^*	R_s^{**}
0.0000M	solvent	2.909±0.004	-	-	1.269	1.363
	Hg ²⁺	5.115±0.129	0.428±0.098	0.758		
	MeHg ⁺	5.708±0.121	0.442±0.083	0.962		
	PhHg ⁺	31.775±0.116	1.729±0.136	9.923		
0.0010M	solvent	2.914±0.004	-	-	1.220	1.073
	Hg ²⁺	5.196±0.004	0.525±0.011	0.783		
	MeHg ⁺	5.696±0.005	0.408±0.083	0.955		
	PhHg ⁺	30.821±0.310	1.283±0.235	9.577		
0.0025M	solvent	2.913±0.004	-	-	1.213	1.154
	Hg ²⁺	5.225±0.037	0.517±0.050	0.794		
	MeHg ⁺	5.719±0.042	0.399±0.038	0.963		
	PhHg ⁺	31.480±0.108	1.055±0.233	9.807		
0.0050M	solvent	2.913±0.002	-	-	1.188	1.048
	Hg ²⁺	5.258±0.008	0.456±0.034	0.805		
	MeHg ⁺	5.697±0.002	0.383±0.034	0.956		
	PhHg ⁺	31.658±0.129	1.472±0.298	9.868		
0.0075M	solvent	2.904±0.005	-	-	1.191	1.163
	Hg ²⁺	5.143±0.060	0.394±0.098	0.771		
	MeHg ⁺	5.571±0.044	0.329±0.025	0.918		
	PhHg ⁺	29.838±0.495	1.287±0.159	9.275		
0.0100M	solvent	2.901±0.001	-	-	1.149	0.890
	Hg ²⁺	5.290±0.085	0.405±0.009	0.823		
	MeHg ⁺	5.646±0.060	0.395±0.149	0.946		
	PhHg ⁺	31.557±0.237	1.433±0.189	9.878		
0.0150M	solvent	2.921±0.001	-	-	1.166	1.099
	Hg ²⁺	5.338±0.022	0.317±0.141	0.827		
	MeHg ⁺	5.738±0.025	0.411±0.114	0.964		
	PhHg ⁺	36.362±1.096	1.241±0.012	11.448		

* selectivity factor between inorganic and methylmercury

** peak resolution between inorganic and methylmercury

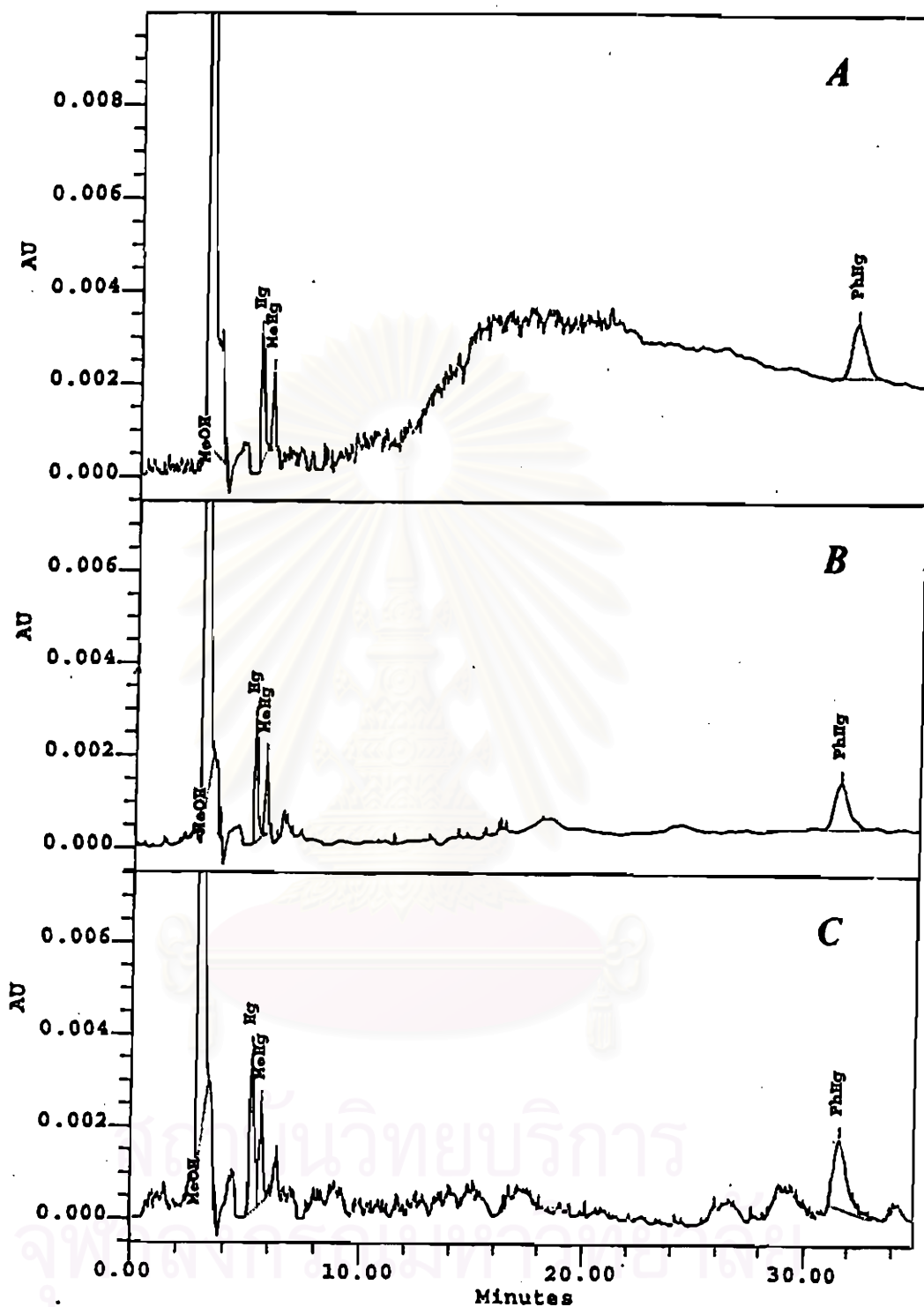


Figure A-4: The chromatograms of mercury compounds at variety of tetrabutylammonium bromide concentration (chromatographic conditions are shown in Table A-7)

A) 0.0000M

B) 0.0025M

C) 0.0050M

B) 0.0075M

E) 0.0100M

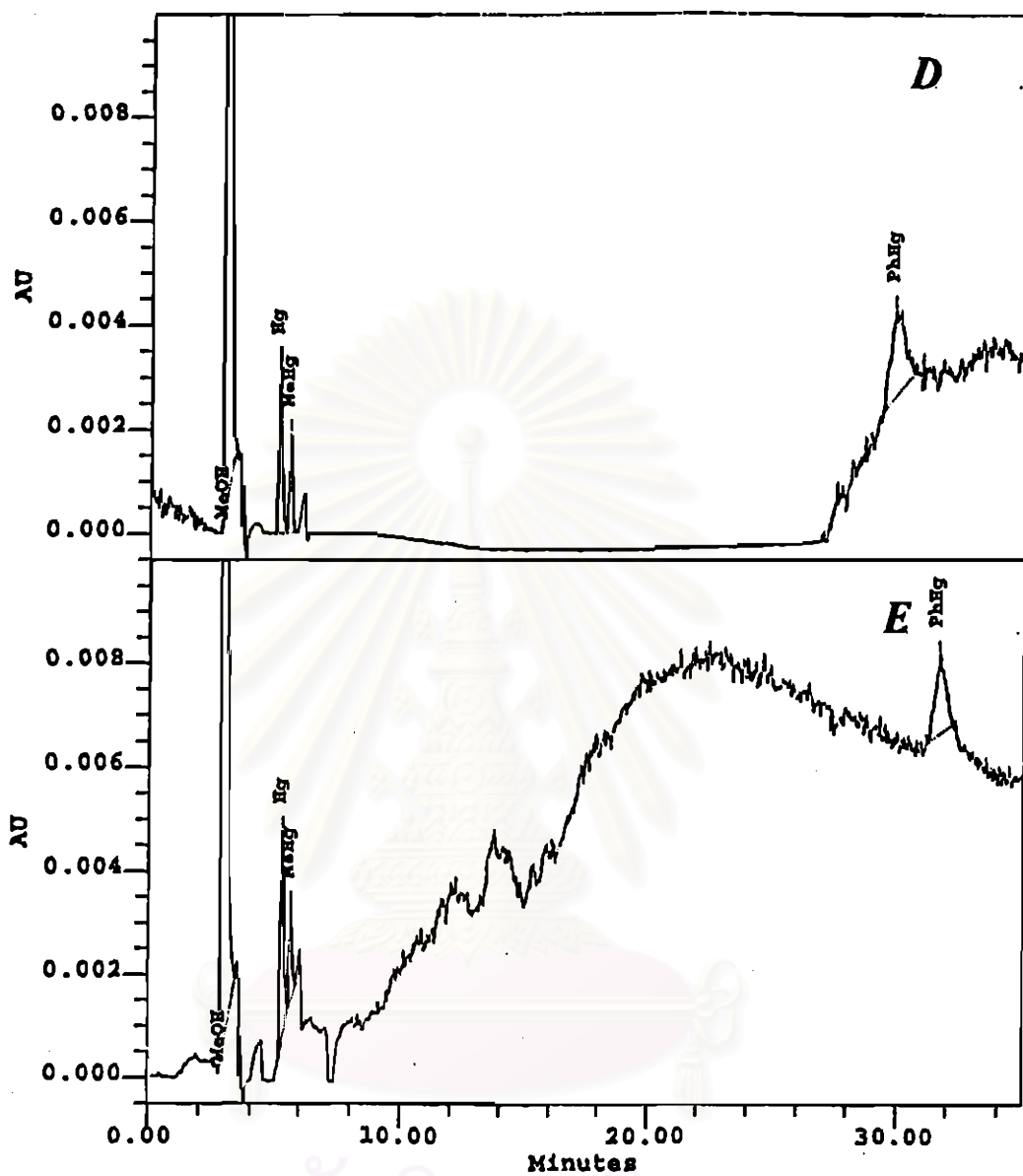


Figure A-4 (continue)

สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย

VITA

Miss Apinya Tunheng was born on September 5, 1974 at Chonburi Province. She received a Bachelor Degree of Science in Chemistry (second class honours) from Faculty of Science, Chulalongkorn University in 1996. After that, she started to study in Graduate School, Chulalongkorn University for the Master Degree of Science. She has been a graduate student, study in Analytical Chemistry in Chulalongkorn University.



สถาบันวิทยบริการ
จุฬาลงกรณ์มหาวิทยาลัย